

Hypothesis testing with a phylogeny

The challenge of accounting for phylogenetic non-independence

Guillaume Louvel

march 22, 2018

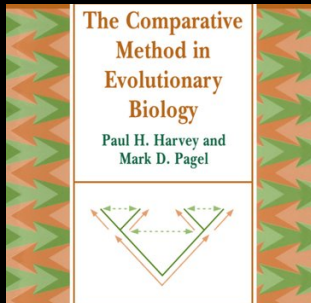
In the literature: the **Phylogenetic Comparative Method**.

László Zsolt Garamszegi
Editor



Modern Phylogenetic Comparative Methods and Their Application in Evolutionary Biology

Concepts and Practice



Syst. Biol. 67(1):14–21, 2018

Multivariate Phylogenetic Comparative Methods: Evaluations, Comparisons, and Recommendations

DEAN C. ADAMS^{1,2,*} AND MICHAEL L. COLLYER³

Test *evolutionary* hypotheses using observations from a set of species.

Concrete questions

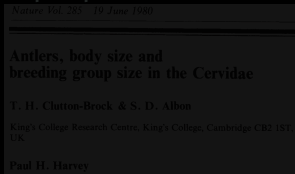
2 morphological traits

- Does one appear only when the other one exists?
- Are their value correlated? (magnitude and direction of change given the other)
- What is their *rate* of evolution?

What's the problem with a standard linear regression?

Disclaimer: strategy applicable for any *structured* data (spatial, etc).

example publications



Maddison (1990)
(discrete binary traits)
Social Brain Hypothesis
(continuous traits)
(DeCasien et al., 2017)

Concrete questions

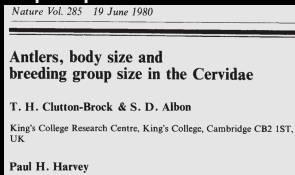
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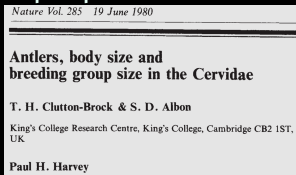
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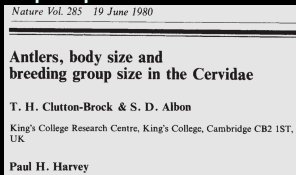
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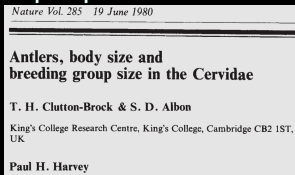
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Comparative data \neq controlled experiment

What explains morphological diversity? Phylogenetic inheritance, chance events, adaptation.

Phylogenetic inertia

- phylogenetic niche conservatism;
- phylogenetic time lag;
- different adaptive responses.

Losos 1994:

In a comparative analysis, a wrong phylogeny is better than no phylogeny at all.

The sister-clade method

2 binary traits

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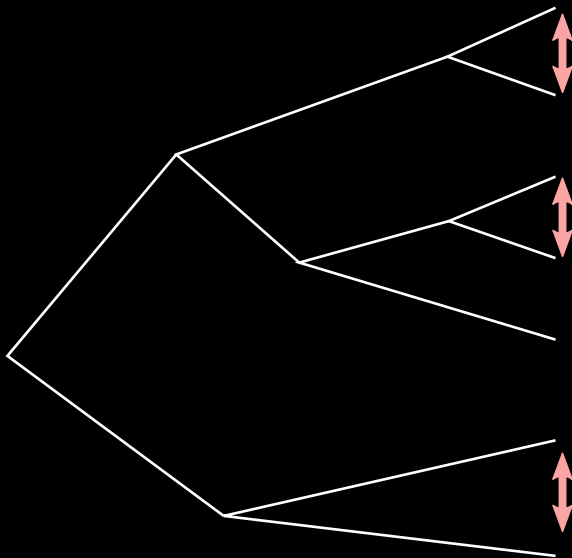
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Question: how would you normally do?

Fisher exact test / χ^2 test

example contingency table

rows: trait 1

columns: trait 2

cell content: number of *extant* species.

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Question: how would you normally do?

Fisher exact test / χ^2 test

example contingency table

rows: trait 1

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cell content: number of *extant* species.

- Ridley 1983: consider *branches*: count trait transitions.
- Maddison 1990: adapted to detect directionality of change

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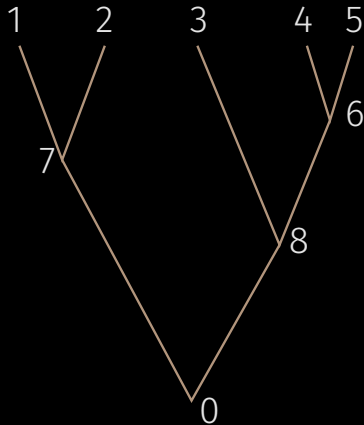
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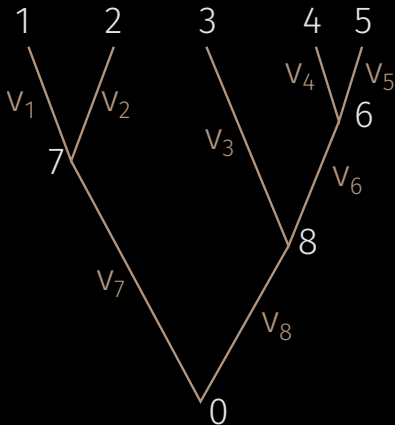
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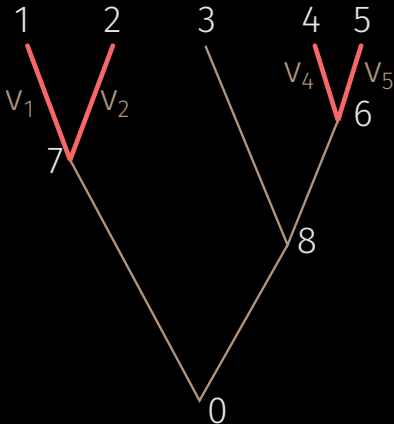
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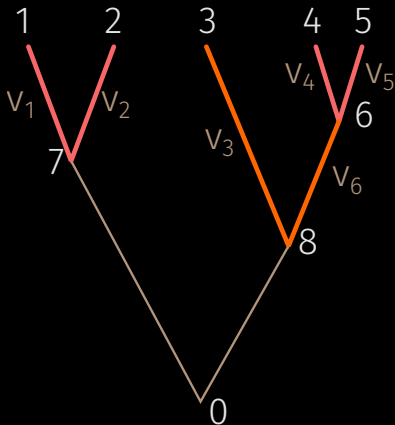
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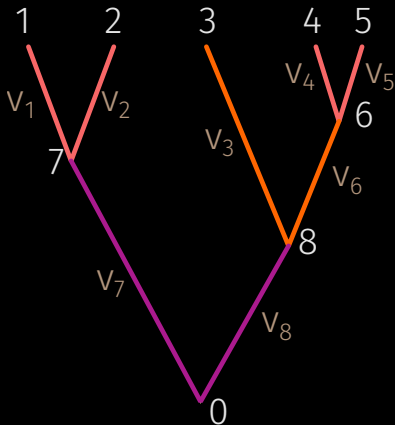
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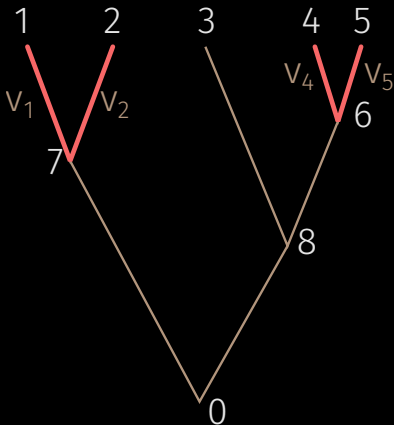
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Contrast 1-2

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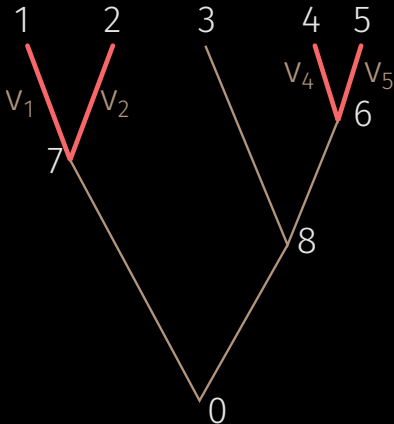
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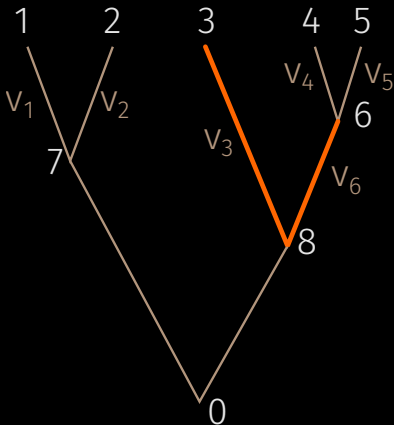
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Contrast 3-6

$$X_3 - X_6$$

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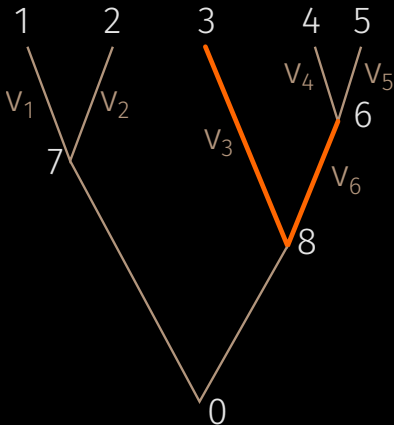
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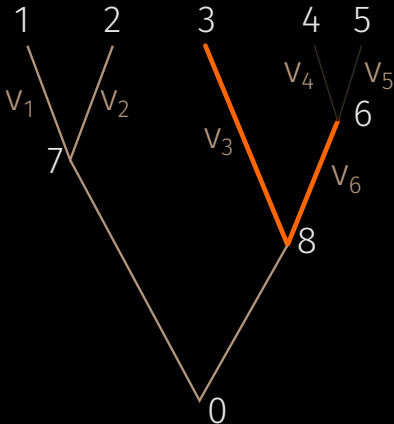
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$$X_3 - X_6$$

where $X_6 = \frac{X_4/v_4 + X_5/v_5}{1/v_4 + 1/v_5}$

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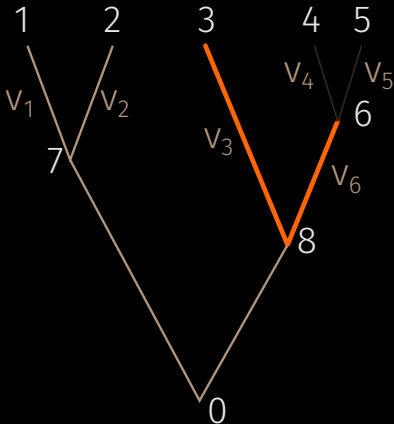
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$$\text{where } X_6 = \frac{X_4/v_4 + X_5/v_5}{1/v_4 + 1/v_5}$$

$$\text{Var}(X_3 - X_6) = v_3 + v'_6$$

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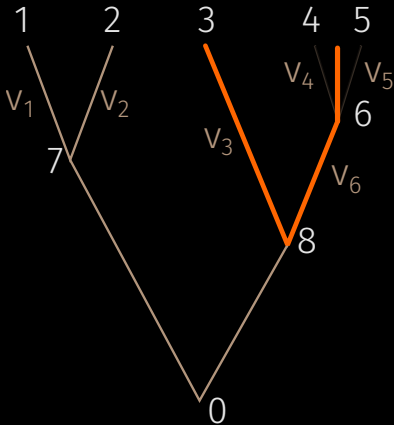
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$$\text{where } X_6 = \frac{X_4/v_4 + X_5/v_5}{1/v_4 + 1/v_5}$$

$$\text{Var}(X_3 - X_6) = v_3 + v_6 + \frac{v_4 v_5}{v_4 + v_5}$$

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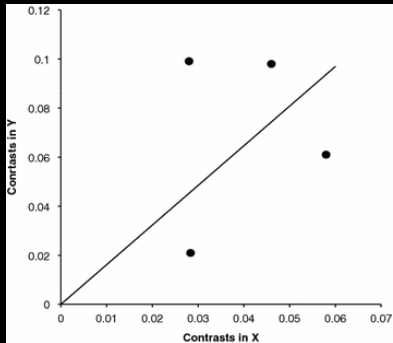
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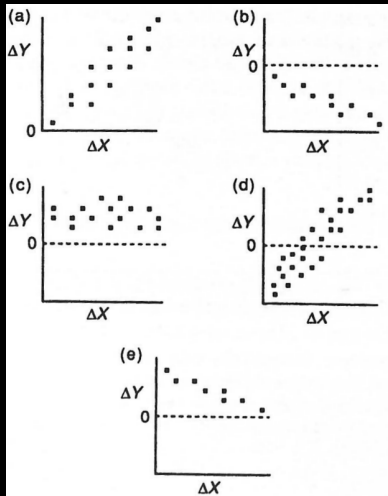
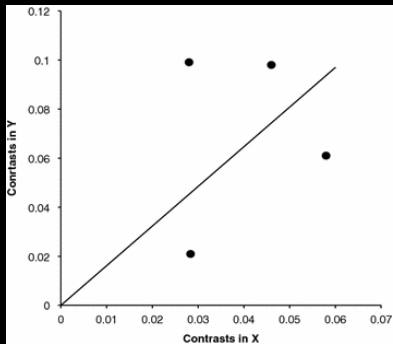
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From Harvey, Pagel, et al. 1991
p.160

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Example

```
library(ape)
# extract data with: geiger::treedata(df, tree)
pic.X <- pic(data$X, tree)
pic.Y <- pic(data$Y, tree)
cor.test(pic.X, pic.Y)
```

Reminder: the Linear Model

2D visualisation

$$y_i = a + bx_i + e_i$$

- i experimental unit;
- y_i response variable;
- x_i explanatory variable;
- a, b regression coefficients (model parameters);
- e_i residual error (variance not explained by the model).

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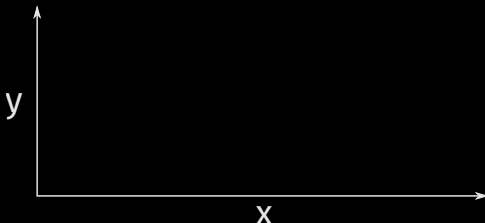
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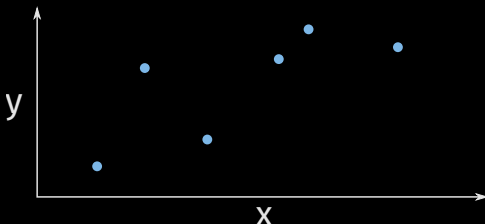
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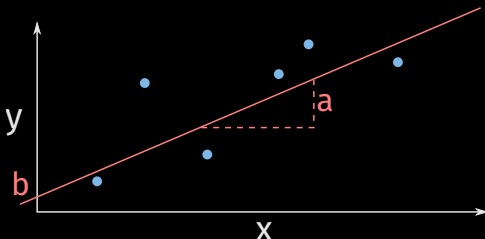


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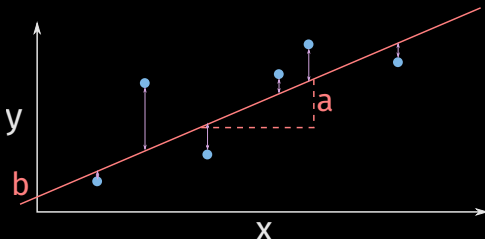


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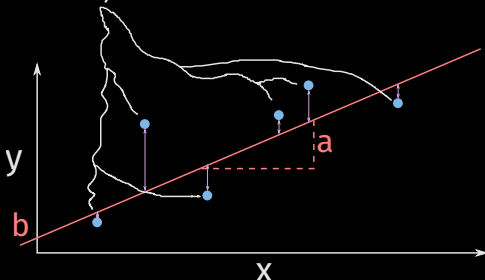


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Reminder: the Linear Model

With multiple explanatory variables:

$$y_i = \theta_0 + \theta_1 x_{i1} + \dots + \theta_p x_{ip} + \dots + e_i$$

$$Y = X\theta + E$$

$$\begin{bmatrix} \dots \\ y_i \\ \dots \end{bmatrix} = \text{exp units} \begin{bmatrix} \dots \\ X_{ip} \\ \dots \end{bmatrix} \begin{bmatrix} \dots \\ \theta_p \\ \dots \end{bmatrix} + \begin{bmatrix} \dots \\ e_i \\ \dots \end{bmatrix}$$

Linear Model Assumptions

- **linearity** : $\mathbb{E}(Y) = X\theta$
- e_i are:
 - *independent and identically distributed* (homoscedasticity).
 - normally distributed $\mathcal{N}(0, \sigma^2)$

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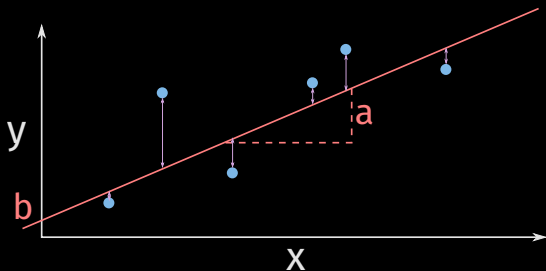
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Reminder: the Linear Model



Linear Model Assumptions

- **linearity** : $\mathbb{E}(Y) = X\theta$
- e_i are:
 - ▶ *independent and identically distributed* (homoscedasticity) .
 - ▶ normally distributed $\mathcal{N}(0, \sigma^2)$

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OLS vs GLS

PGLS

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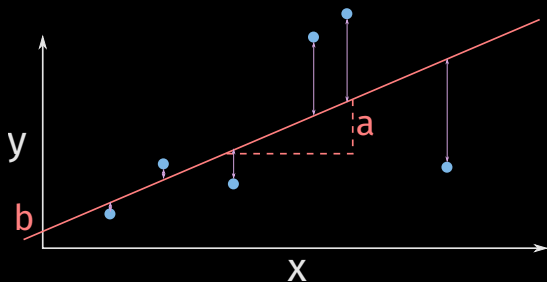
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Reminder: the Linear Model



General Linear Model Assumptions

- **linearity** : $\mathbb{E}(Y) = X\theta$
- e_i are:
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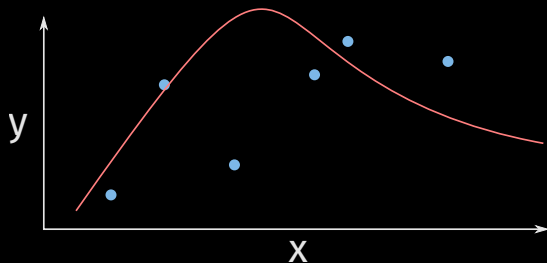
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Reminder: the Linear Model



Generalized Linear Model Assumptions

- **linearity + link function:** $\mathbb{E}(Y) = g(X\theta)$
- e_i are:
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Fitting a Linear model with Least Squares

Ordinary Least Squares

Minimizing the sum of squared errors:

$$\operatorname{argmin}_{a,b} S(a, b)$$

$$\begin{aligned} S(a, b) &= \sum_{i=1}^n e_i^2 \\ &= \sum_{i=1}^n (y_i - (a + bx_i))^2 \end{aligned}$$

Generalized Least Squares

Residues *covariate*:

so we use a covariance matrix $\operatorname{Cov}(E) = \Omega$

$$\hat{\theta} = (X^t \Omega^{-1} X)^{-1} X^t \Omega^{-1} Y$$

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Ordinary Least Squares

Minimizing the sum of squared errors:

$$\hat{\theta} = \operatorname{argmin}_{\theta} \|Y - X\theta\|^2$$

We assume:

$$\mathbb{E}(E) = 0 \text{ and } \operatorname{Cov}(E) = \sigma^2 I$$

There is a solution:

$$\hat{\theta} = (X^t X)^{-1} X^t Y$$

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Some terminology

General Linear Model: t-test, multiple regression, ANOVA, ANCOVA

⚠ Linearity: $y = \alpha + \beta x_1 + \gamma x_2 + e$

⚠ “Generalized”:

Generalized Linear Model Assumptions

- **linearity + link function:**
 $\mathbb{E}(Y) = g(X\theta)$
- e_j are:
 - *independent and identically distributed.*

Generalized Least Squares Assumptions

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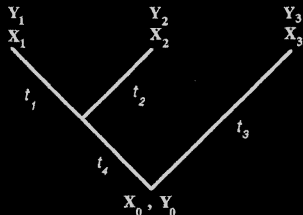
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Grafen 1989

Getting the covariance matrix from the phylogeny:



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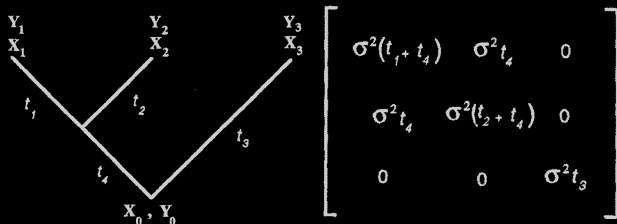
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Getting the covariance matrix from the phylogeny:



Phylogenetic Independent Constrasts VS Phylogenetic Generalized Least Squares

Independent Contrasts and PGLS Regression Estimators Are Equivalent FREE

Simon P. Blomberg , James G. Lefevre, Jessie A. Wells, Mary Waterhouse [Author Notes](#)

Systematic Biology, Volume 61, Issue 3, 1 May 2012, Pages 382–391,
<https://doi.org/10.1093/sysbio/syr118>

Published: 03 January 2012 [Article history](#) ▼

- PIC do not provide the intercept directly;
- Both can be generalized to other evolution processes (Ornstein-Uhlenbeck);
- PGLS easier with partially unresolved phylogenies.

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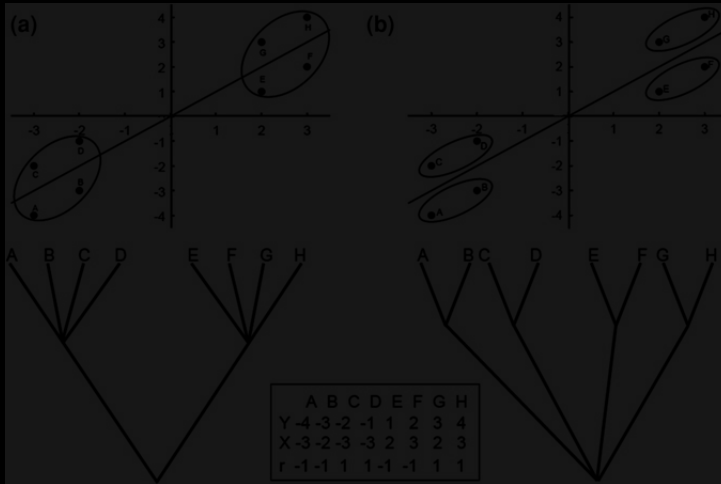
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Should you correct for phylogenetic structure?

Yes if there is *phylogenetic signal* in the *residuals*.

⚠ in residuals \neq in the response variable !



From Labra et al 2009

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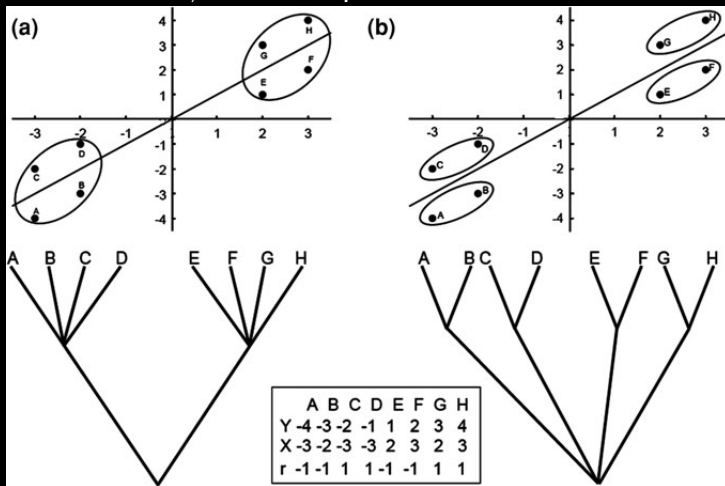
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Brief tutorial in R

Example from

<http://www.mpcm-evolution.org/practice>.

Some packages

```
library(ape) # tree handling
library(nlme) # regression modelling
# or
library(caper) # pglS() function
```

data

```
library(ade4); data(lizards)
tree <- read.tree(text = lizards$hprA)
dat <- lizards$traits[tree$tip.label, ] # sort data
      according to tree
plot(tree, main = "Phylogeny for 18 Lizard Species",
      direction = "up", srt = -90, label.offset = 1)
```

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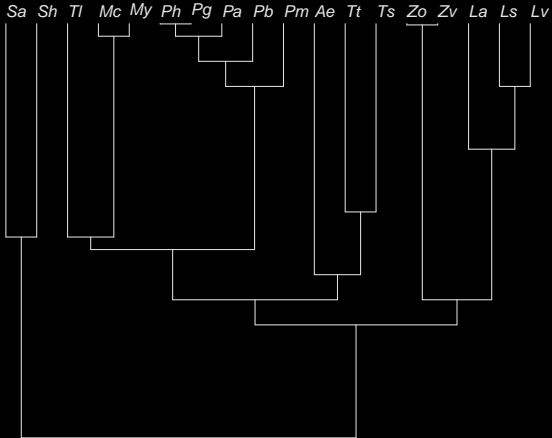
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	mean.L	matur.L	max.L	hatch.L	hatch.m	clutch.S	age.mat	clutch.F
Sa	69.2	58	82	27.8	0.572	6.0	13	1.5
Sh	48.4	42	56	22.9	0.310	3.2	5	2.0
Tl	168.4	132	190	42.8	2.235	16.9	19	1.0
Mc	66.1	56	72	25.0	0.441	7.2	11	1.5
My	70.1	60	81	26.6	0.550	5.4	10	1.0
Ph	49.6	39	57	23.8	0.310	2.1	8	2.0

```
fit <- gls(matur.L ~ age.mat, correlation=corBrownian
(tree), data=dat)
```

```
# Custom correlation matrix:
myamat <- vcv(tree, corr=TRUE) # construct correlation
matrix
corSymm(myamat[lower.tri(mat)], fixed=TRUE)
```

	mean.L	matur.L	max.L	hatch.L	hatch.m	clutch.S	age.mat	clutch.F
Sa	69.2	58	82	27.8	0.572	6.0	13	1.5
Sh	48.4	42	56	22.9	0.310	3.2	5	2.0
Tl	168.4	132	190	42.8	2.235	16.9	19	1.0
Mc	66.1	56	72	25.0	0.441	7.2	11	1.5
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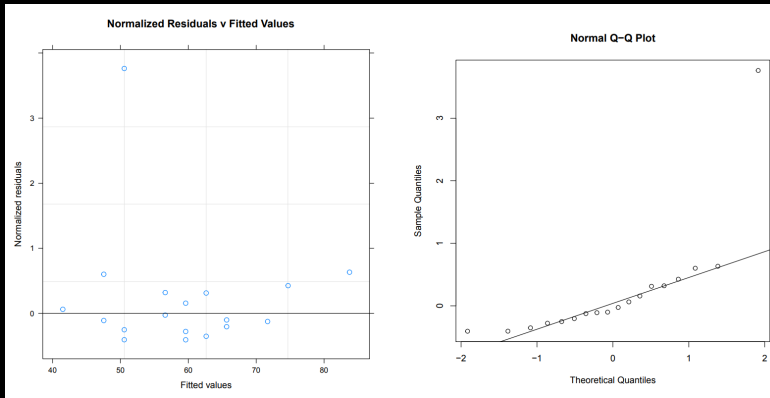
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```

plot(fit2 , resid(., type="n")~fitted(.), main="
  Normalized Residuals v Fitted Values", abline=c
  (0,0))
res <- resid(fit2 , type="n")
qqnorm(res)
qqline(res)

```



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phylogenetic signal

```
fitPagel <- gls(matur.L ~ age.mat, correlation=
  corPagel(value=0.8, phy=tree3), data=dat3)
intervals(fitPagel, which="var-cov")
## Approximate 95% confidence intervals
##
## Correlation structure:
## lower est. upper
## lambda 0.49 0.899 1.308
## attr(,"label")
## [1] "Correlation structure:"
##
## Residual standard error:
## lower est. upper
## 11.76 21.88 40.72
```

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```

fitPagelo <- gls(matur.L ~ age.mat, correlation =
  corPagel(value = 0, phy = tree3, fixed = TRUE),
  data = dat3) # independence
fitPagel1 <- gls(matur.L ~ age.mat, correlation =
  corPagel(value = 1, phy = tree3, fixed = TRUE),
  data = dat3) # Brownian motion

anova(fitPagel, fitPagelo)
## Model df AIC BIC logLik Test L.Ratio p-value
## fitPagel 1 4 140.2 143.0 -66.08
## fitPagelo 2 3 137.8 139.9 -65.91 1 vs 2 0.3439
0.5576

```

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References to get started

Felsenstein 1985 “Phylogenies and the Comparative Method”

Garamszegi 2014 *Modern Phylogenetic Comparative Methods and Their Application in Evolutionary Biology*

www.mpcm-evolution.org/practice

Harvey, Pagel, et al. 1991 *The comparative method in evolutionary biology*

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That's not all!

- phylogenetic signal (Münkemüller et al. 2012)
- phylogenetic ANOVA, ANCOVA, multivariate analysis...
- inferring causality
- The seven deadly sins of comparative analysis (Freckleton 2009)

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Ordinary and Generalized Least Squares

Least Squares Estimator:

$$\hat{\theta} = \operatorname{argmin}_{\theta} \|Y - X\theta\|^2$$

It verifies:

$$X^t X \hat{\theta} = X^t Y$$

So when $X^t X$ is invertible (*i.e.* the matrix is of full rank, there are more data points than explanatory variables, and explanatory variables are independent):

$$\hat{\theta} = (X^t X)^{-1} X^t Y$$

When there is covariance between residuals (Generalized Least Squares):

$$\hat{\theta} = (X^t \Omega^{-1} X)^{-1} \Omega^{-1} X^t Y$$

(corresponds to minimizing the squared Mahalanobis length of the residual vector)