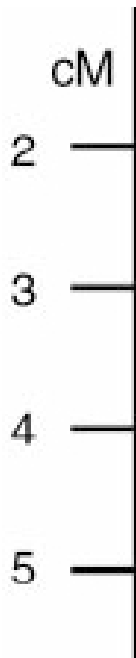


Mapping the genes underlying phenotypic changes of interest

**Virginie Courtier-Orgogozo
Institut Jacques Monod, Paris**

Tomato shape



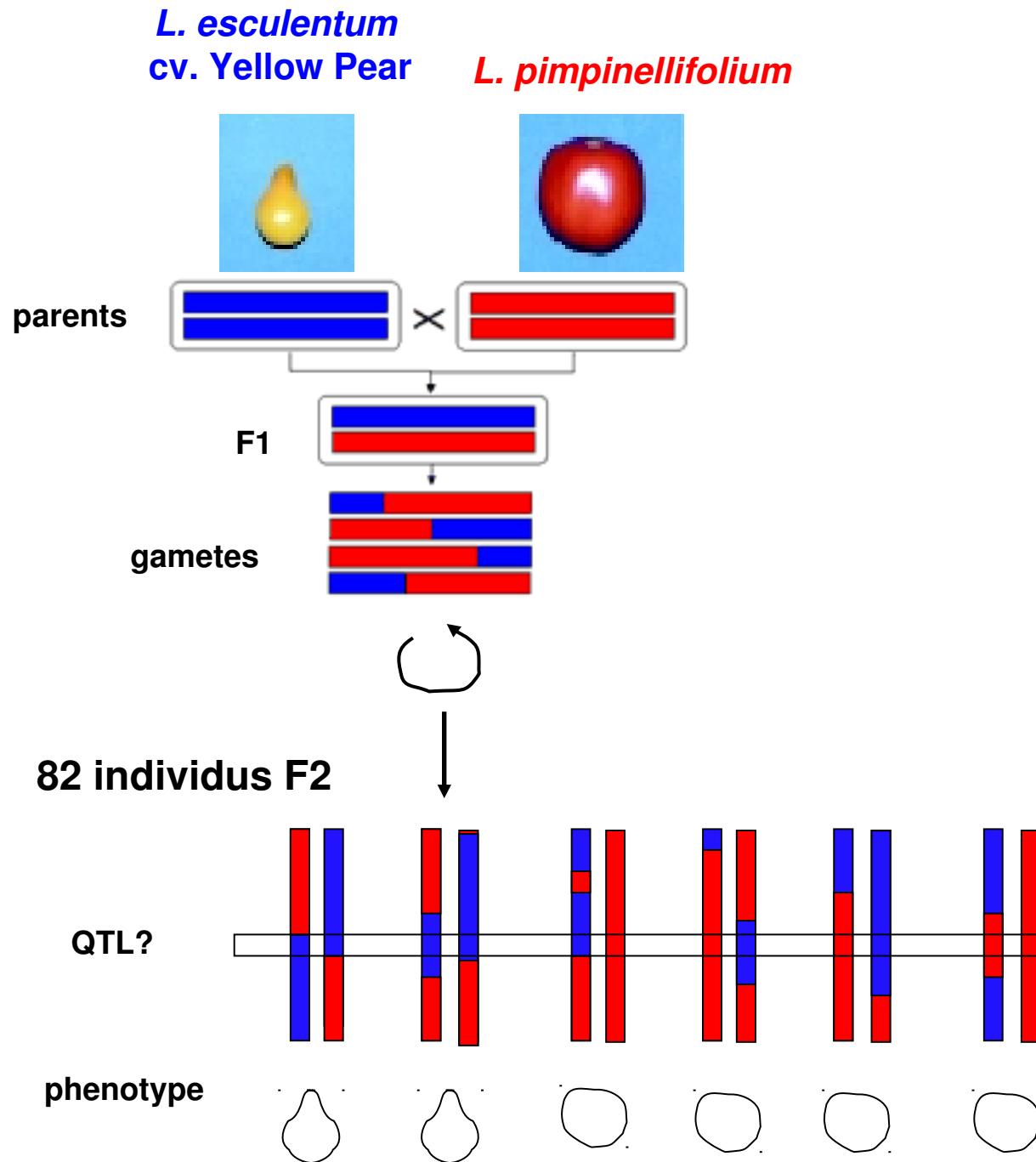
Lycopersicon esculentum



Lycopersicon esculentum cv. Yellow Pear

(Ku et al., 1999; Liu et al., 2002)

QTL mapping

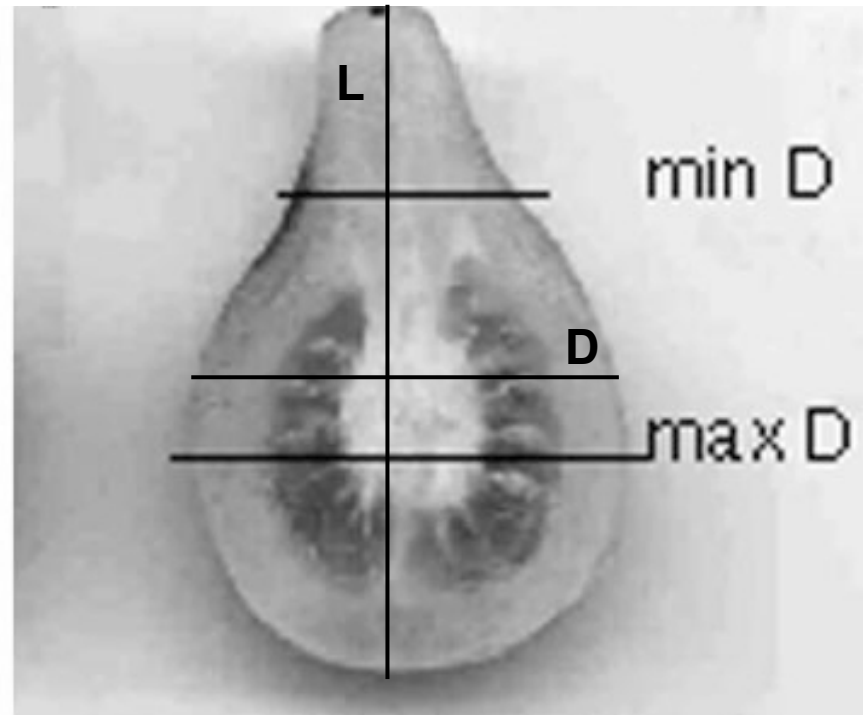
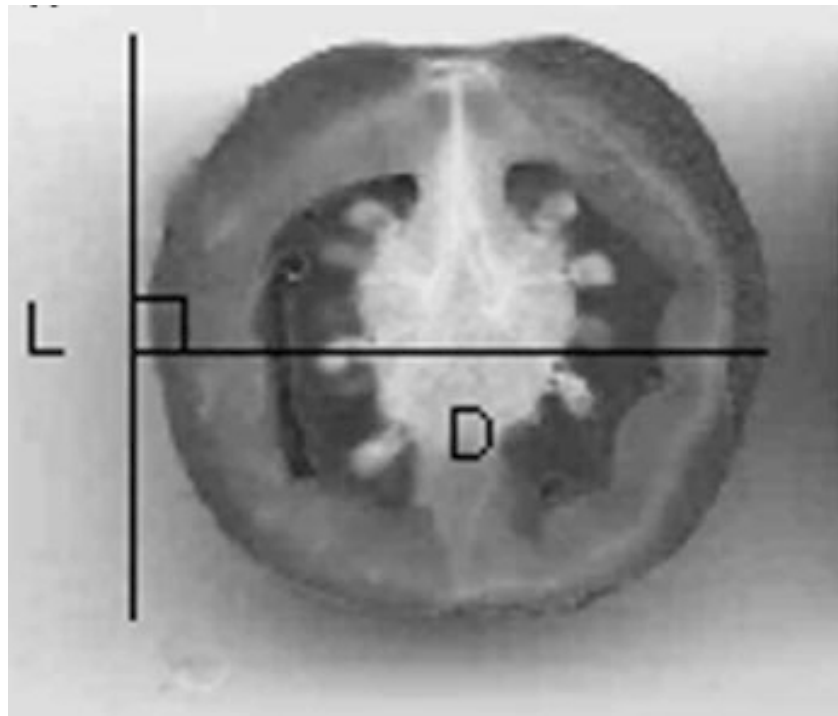


Quantitative measure of the phenotype

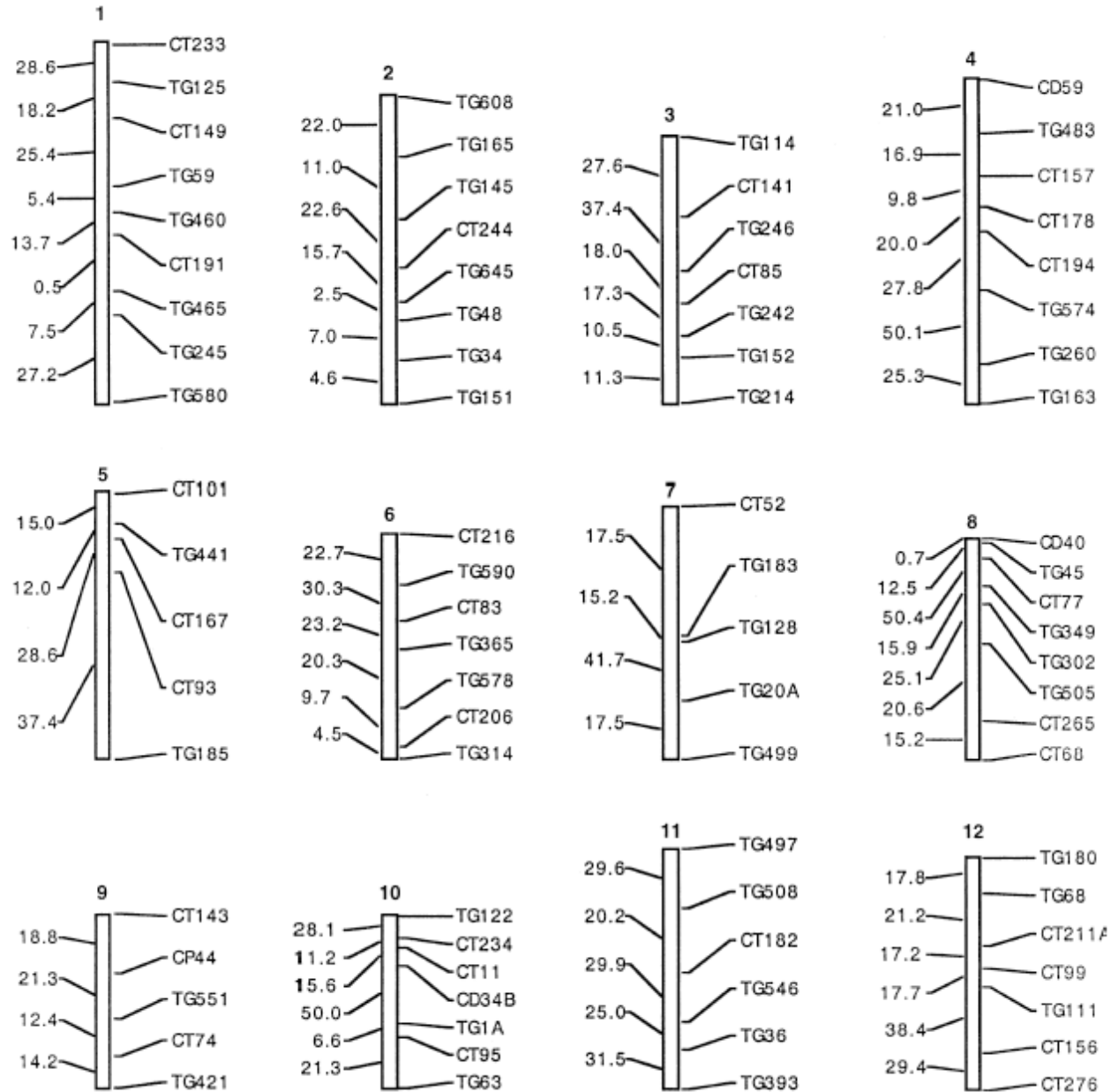
Measure of 2 indexes L/D and D_{min}/D_{max} for 10 fruits per plant

L/D : L = length, D = diameter at equator

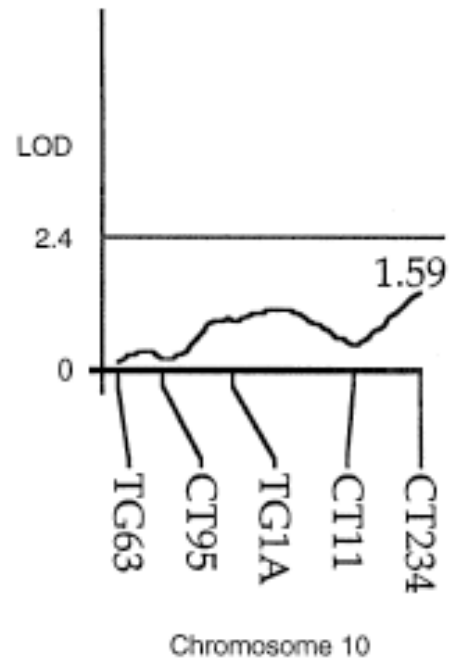
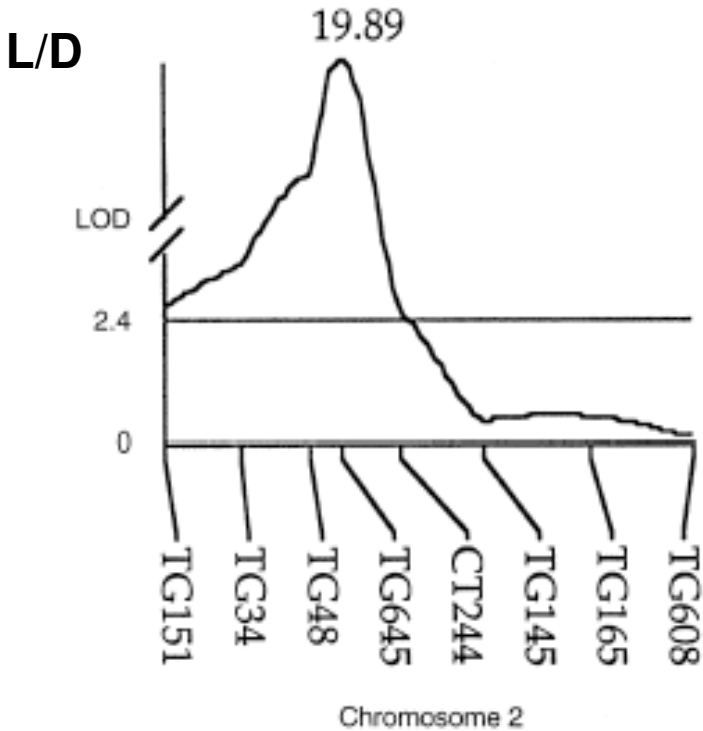
D_{min}/D_{max}



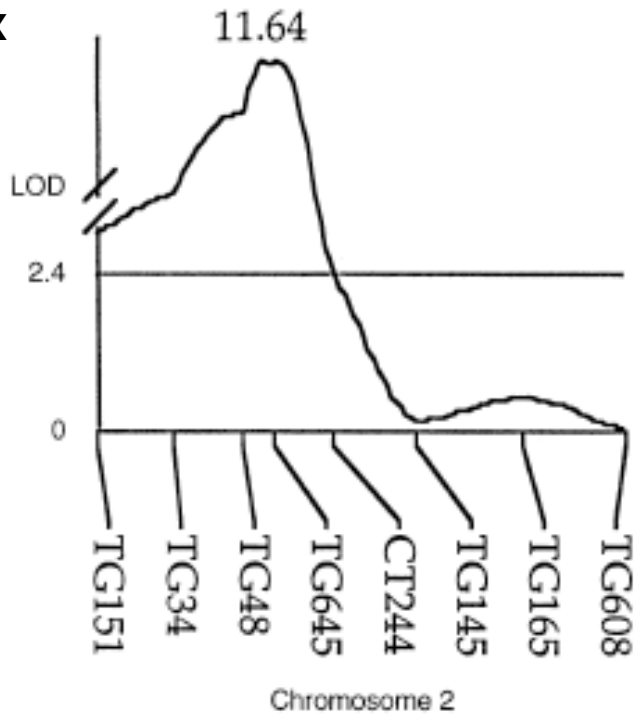
82 molecular markers on the 12 tomato chromosomes



One major locus near marker TG645



Dmin/Dmax



responsible for 67% of L/D variance

allele YP = recessive

BAC library (Bacterial Artificial Chromosomes)

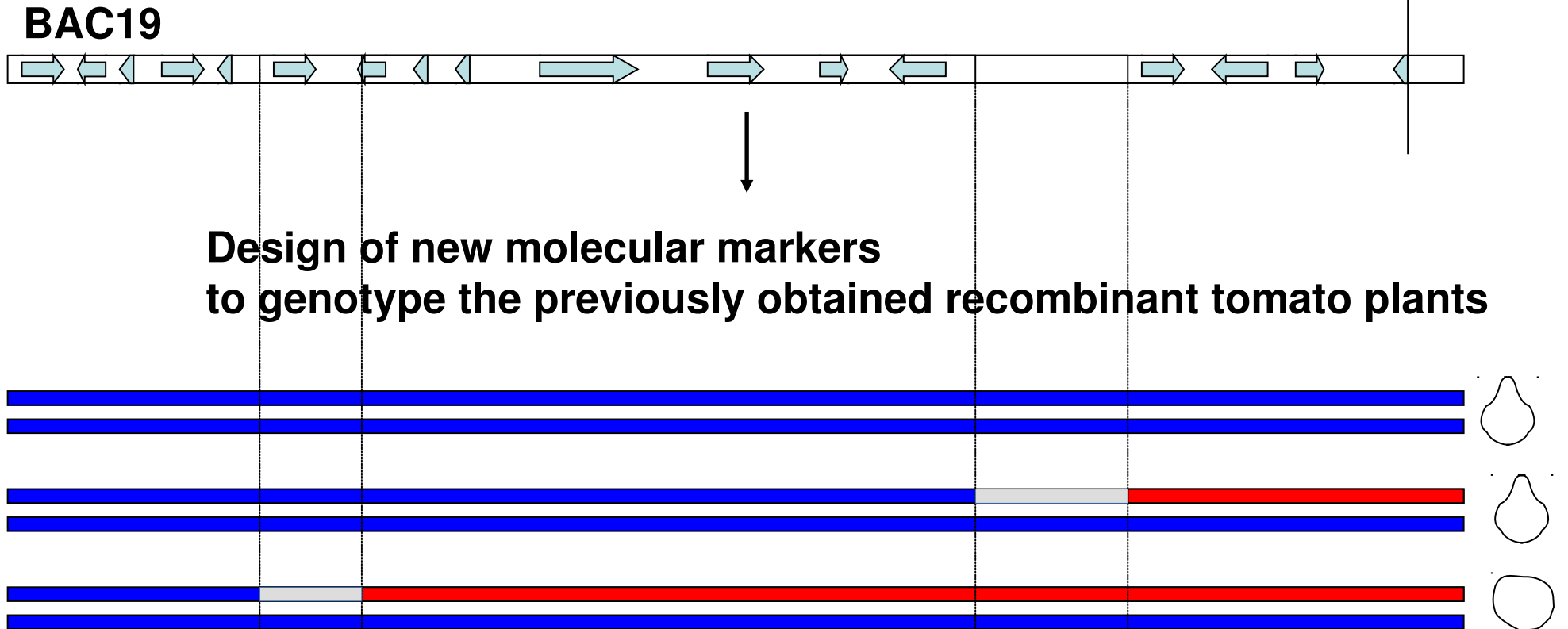
contains genomic DNA fragments of 100-350kb

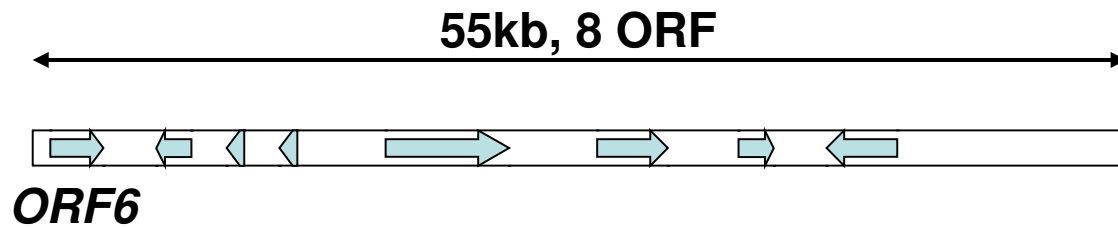


Screen of the library with marker TG645



BAC19 containing 105kb, 17 ORF (open reading frame) TG645





Sequencing of the region in the 2 tomato varieties

L. esculentum
cv. Yellow Pear



1 SNP (single nucleotide polymorphism)
et 1 indel (insertion-deletion) of 2bp
in non-coding regions

L. esculentum



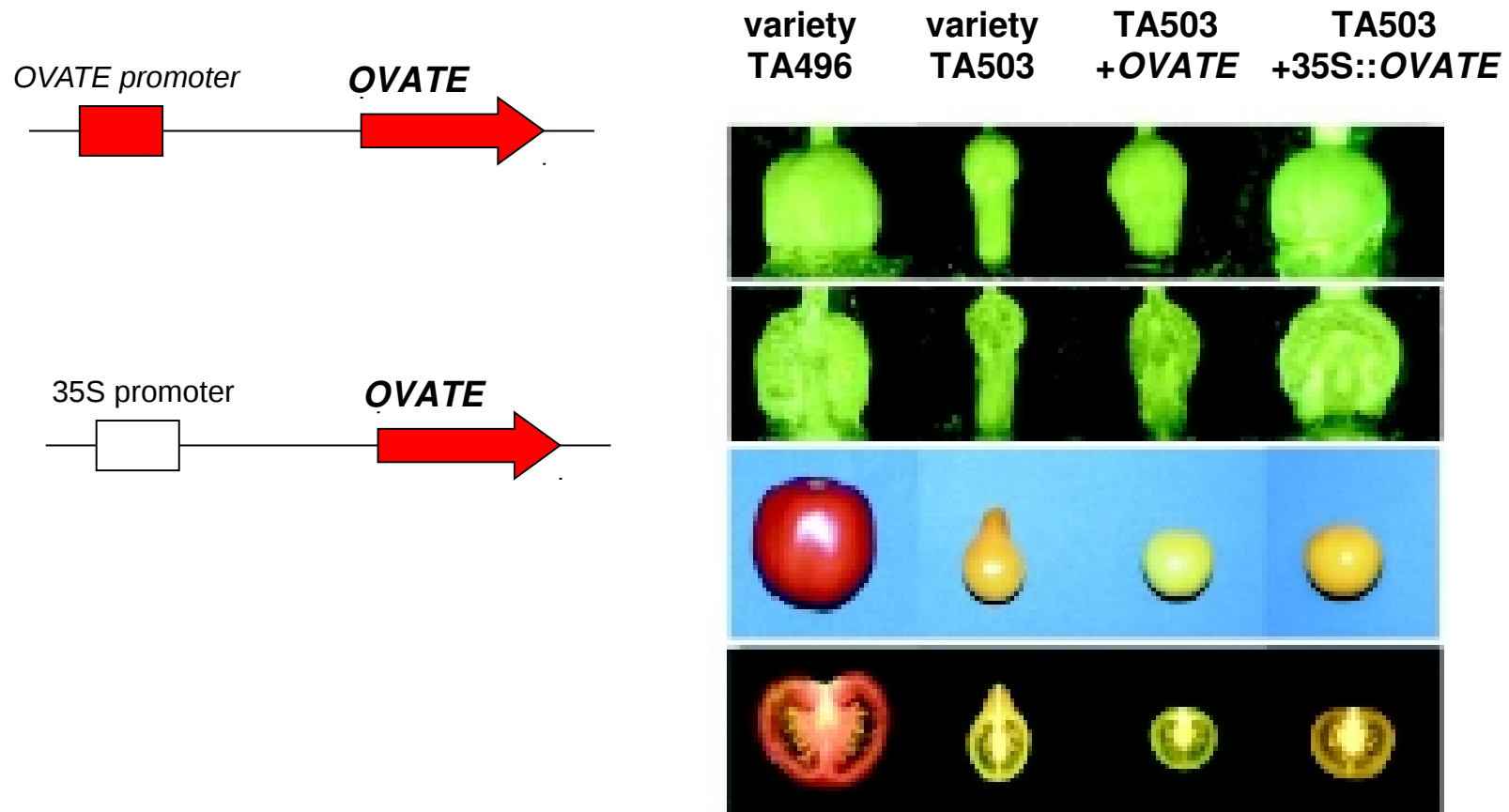
1 SNP in *ORF6* : G496T, stop codon stop,
truncated protein with last 75 amino acids
missing

Hypothesis: the causing gene is *ORF6* = *OVATE*

The causing gene is *OVATE/ORF6*

Same mutation in 3 other pear tomato varieties

Complementation of the mutation by transgenesis



OVATE = protéine à domaine NLS (nuclear localisation signal), fonction inconnue, exprimée dans les fruits en développement mais pas dans l'appareil végétatif

Evolution of morphology in threespine sticklebacks



marine



Paxton Lake, Canada

Gasterosteus aculeatus

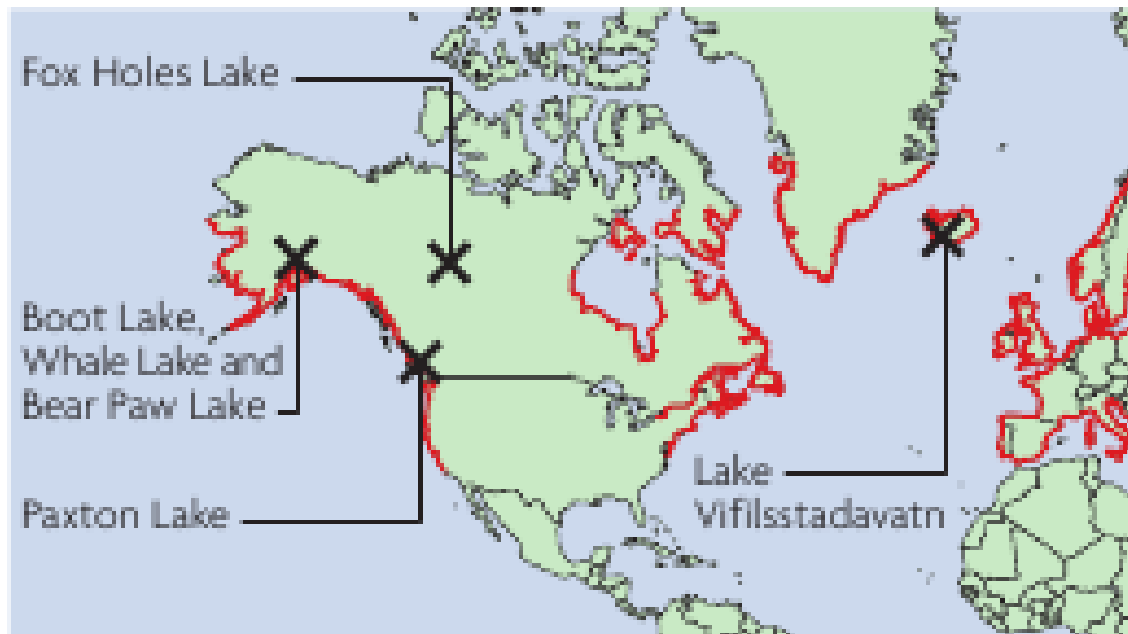
(Peichel et al., 2001 ; Shapiro et al, 2004 ; Chan et al. 2010)

Marine fishes with robust pelvis = ancestral

**Freshwater fishes with reduced pelvic structures = derived,
independently at least 20 times**

- limited calcium availability
- absence of gape-limited predatory fishes
- predation by grasping insects

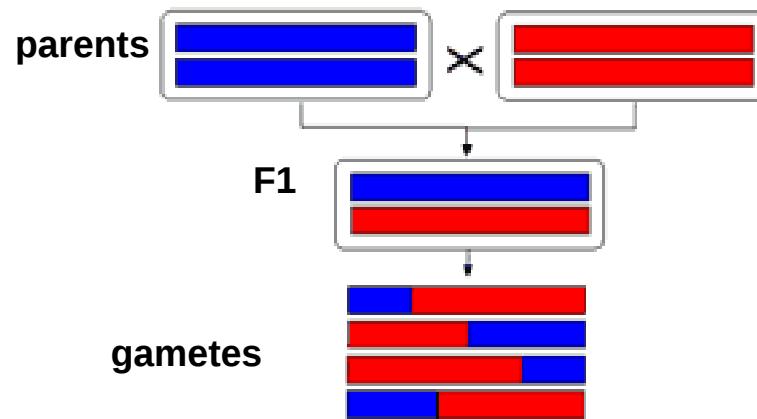
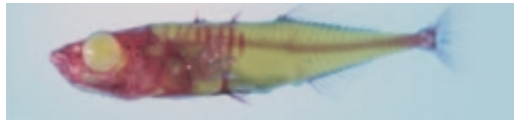
Last glacier retreat = 10 000 – 20 000 years ago



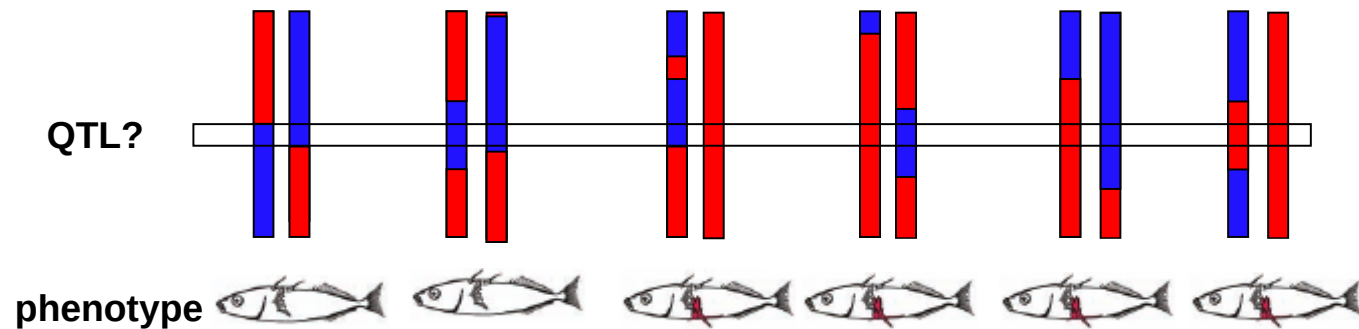
QTL mapping

lake

marine



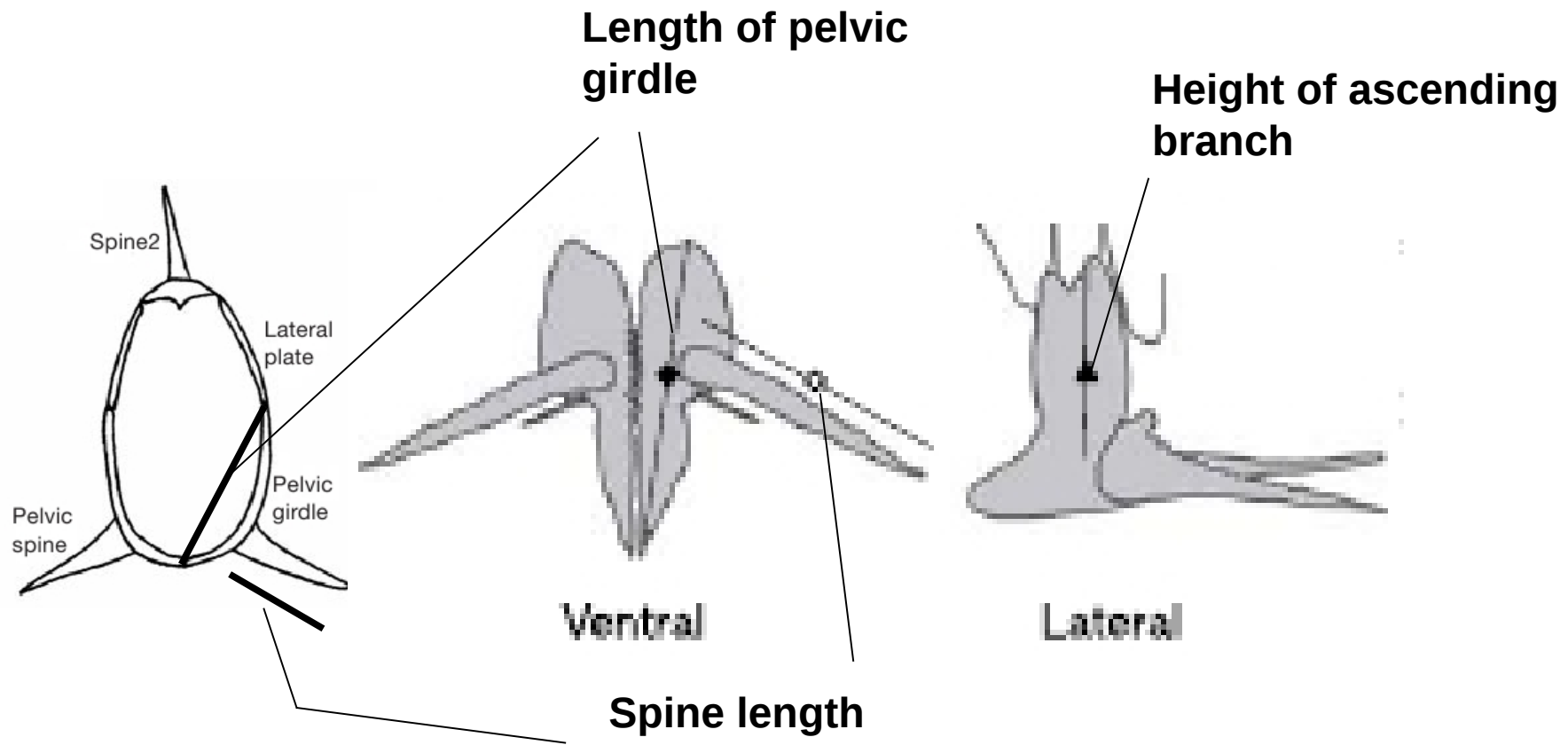
375 F2 individuals



phenotype

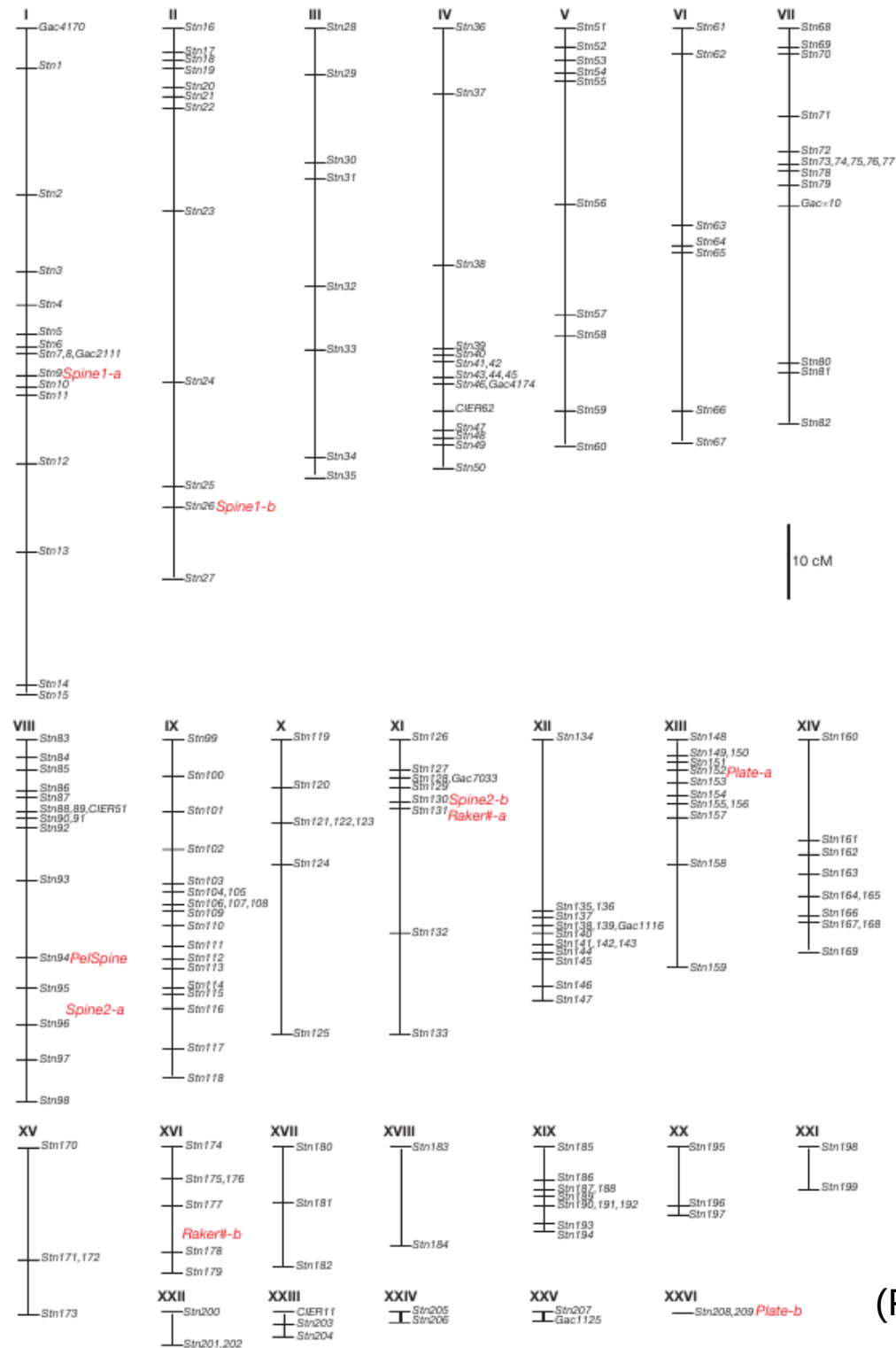
(Shapiro et al., 2004)

Quantitative measurement of the phenotype



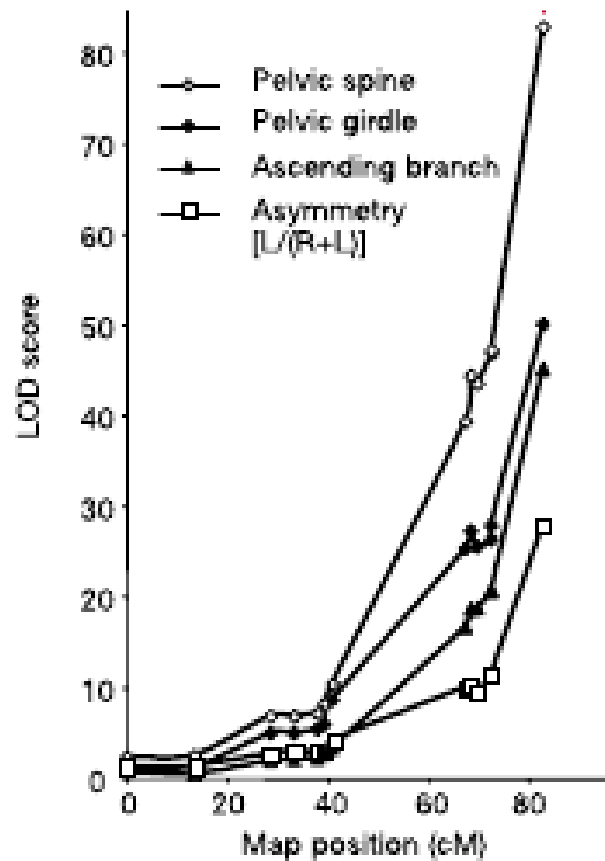
1000 microsatellite markers

26 linkage groups



(Peichel et al., 2001)

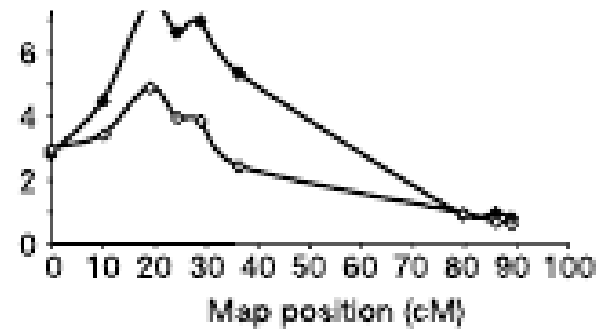
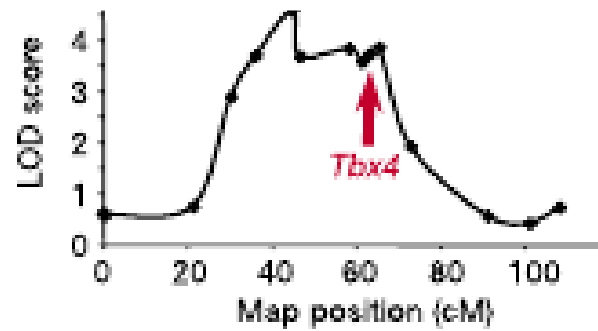
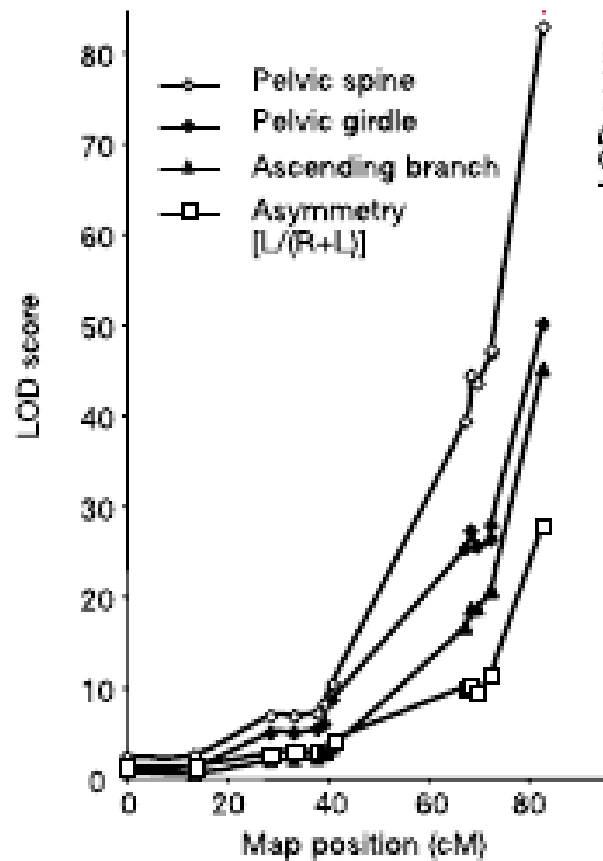
One major locus at the end of linkage group 7



Major locus responsible for 65% of the variance

One major locus at the end of linkage group 7

A few minor loci



Major locus responsible for 65% of the variance

Three candidate genes: *Pitx1*, *Pitx2*, *Tbx4*



Screen of a BAC library (Bacterial Artificial Chromosomes)
containing 100-350-kb fragments of stickleback genomic DNA



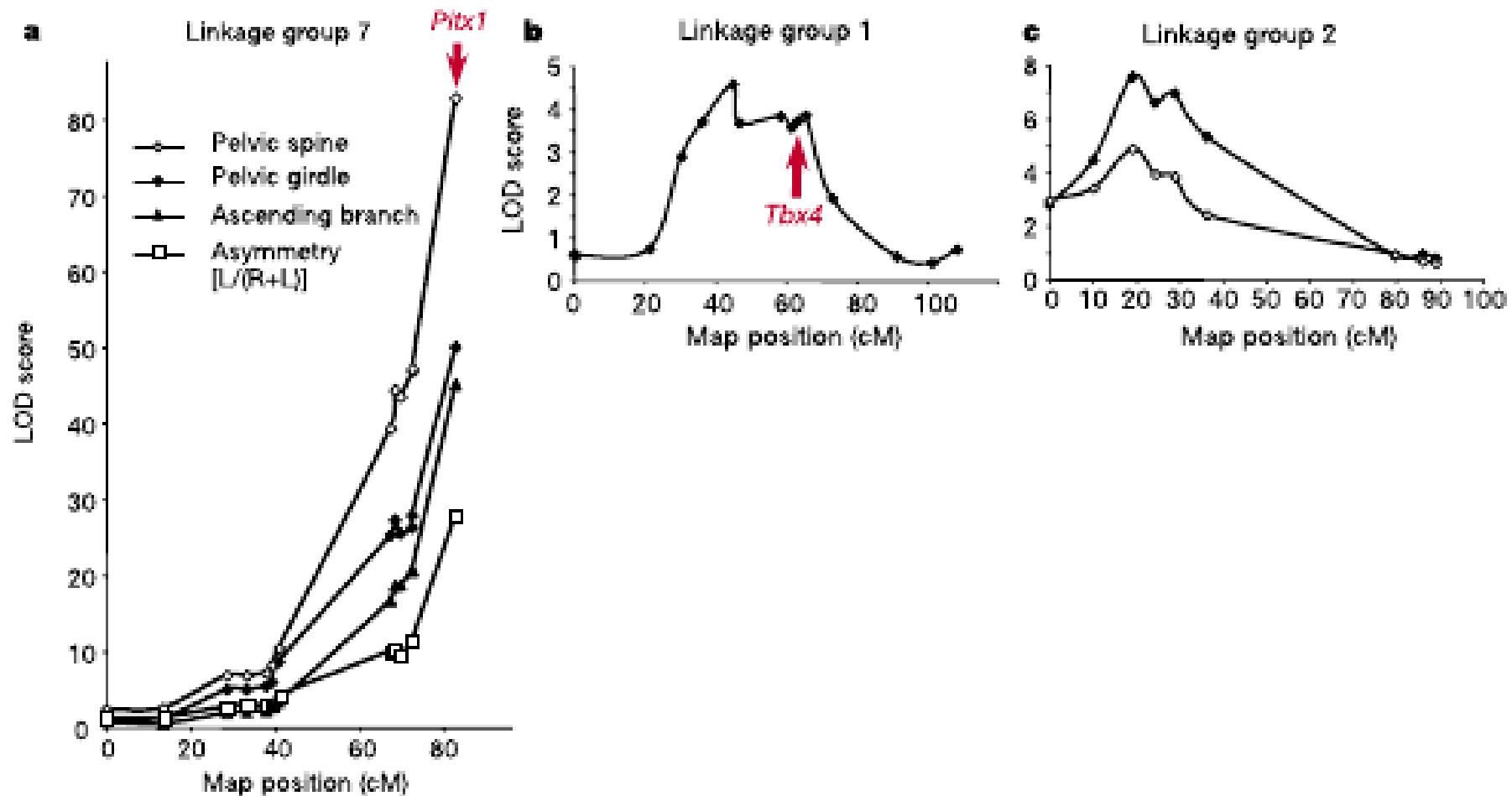
Partial sequencing and generation of new markers



Genotyping of 375 F2 individuals

One major locus at the end of linkage group 7

A few minor loci



***Pitx1*, responsible for the phenotypic change?**

Pitx1 null mutations in mice
(pelvis reduction, stronger on right side)

QTL mapping

Same coding sequence in lake and marine forms

Pitx1 expressed at stage 29 in marine individuals but not in lake individuals

***Pitx1*, responsible for the phenotypic change?**

Pitx1 null mutations in mice
(pelvis reduction, stronger on right side)

QTL mapping

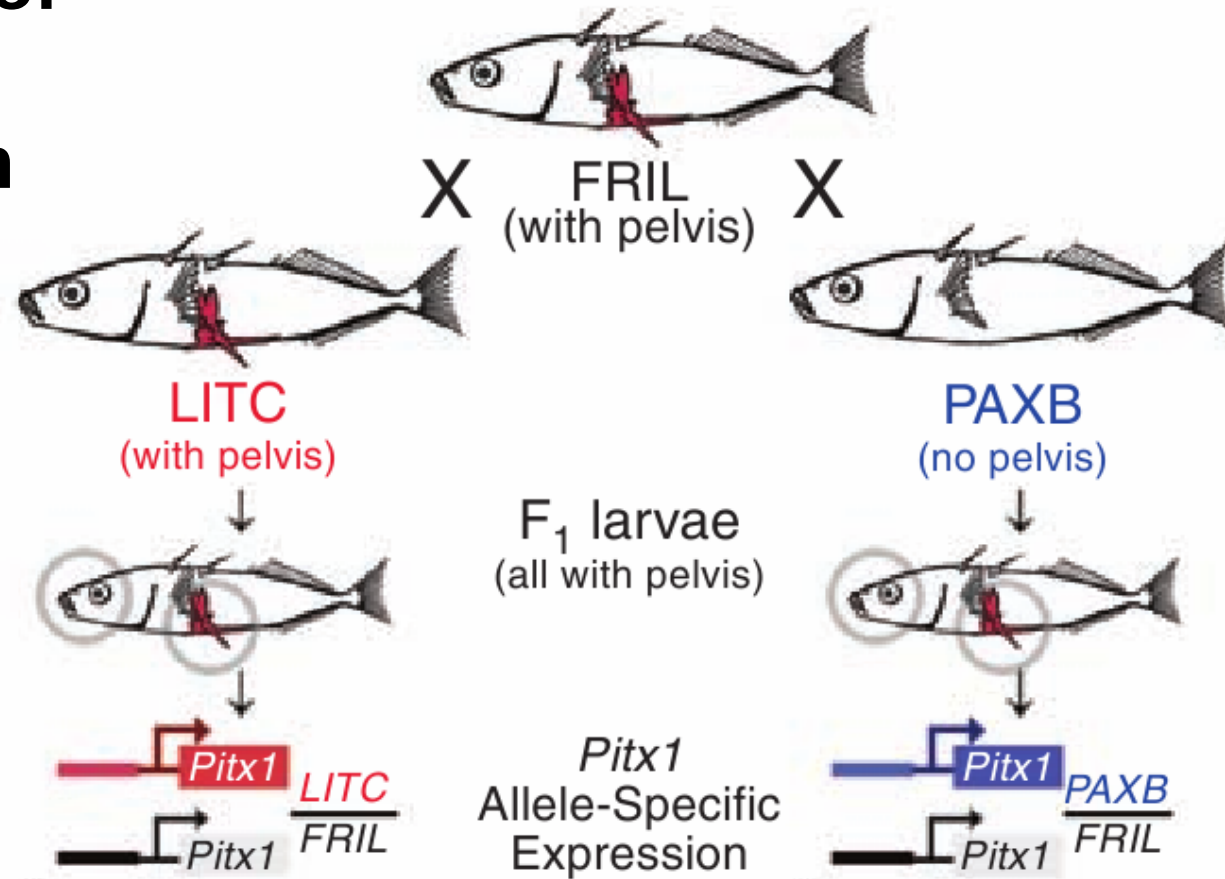
Same coding sequence in lake and marine forms

Pitx1 expressed at stage 29 in marine individuals but not
in lake individuals

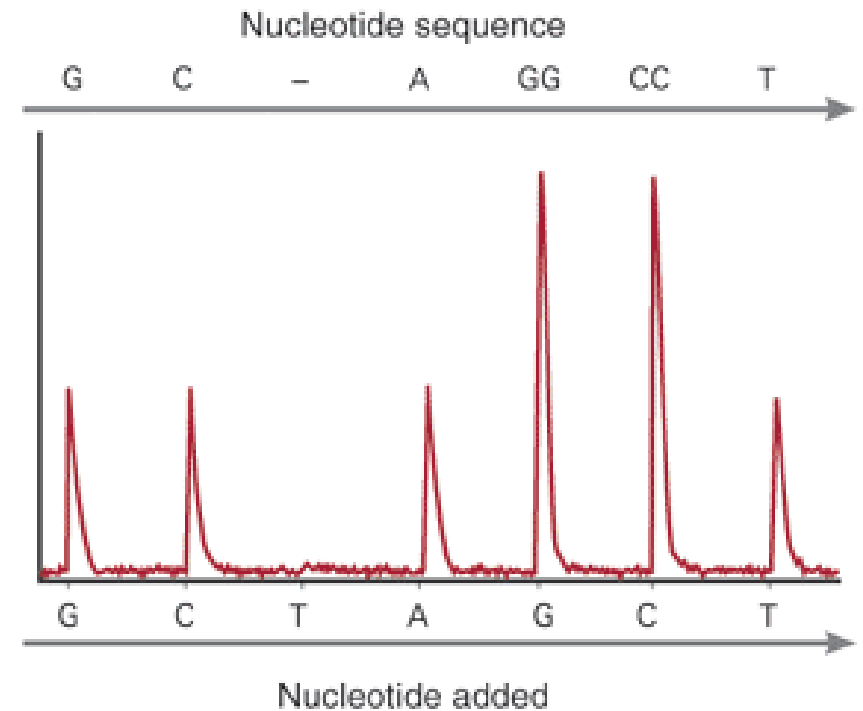
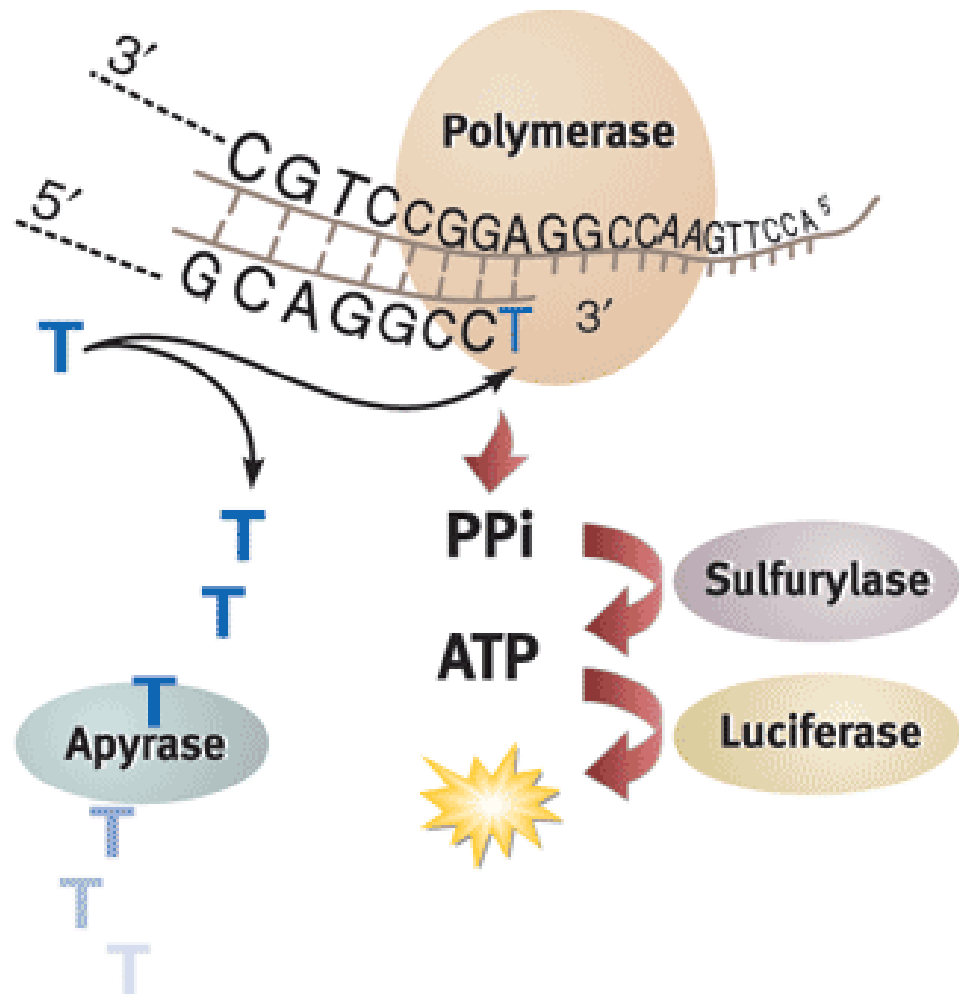
BUT

The decrease in *Pitx1* expression levels might
have evolved due to mutations in an upstream
regulatory gene

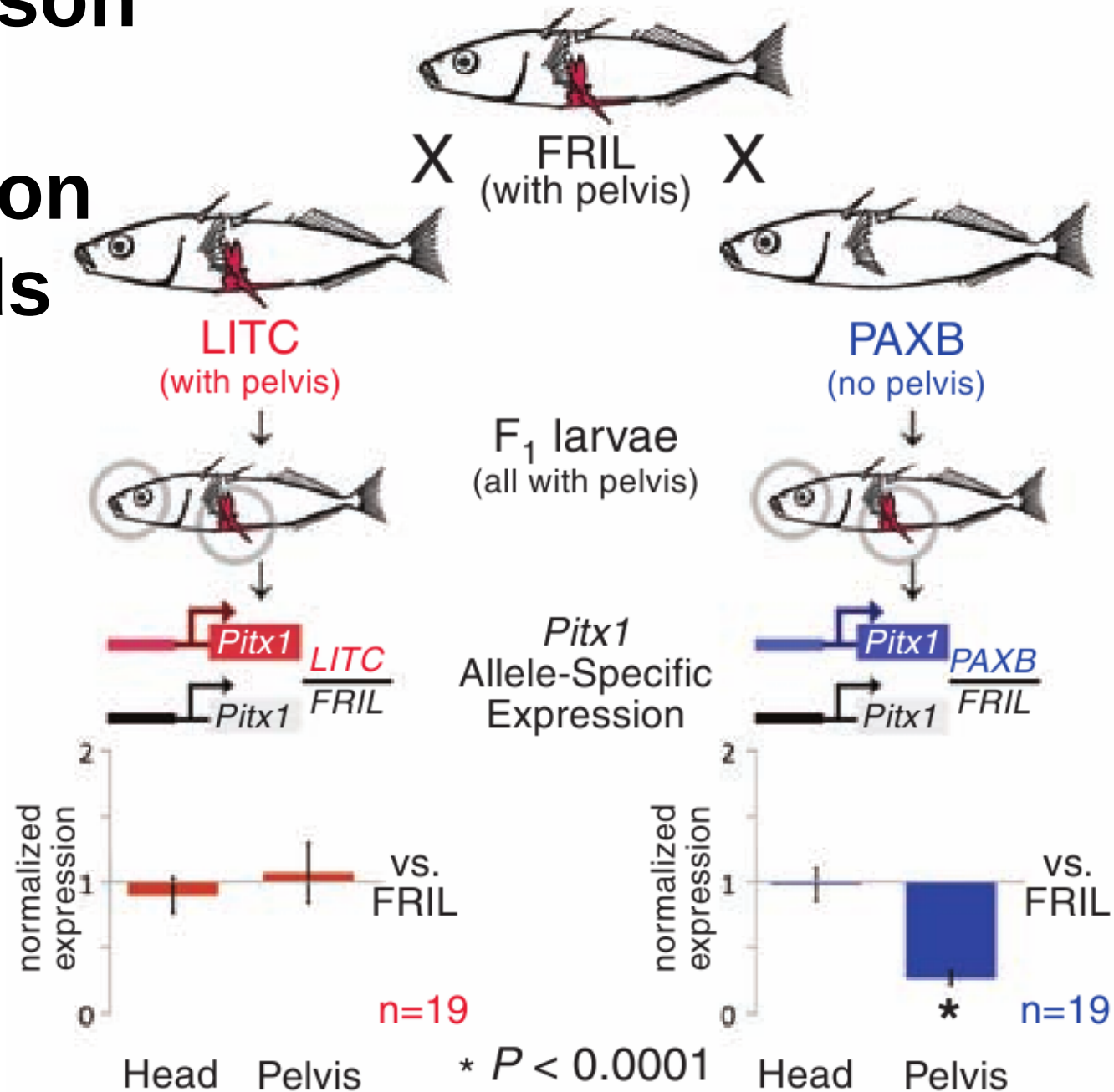
Comparison of allele expression in hybrids



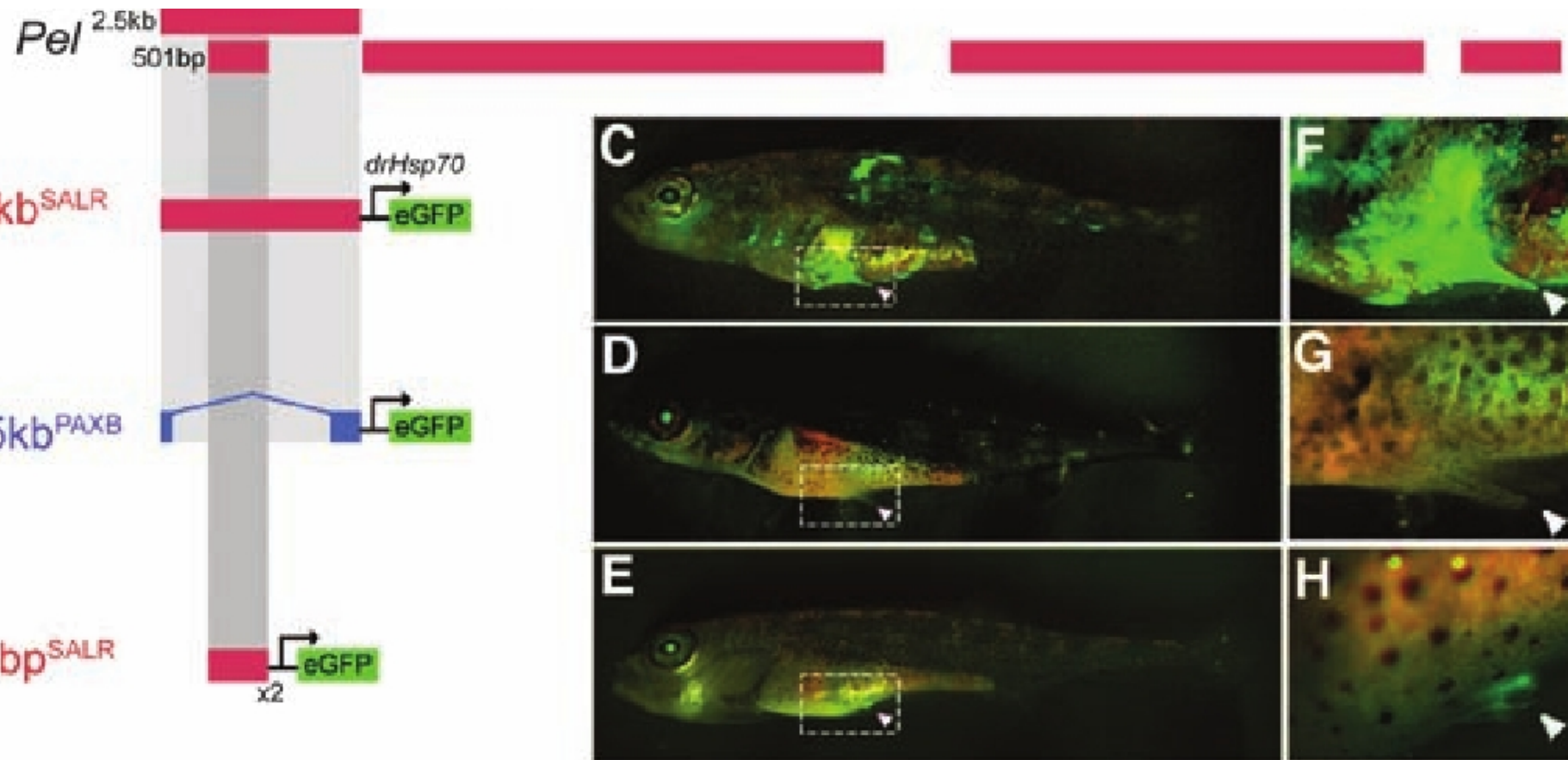
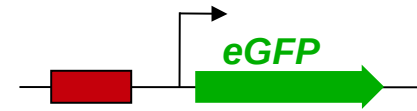
Pyrosequencing



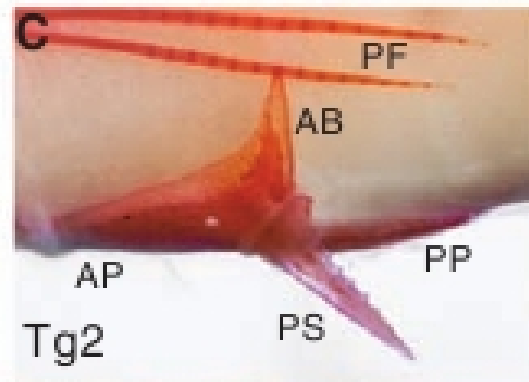
Comparison of allele expression in hybrids



Test of *Pitx1* cis-regulatory regions



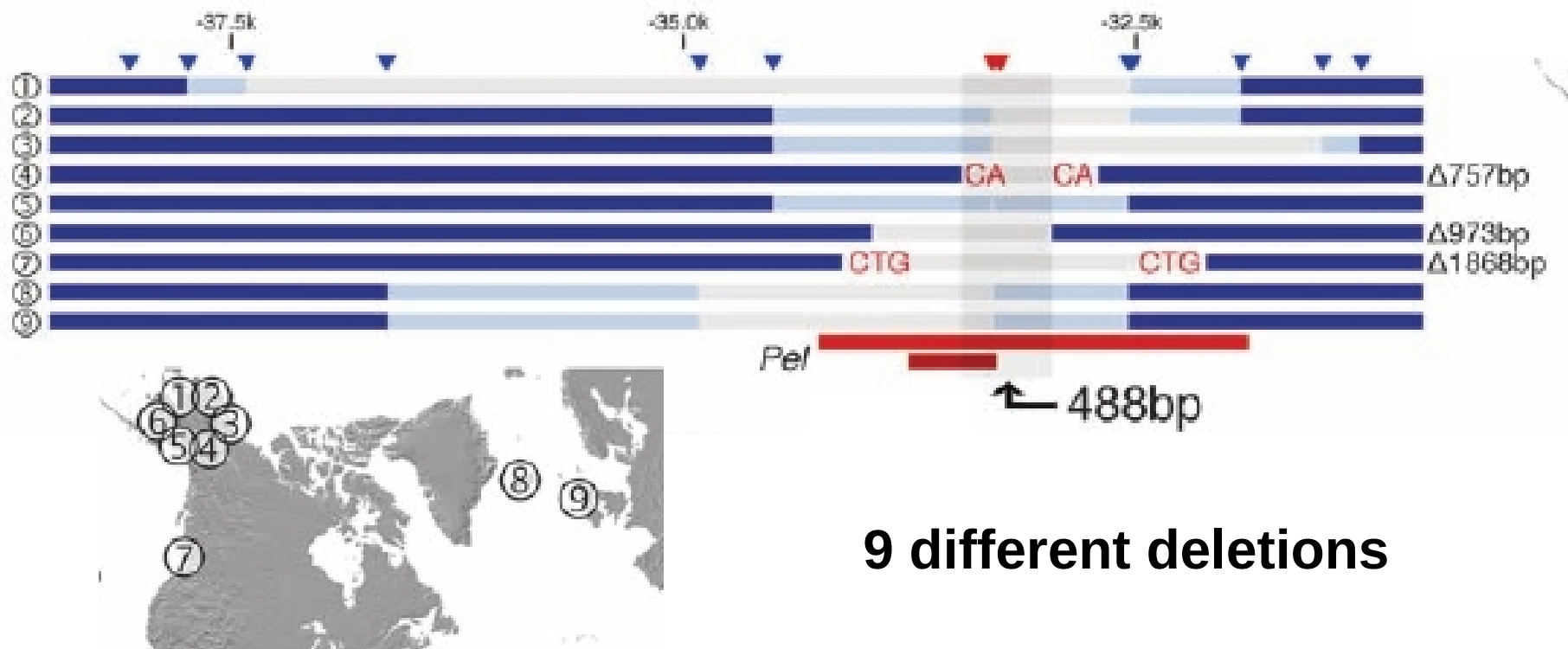
Rescue of a pelvis in freshwater individuals



Several independent deletions in the cis-regulatory region of *Pitx1*

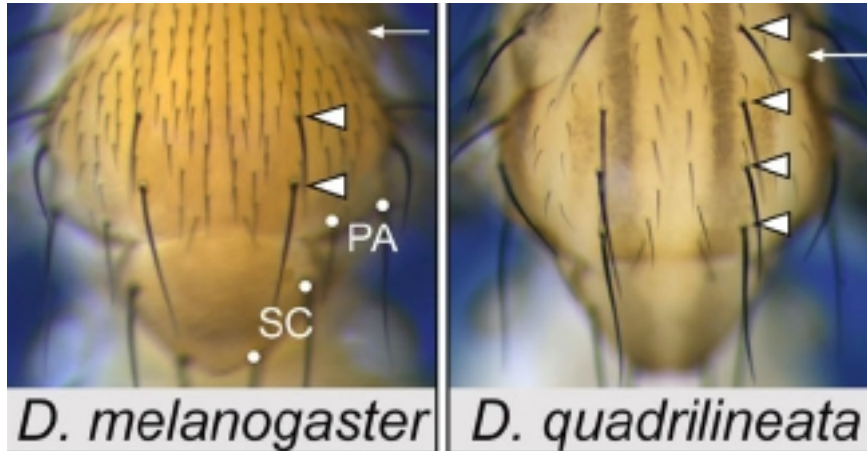
Region sequenced in two lake pop[ulations]: a 2-kb deletion in one
and a 757-bp deletion in the other one

SNP genotyping in 13 populations with reduced pelvis
and in 21 populations with complete pelvis

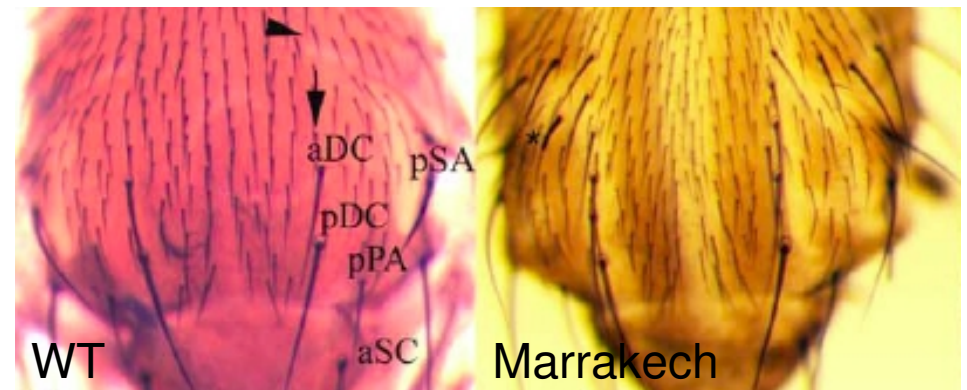


Evolution of extra bristles

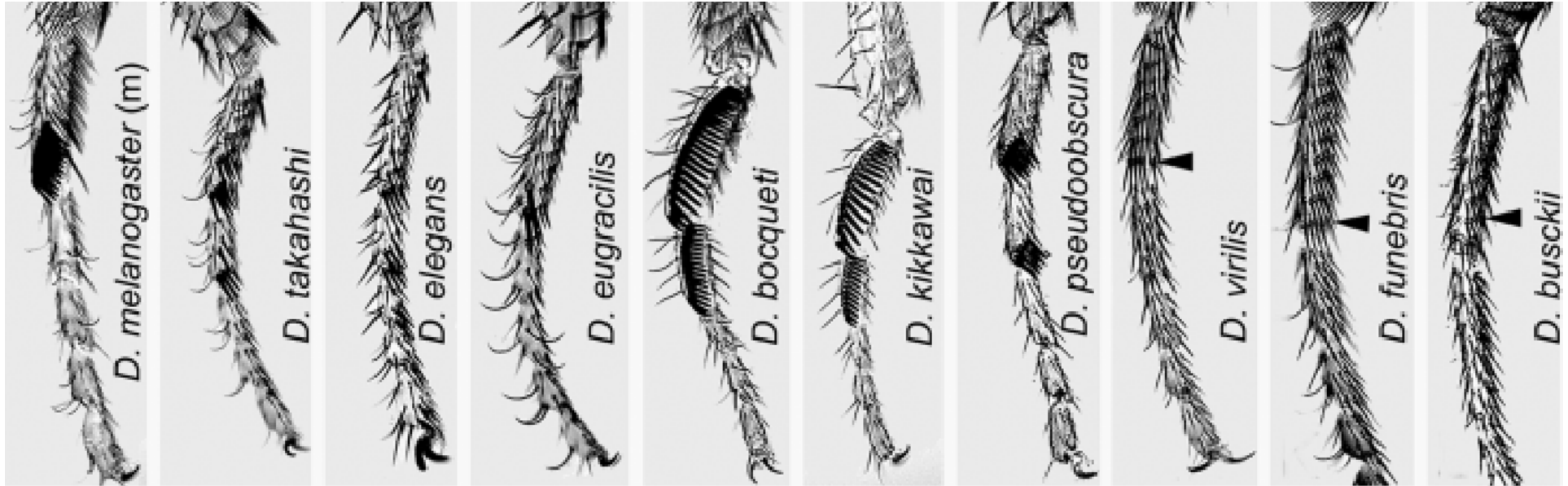
Interspecific change
in *D. quadrilineata*



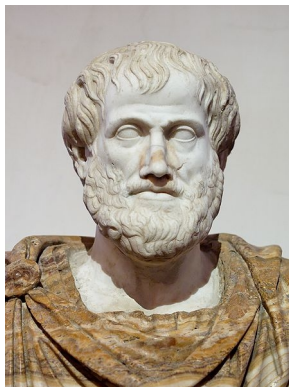
Intraspecific change
in *D. melanogaster*



Finding genetic rules on bristle evolution



Randsholt and Santamaria 2008



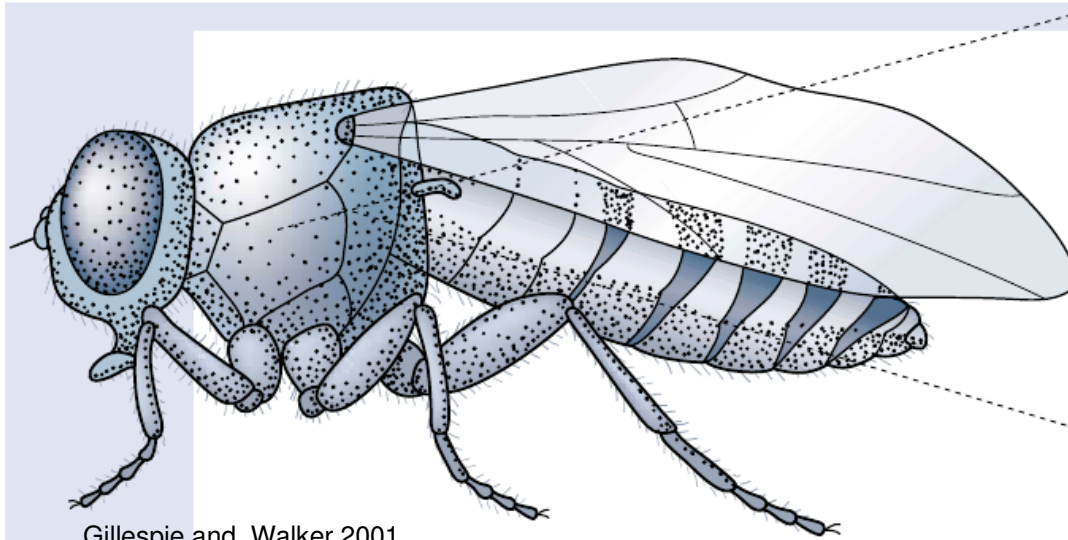
color, type, orientation
shape and size
presence/absence
position

} CRE mutations
in *achaete-scute*

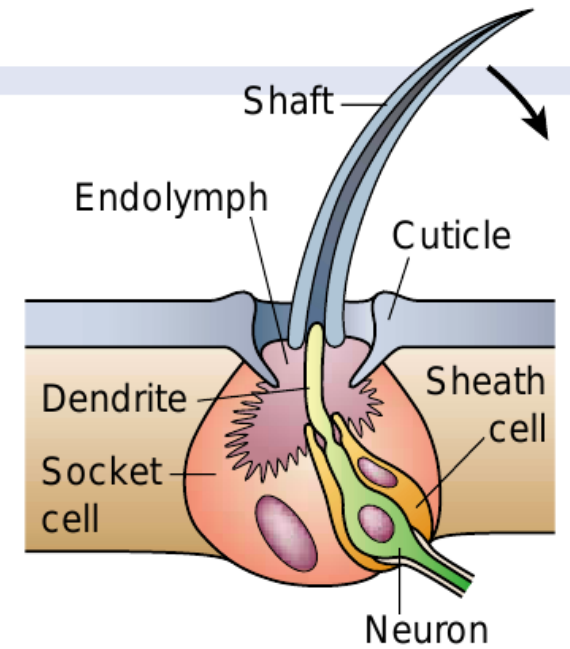
Aristotle, Historia animalium, book I, 2,
300BC

Stern and Orgogozo 2009

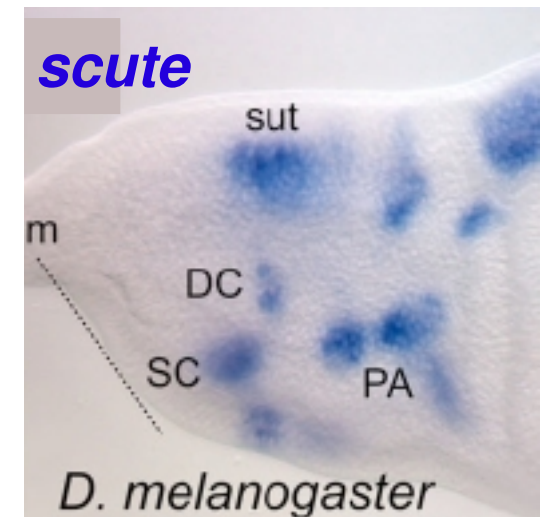
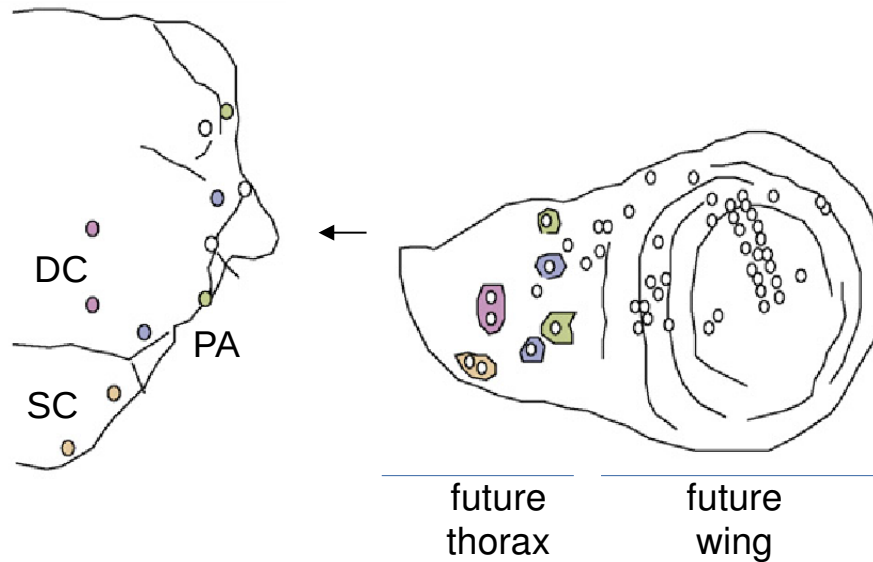
Bristle development

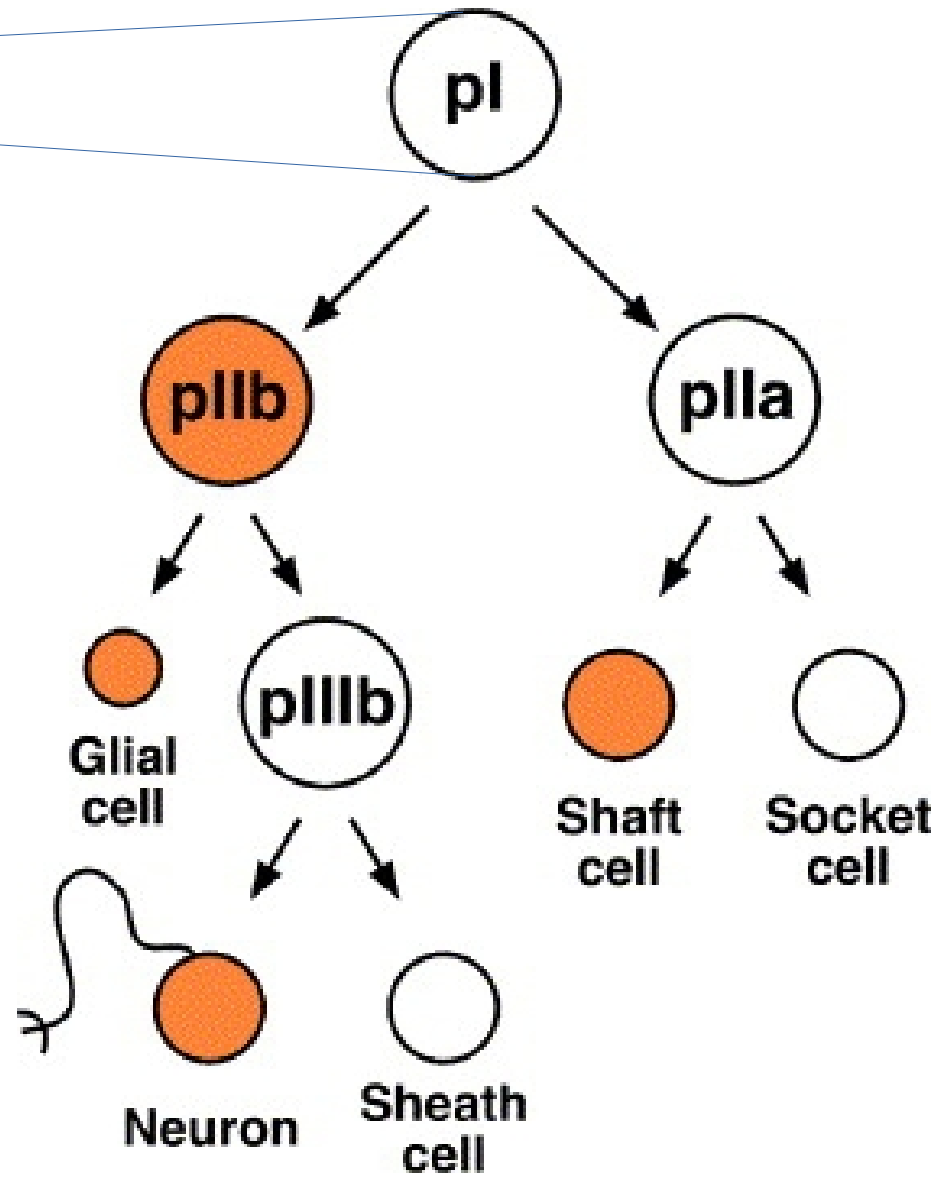
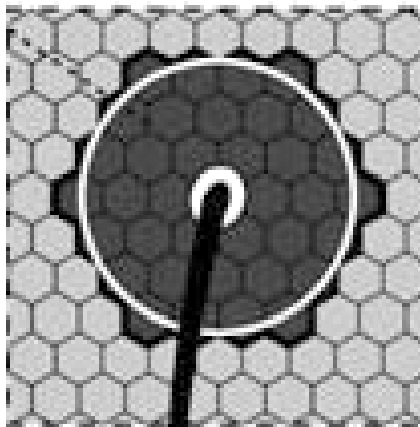
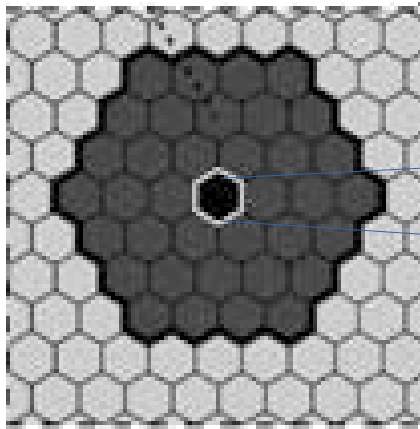


Gillespie and Walker 2001

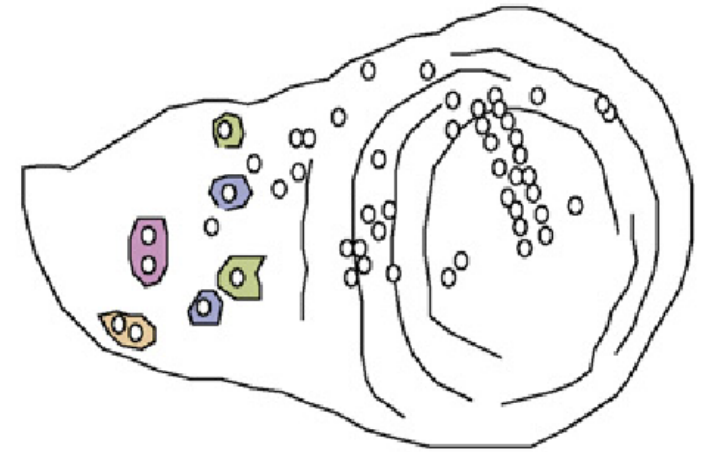
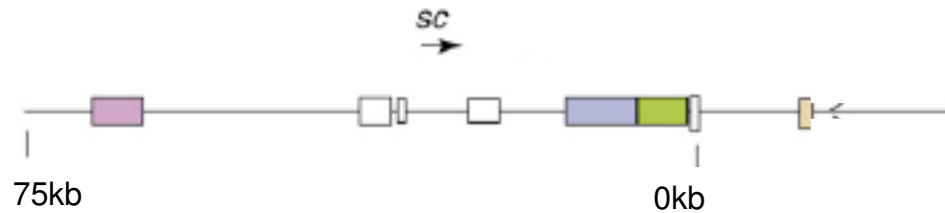


Simpson 2007

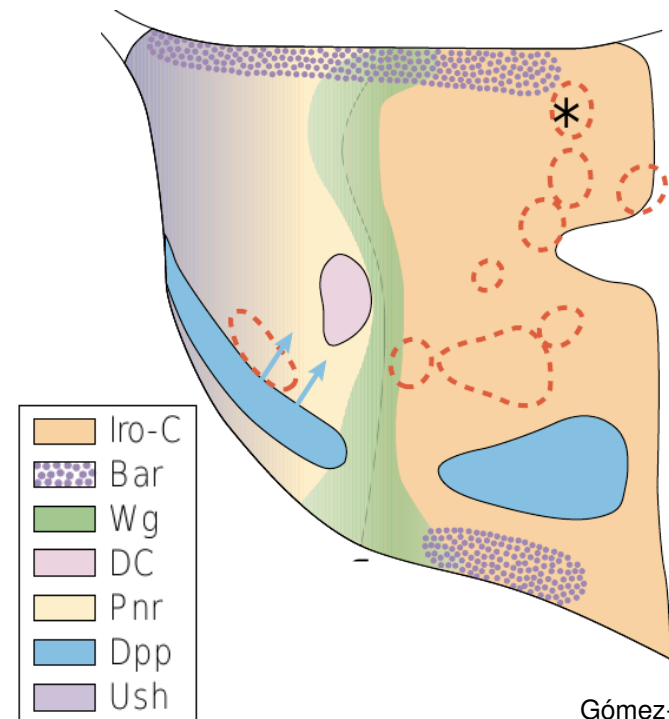
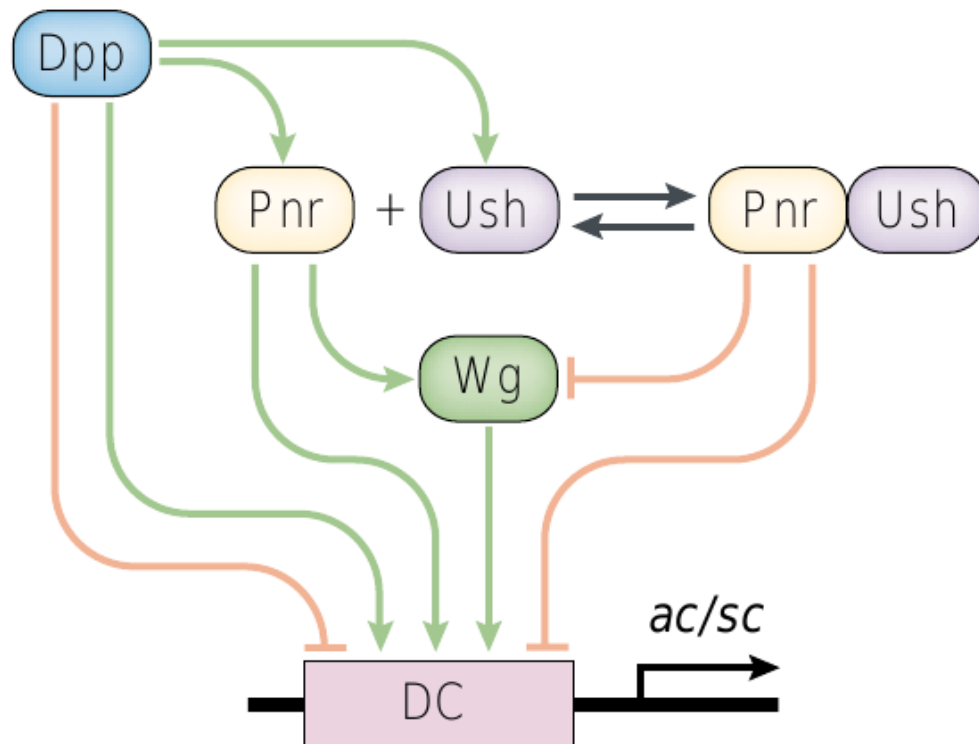




scute cis-regulatory elements are “master switches”

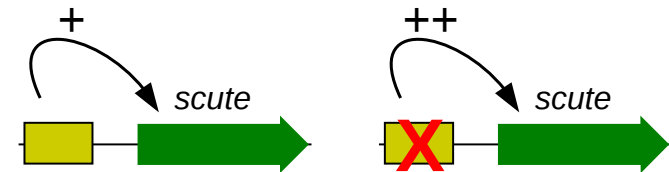
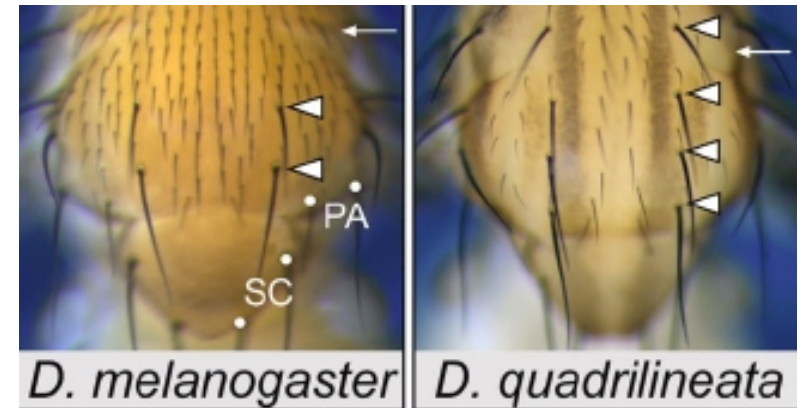
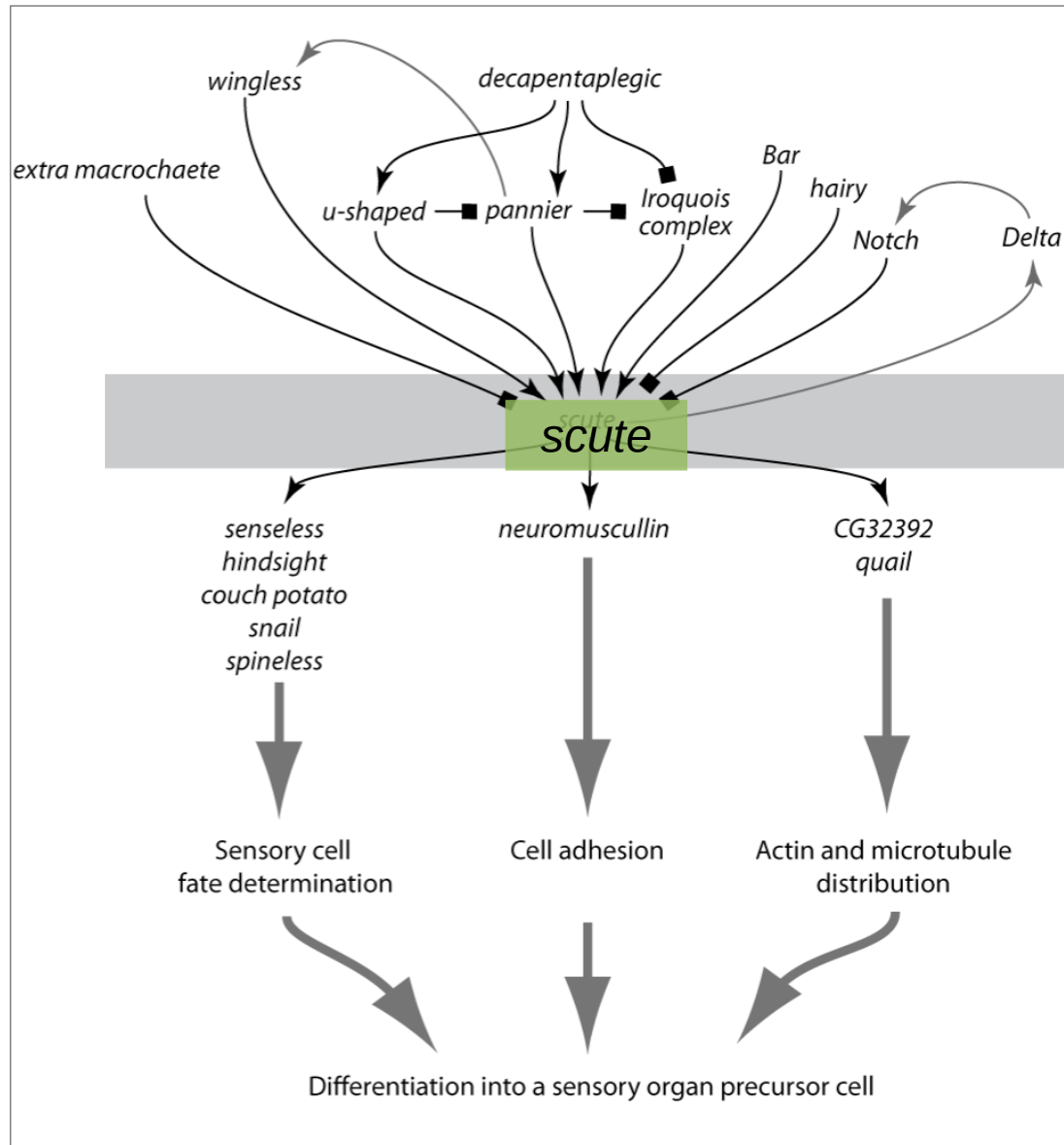


Simpson 2007

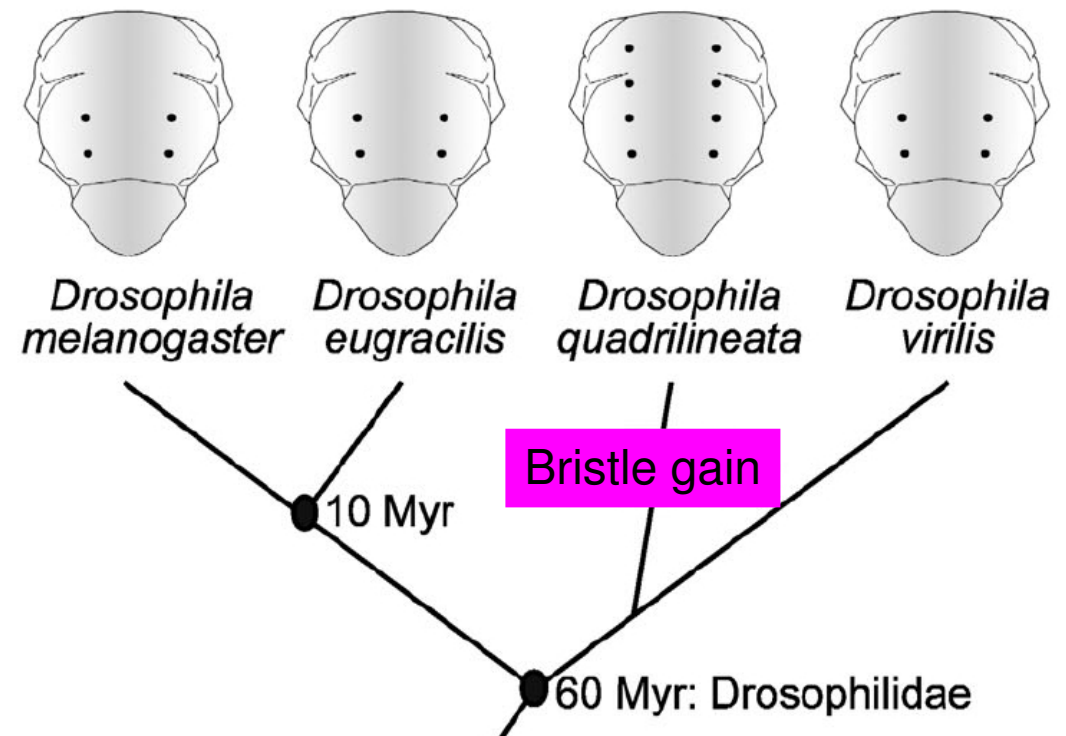
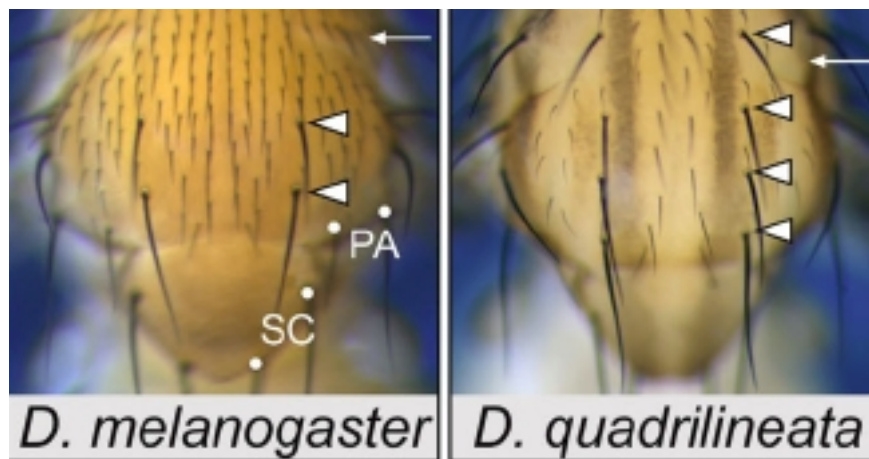


Gómez-Skarmeta 2003

Genetic evolution is partly predictable

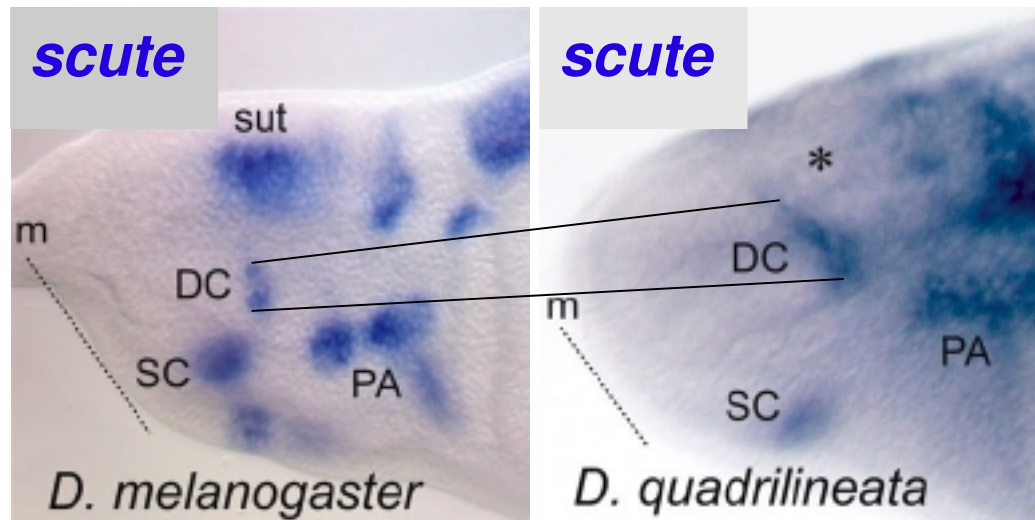


Extra bristles in *D. quadrilineata*



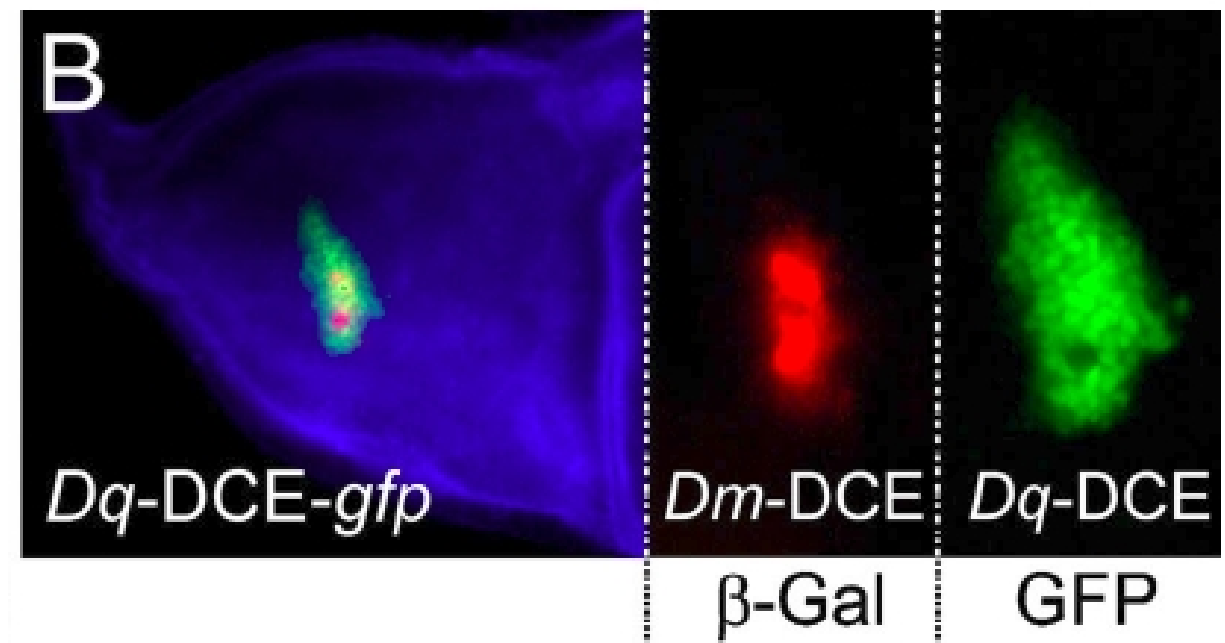
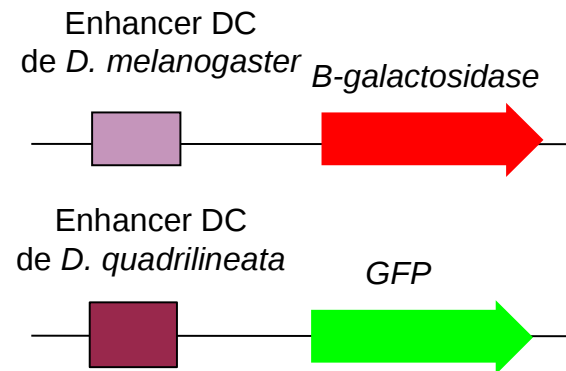
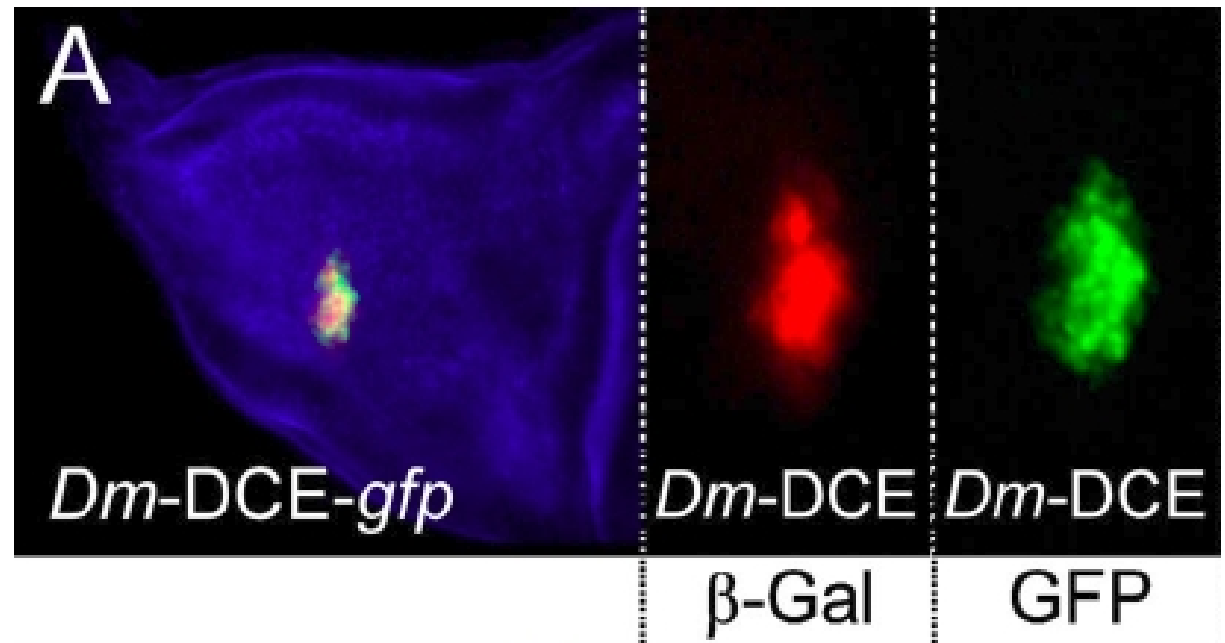
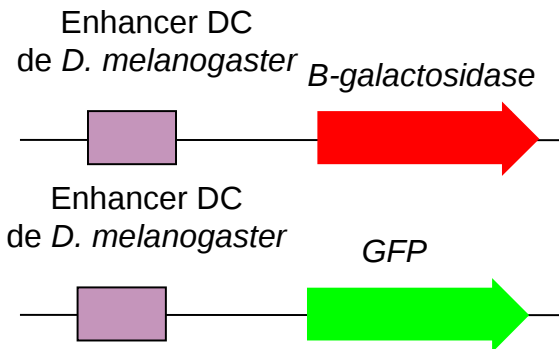
Extra bristles in *D. quadrilineata* correlate with larger *scute* expression domain

In situ hybridization



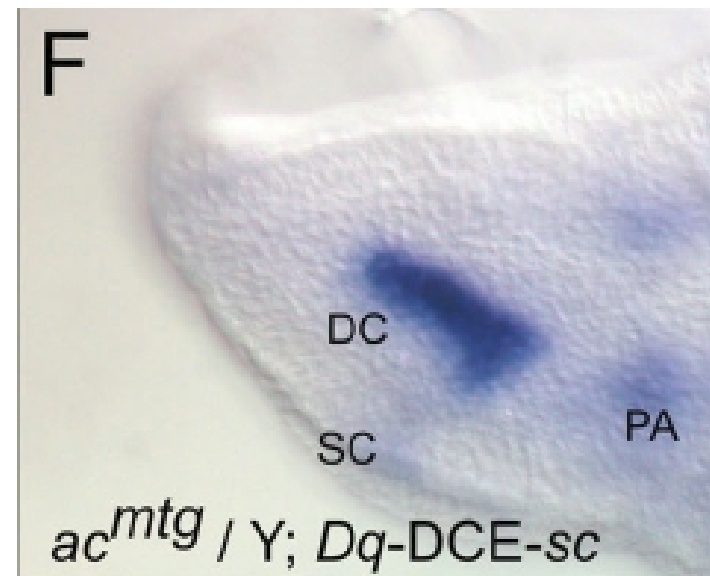
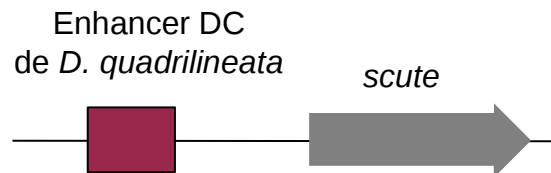
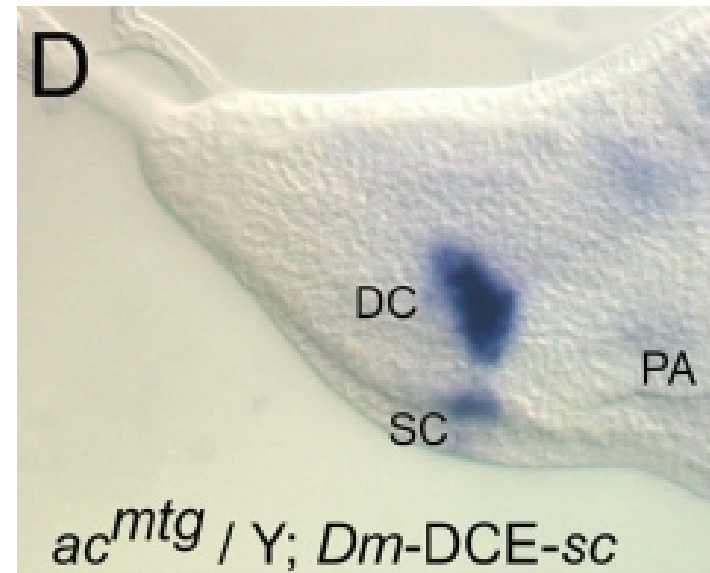
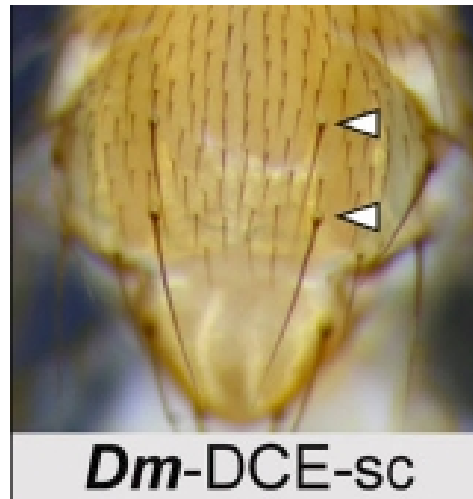
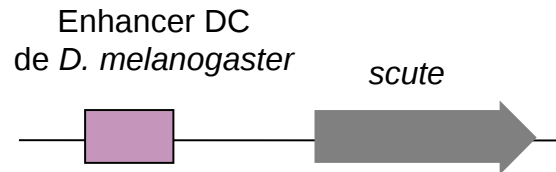
Test for a cis-regulatory change (1)

D. melanogaster
transgenics



Test for a cis-regulatory change (2)

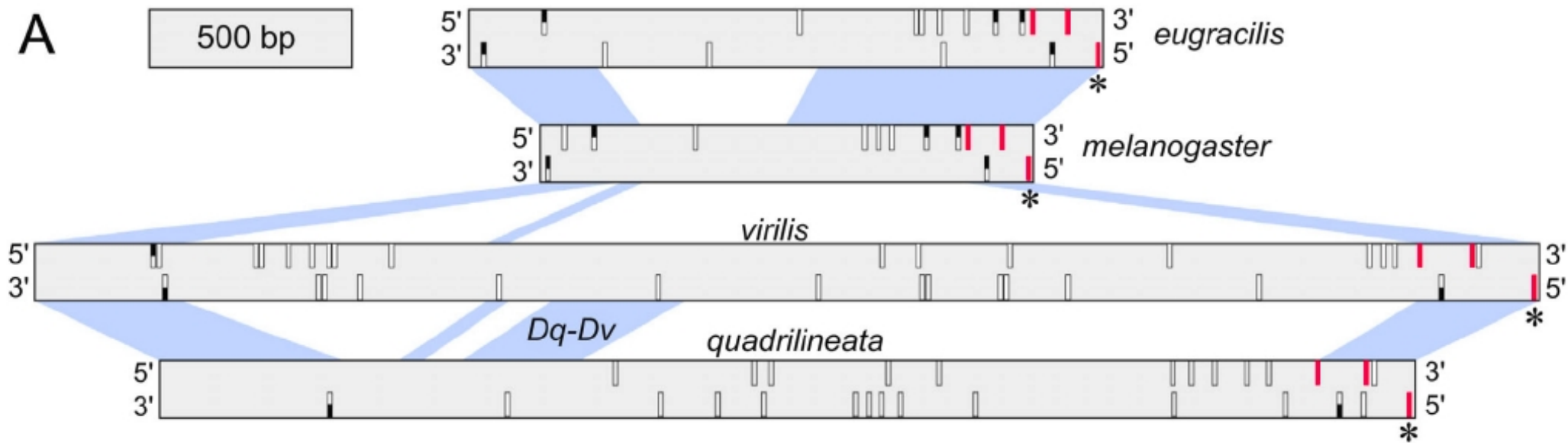
D. melanogaster
transgenics



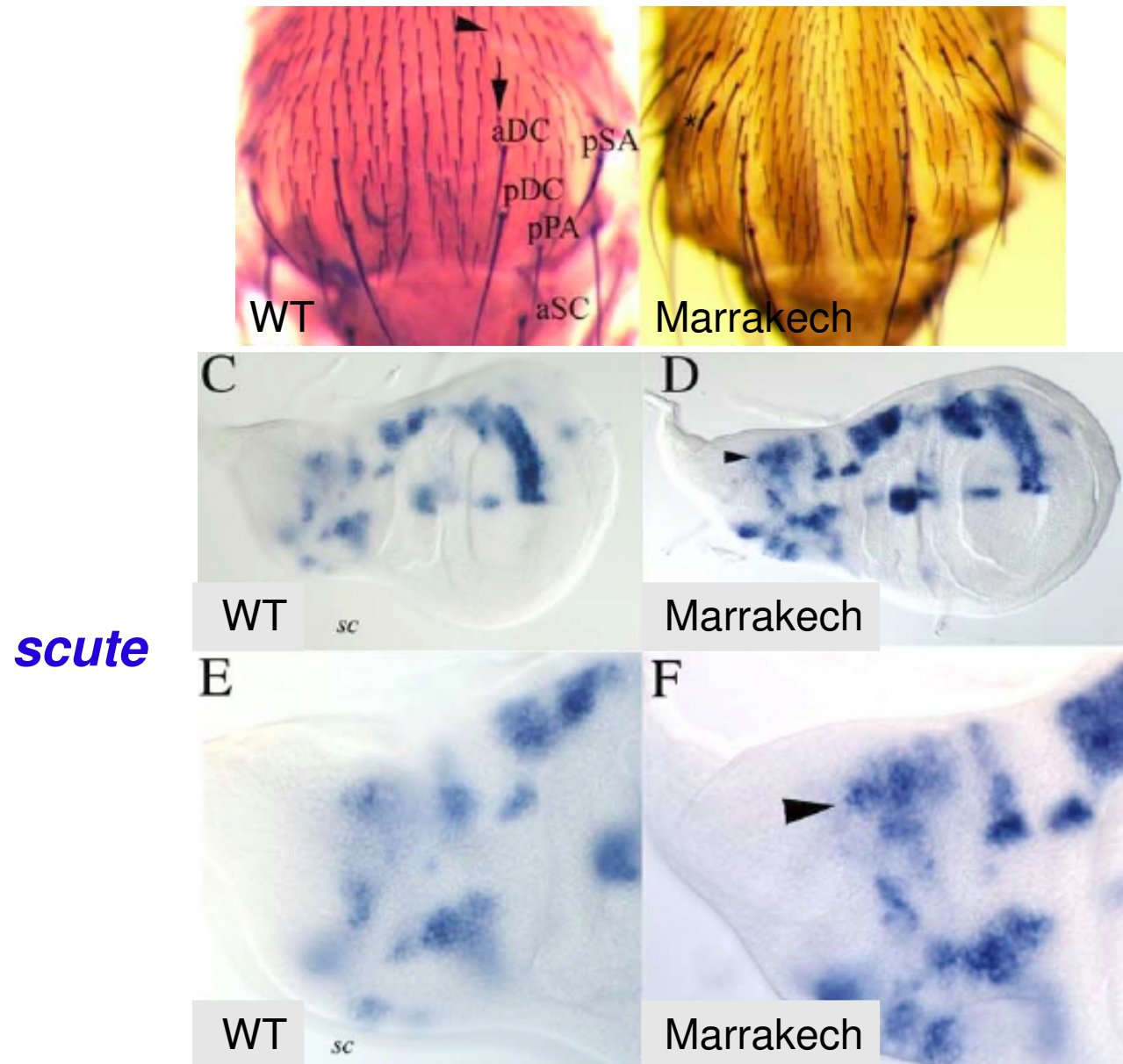
Alignment of the DC region

A

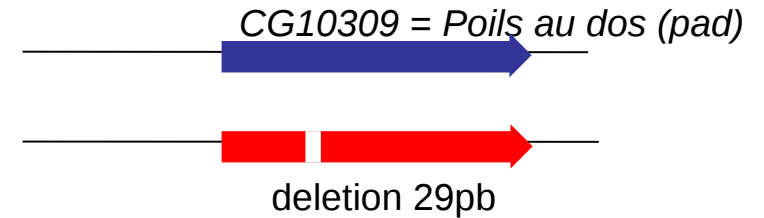
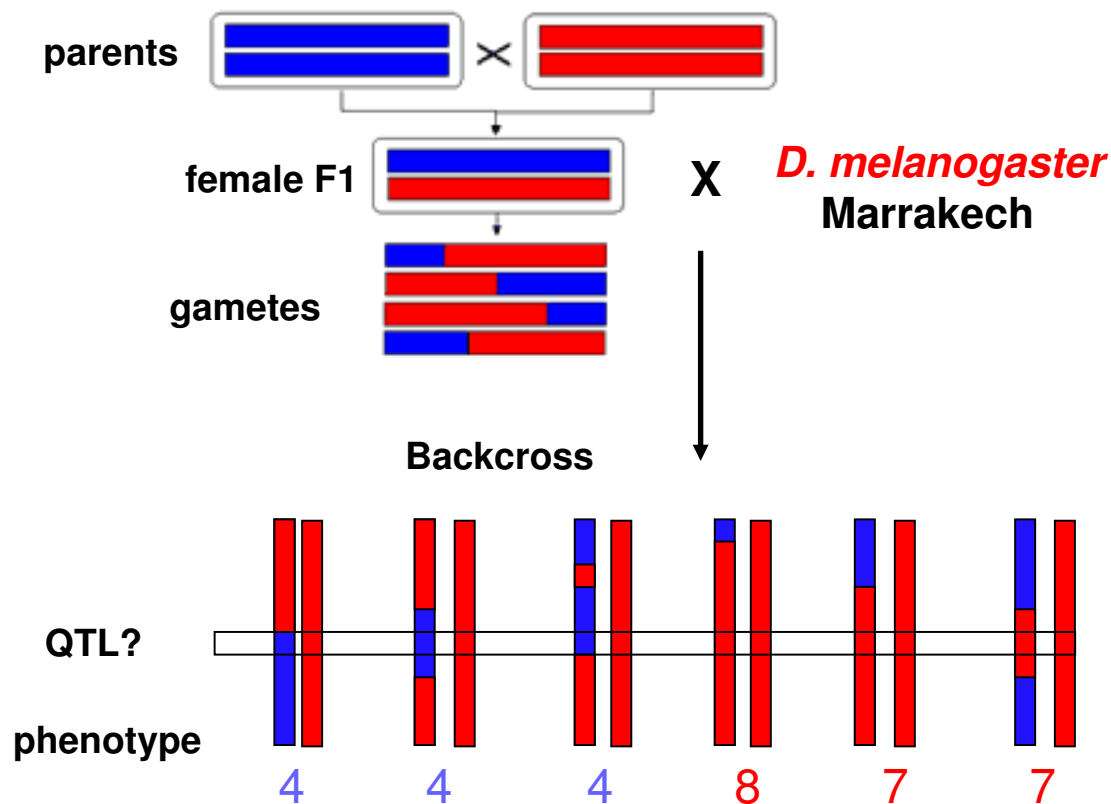
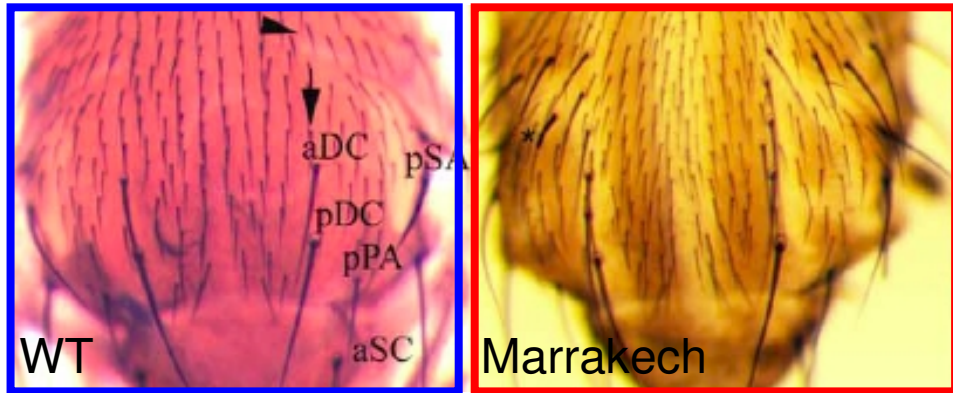
500 bp



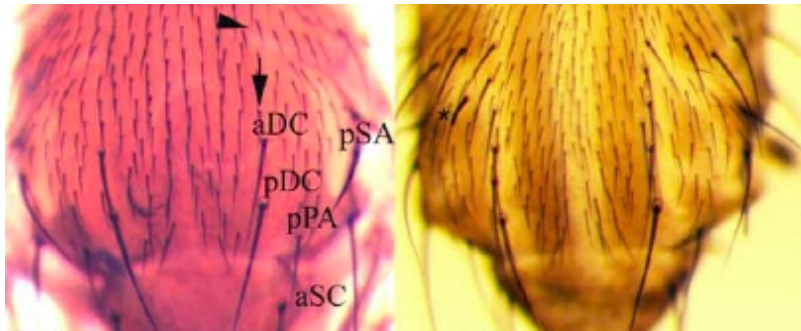
Extra bristles in *D. melanogaster*-Marrakech correlate with larger *scute* expression domain



Extra bristles in *D. melanogaster*-Marrakech due to mutation(s) in *poils-au-dos*

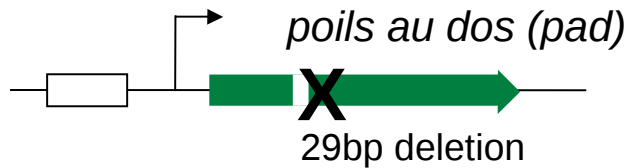


Short-term evolution...



D. melanogaster

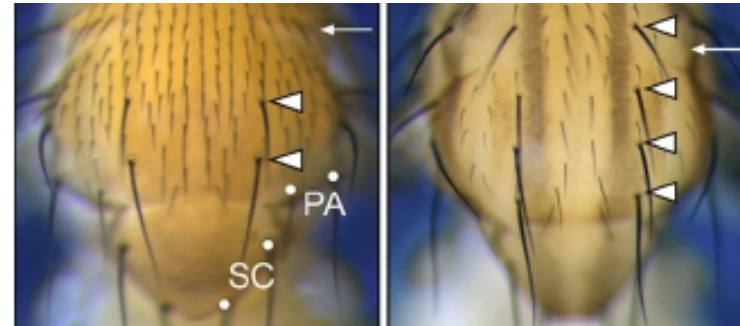
D. melanogaster variant



null mutation in coding region
change in thorax and wing

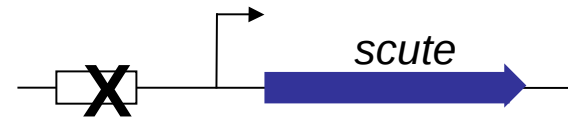
(Gibert et al., 2005)

...versus long-term evolution



D. melanogaster

D. quadrilineata



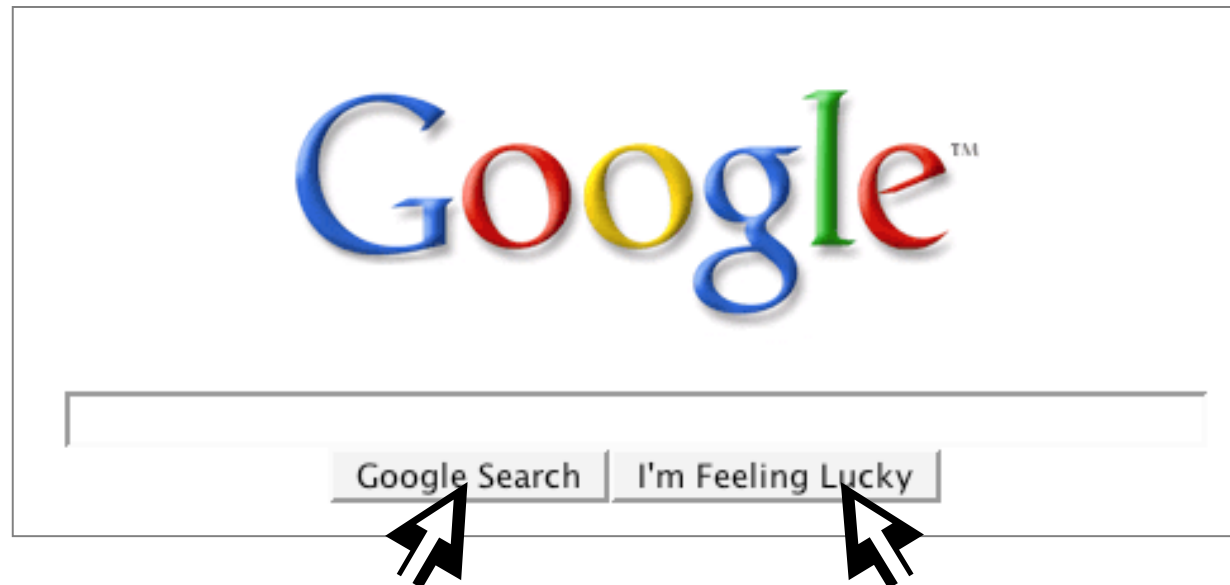
cis-regulatory mutation
change in the thorax only

(Marcellini et al. 2006)

Methods to identify the genes and the mutations responsible for phenotypic evolution



Two types of approaches



Genetic mapping

no a priori, fewer biases
long and tedious
rarely ends with identification of the gene

only with strains/species which produce fertile hybrids

Candidate gene

Based on an a priori idea
can be fast and efficient

will only find known genes

In both cases, genes with small effect are more difficult to identify

Various methods

Genetic

which chromosome (ex: autosomal versus sex)

QTL mapping

Genetic association studies

Complementation tests

General biology

General knowledge of the genes involved in the phenotype

Similarity with a known phenotype

Correlation with a change in gene expression level/pattern

Final test of protein activity

in vitro in *E. coli*, by transgenesis in the studied species or the closest model organism (ex: *beta-defensin* of dogs tested in mouse)

Final test of cis-regulatory regions

- with reporter constructs, transgenesis, comparison of both regions
- comparison of allele expression levels in hybrids (pyrosequencing)

QTL Mapping

4 steps: crosses, genotyping, phenotyping, statistical analysis

(Association study: no crosses, sample of a natural population)

Crosses for QTL mapping

Backcross with one line

Backcross in both directions

F₂

Crosses for several generations

Introgression lines

Recombinant Inbred Lines

...

Always try to maximize the number of recombination events

Markers for QTL mapping

yes-no PCR

PCR length polymorphism

Sequencing

Pyrosequencing

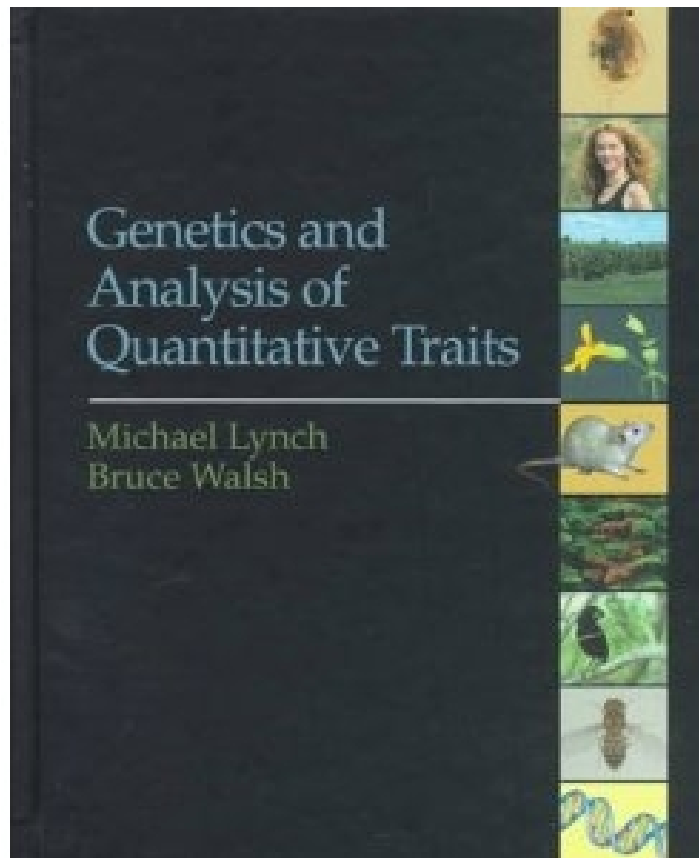
Probe hybridization

Microarray

RAD...

How many markers?

theory



practice

