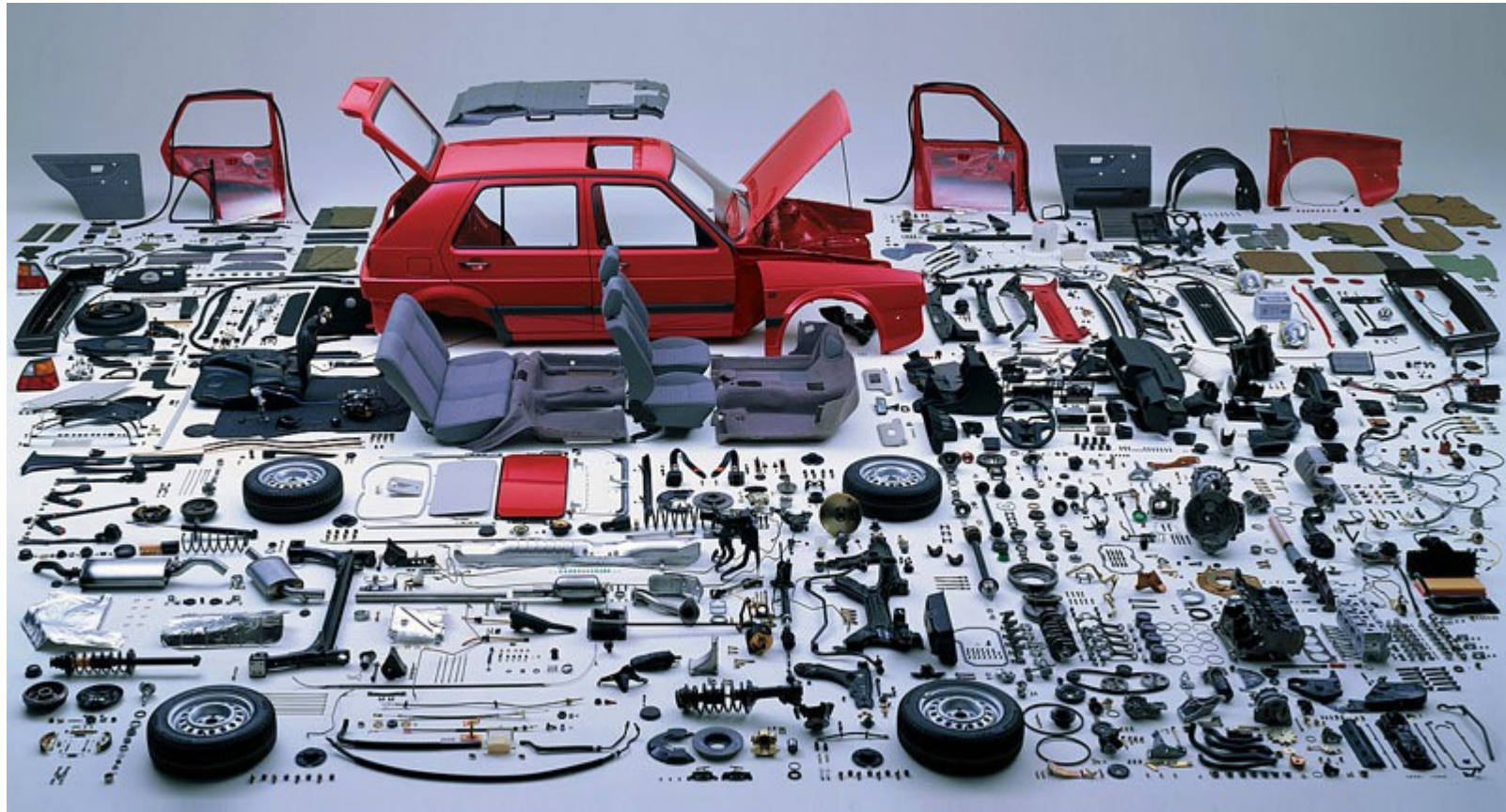


# **Genetic screens, Quantitative Genetics, Noise, Cryptic Variation, Robustness**



**Virginie Courtier-Orgogozo  
Institut Jacques Monod, Paris**

# Biochemistry versus Genetics



# Genetic screens

---

unbiased approach:

Biochemical screen (kinase assay, methylation assay, etc...)

Genetic screen (for genes)

# ***A genetic screen***

is a method used to find genes involved in a given phenotype

3 steps :

1. production of mutations
2. selection of individuals with the phenotype of interest
3. identification of the underlying genes

**Historical screen**

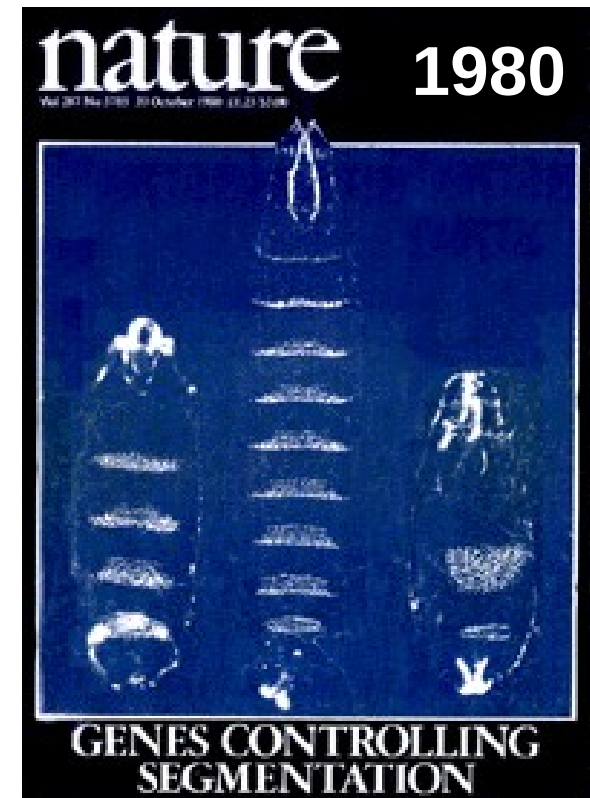
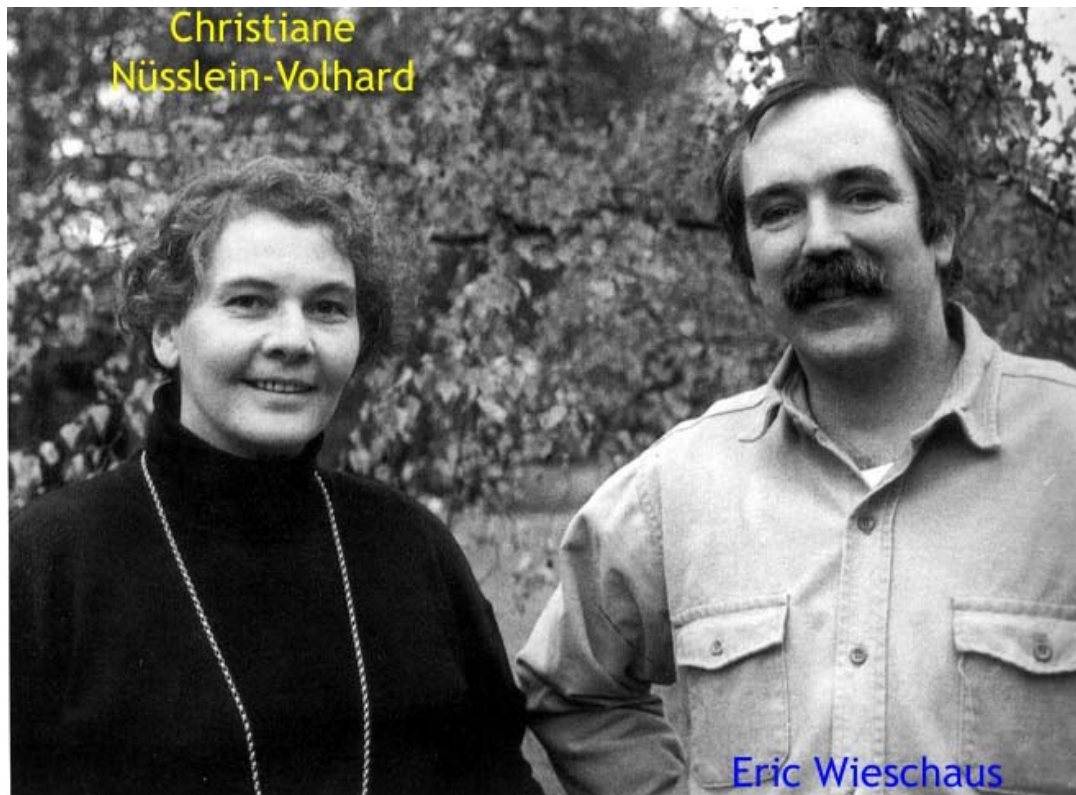
**by C. Nüsslein-Wolhard et E. Wieschaus**

**General principles**

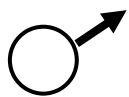
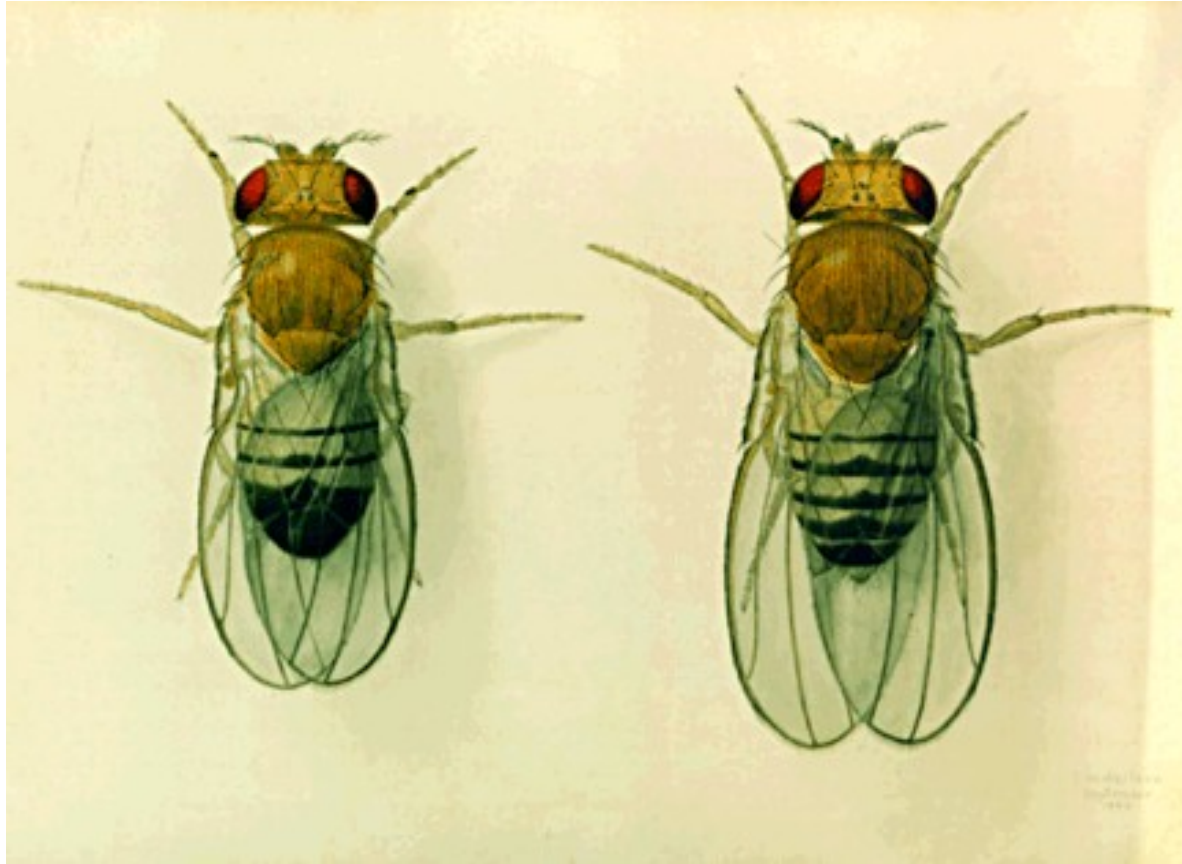
**Other types of screen**



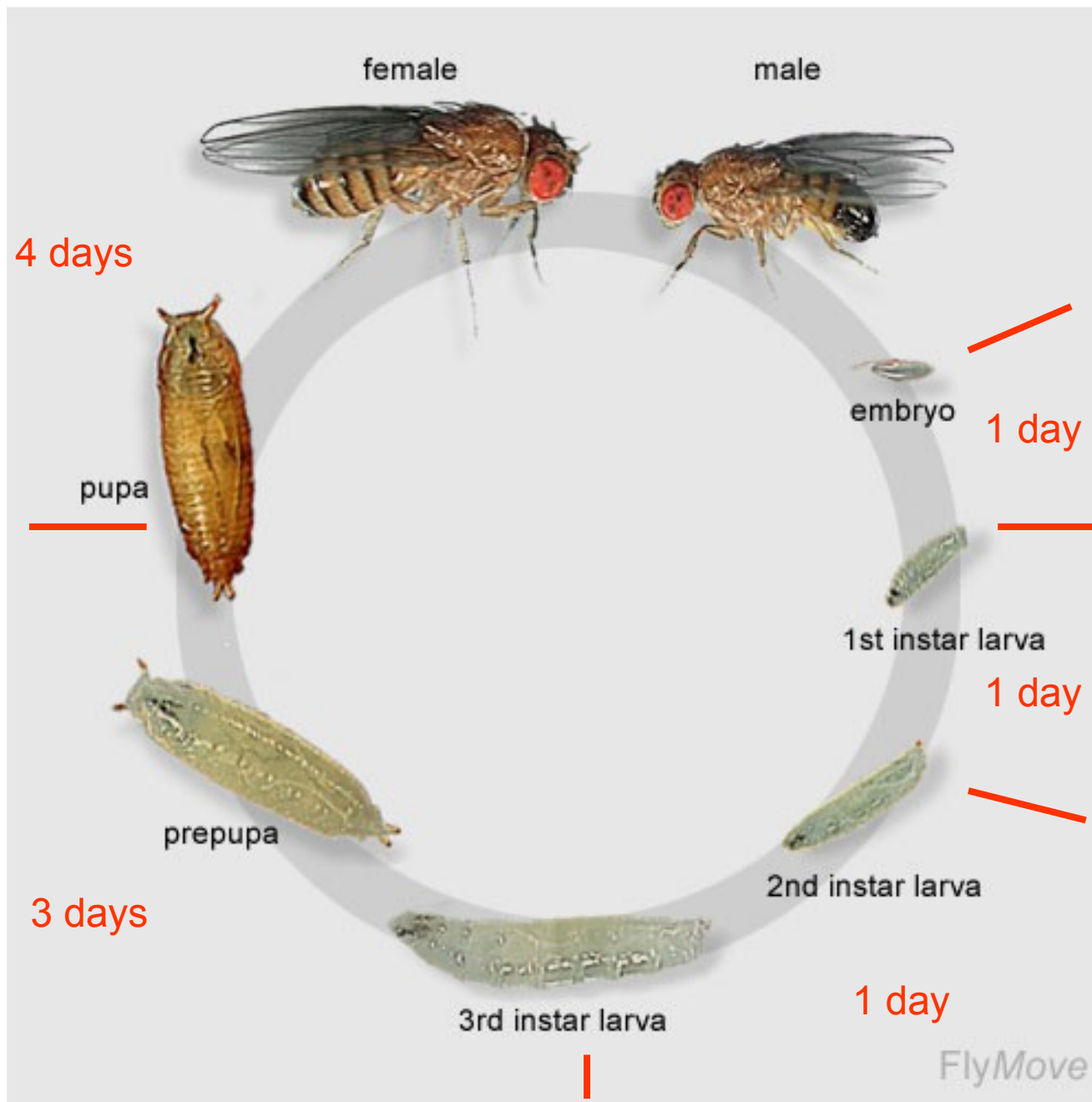
# Historical screen by C. Nüsslein-Volhard et E. Wieschaus



**Nobel Price in Physiology/Medecine in 1995  
with E. Lewis**



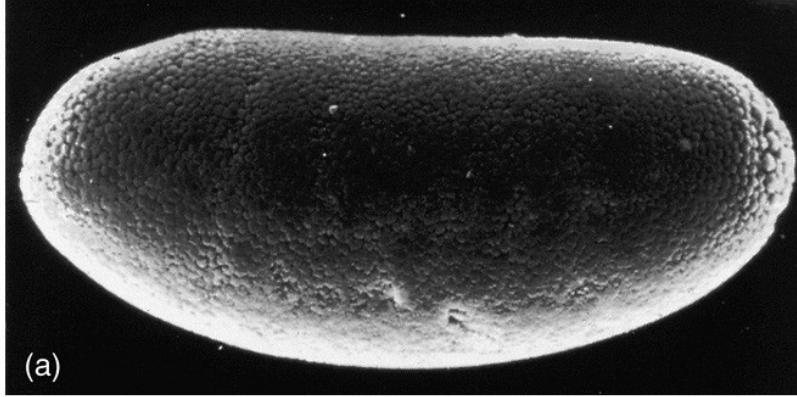
## The life cycle of *Drosophila melanogaster*



One Generation  
=  
10 days

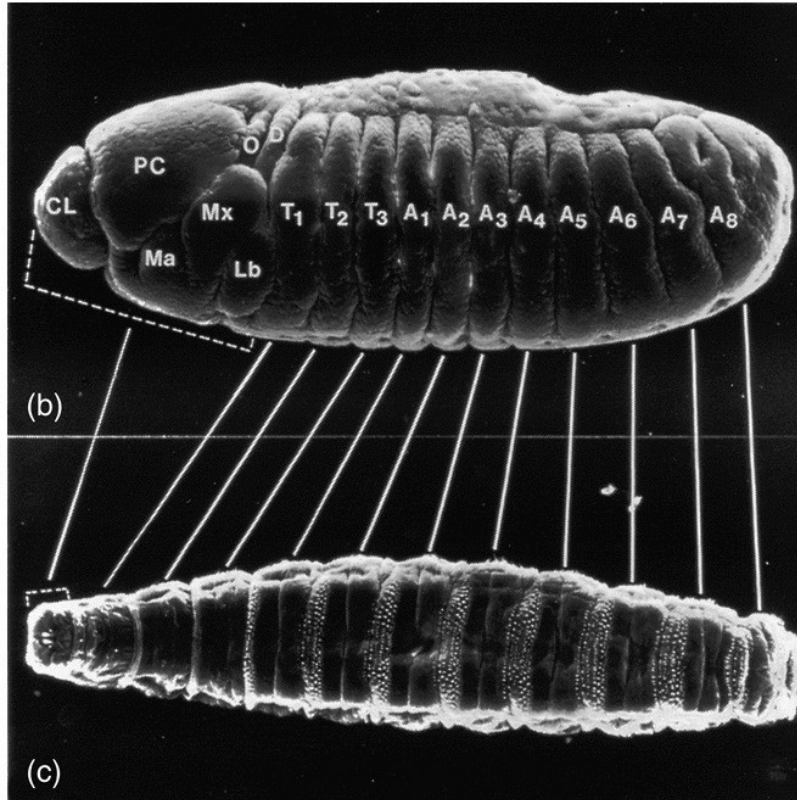
Cellular blastoderm  
stage

3 h



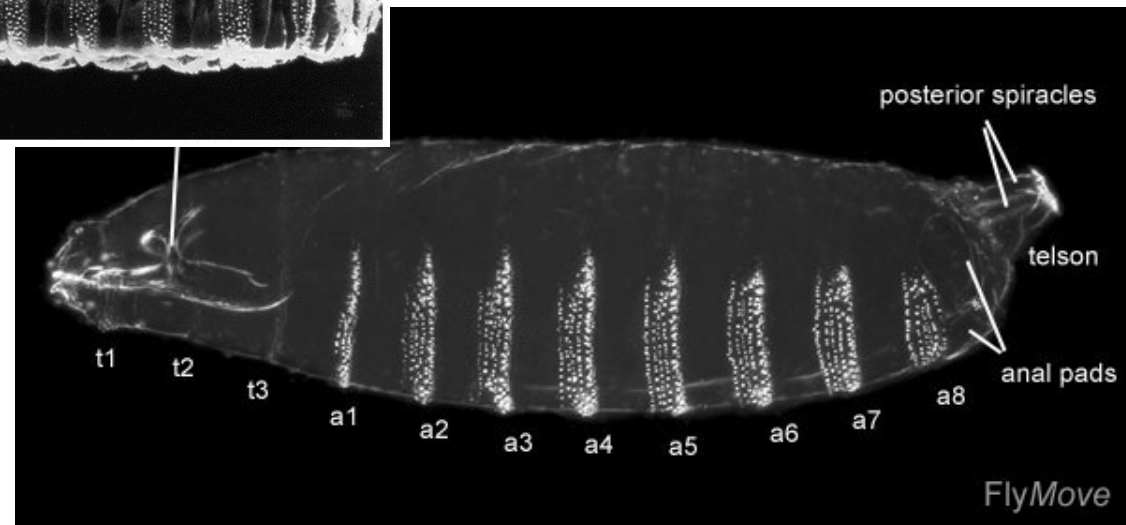
Segmented embryo

10 h



Larva

**Wild-type  
larva**





# General Strategy

Random mutagenesis of the *Drosophila* genome and screen for mutant phenotypes



Identification of the mutated gene

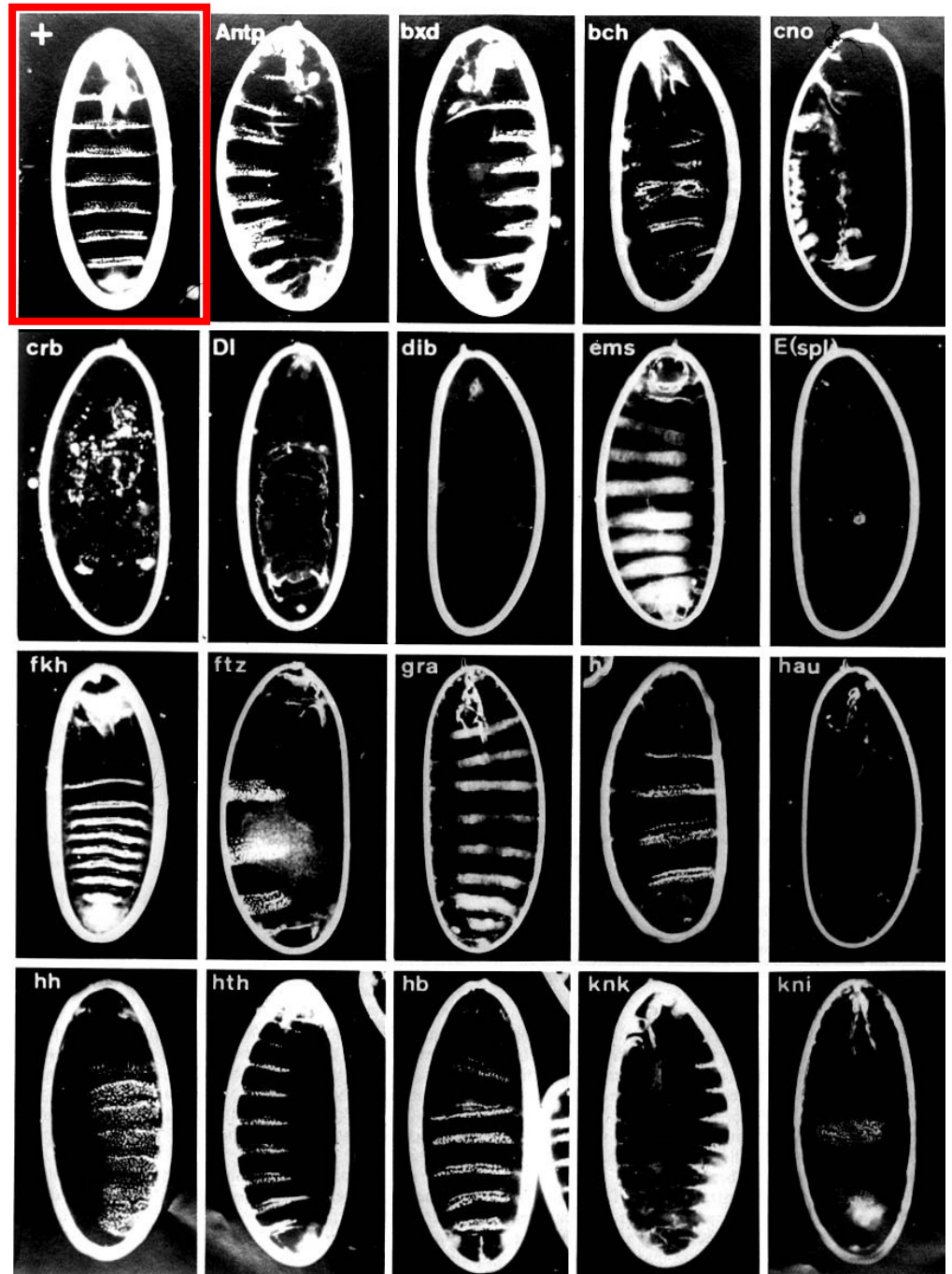


Molecular analysis of the protein function

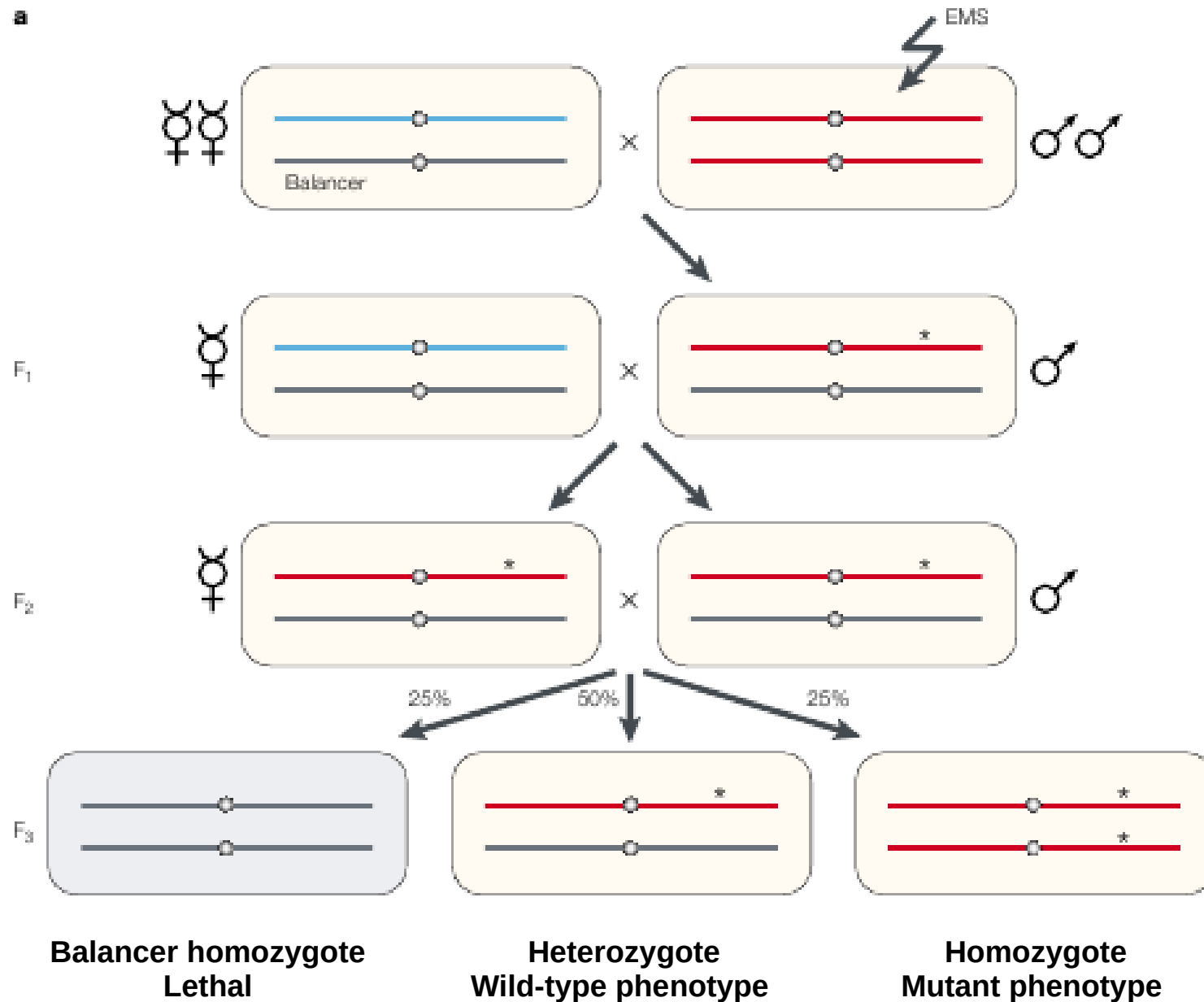
**Wild-type**

**Others= mutants**

**(aberrant position  
/shape of trichomes)**



# Using balancers to screen recessive lethals



# Screen of chromosome 2

Production of 5.764 lines  
including 4.217 homozygote lethal lines

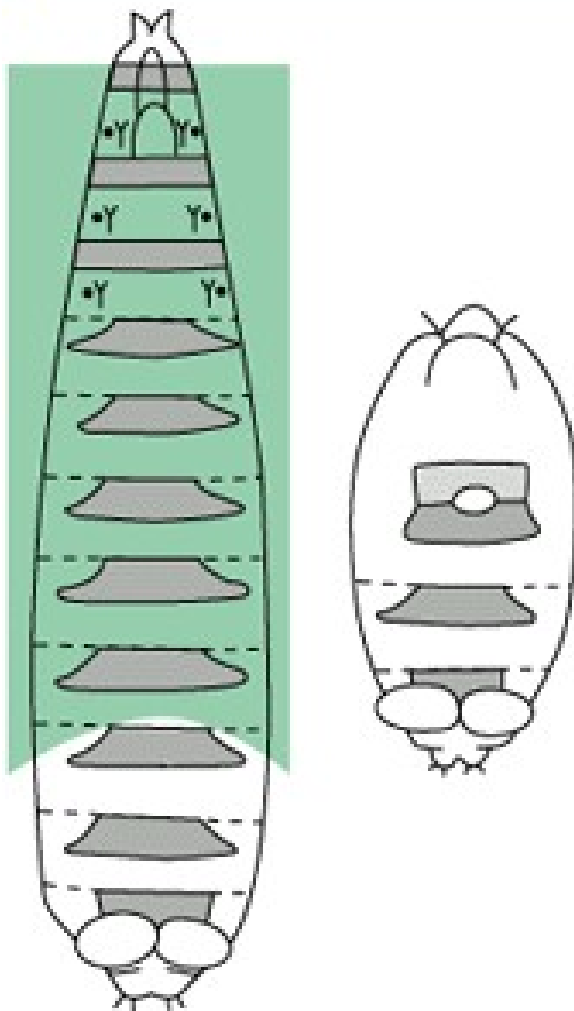
Identification of 7.600 lethal mutations  
including 2.843 mutations causing embryonic lethality  
and 272 mutations embryonic phenotypes

**Complementation test** for mutations with the same phenotype:  
**48 complementation groups** containing on average 5.4 alleles  
13 alleles are complemented by all other mutants

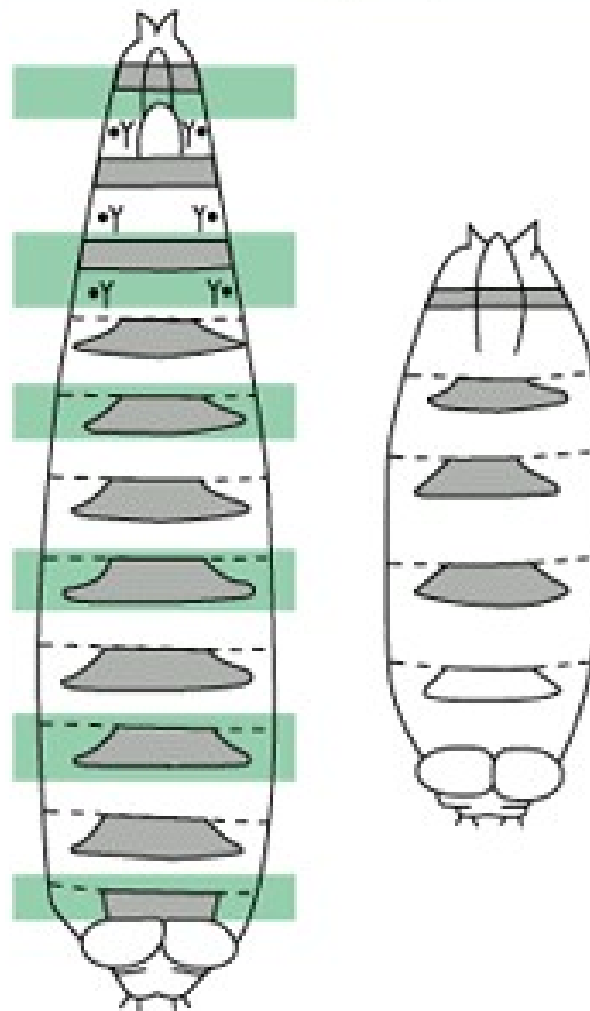
= 61 genes in total



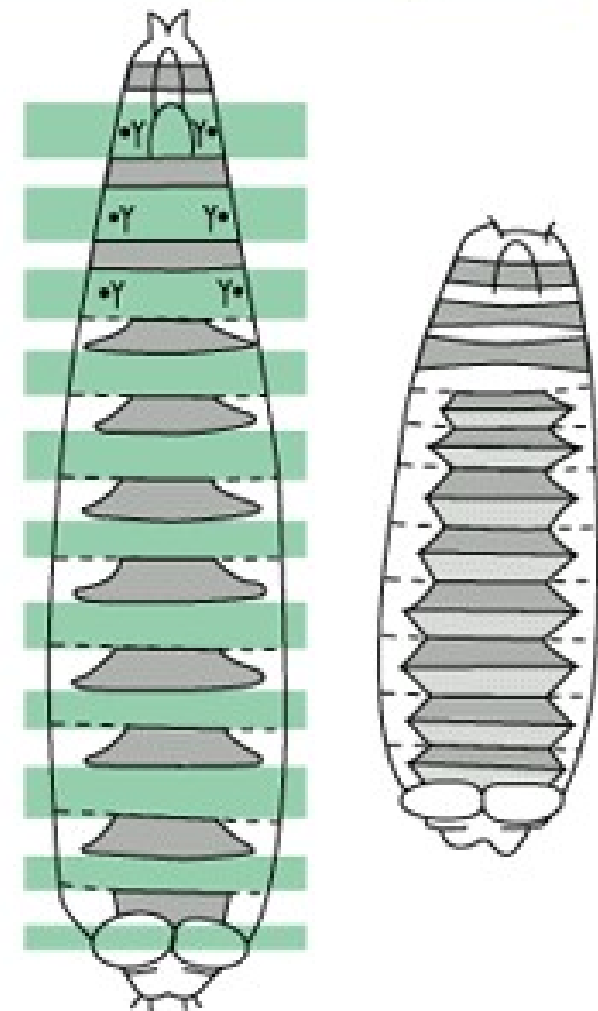
GAP GENE (*Krppel*)



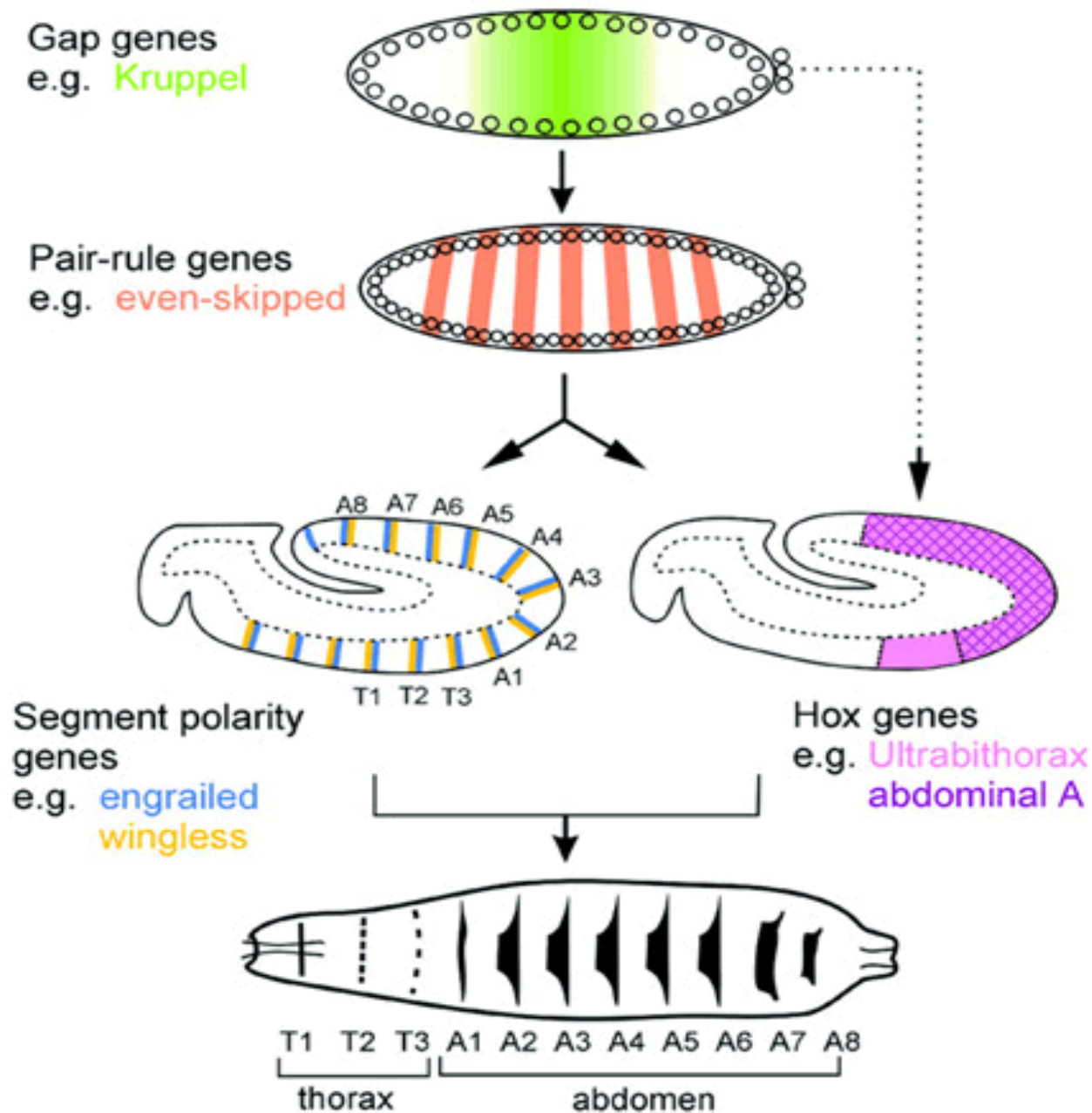
PAIR-RULE GENE (*even-skipped*)



SEGMENT-POLARITY GENE (*gooseberry*)

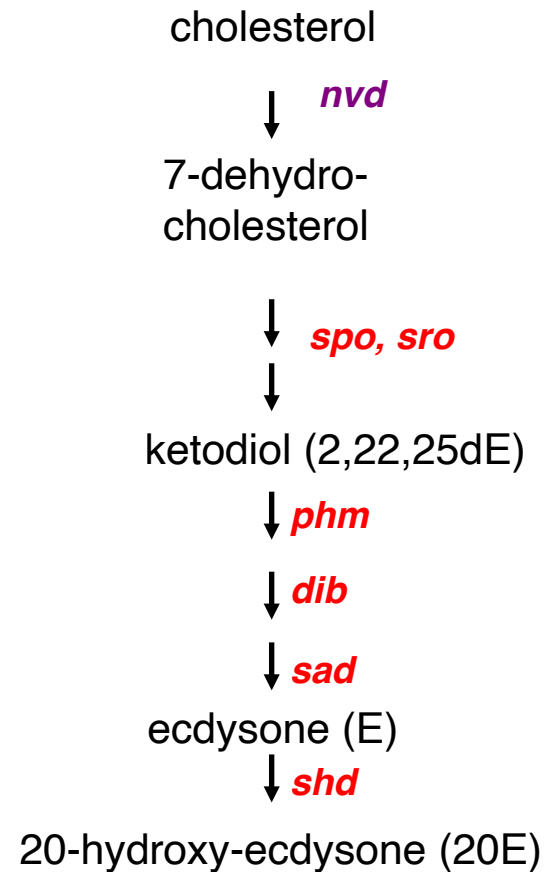
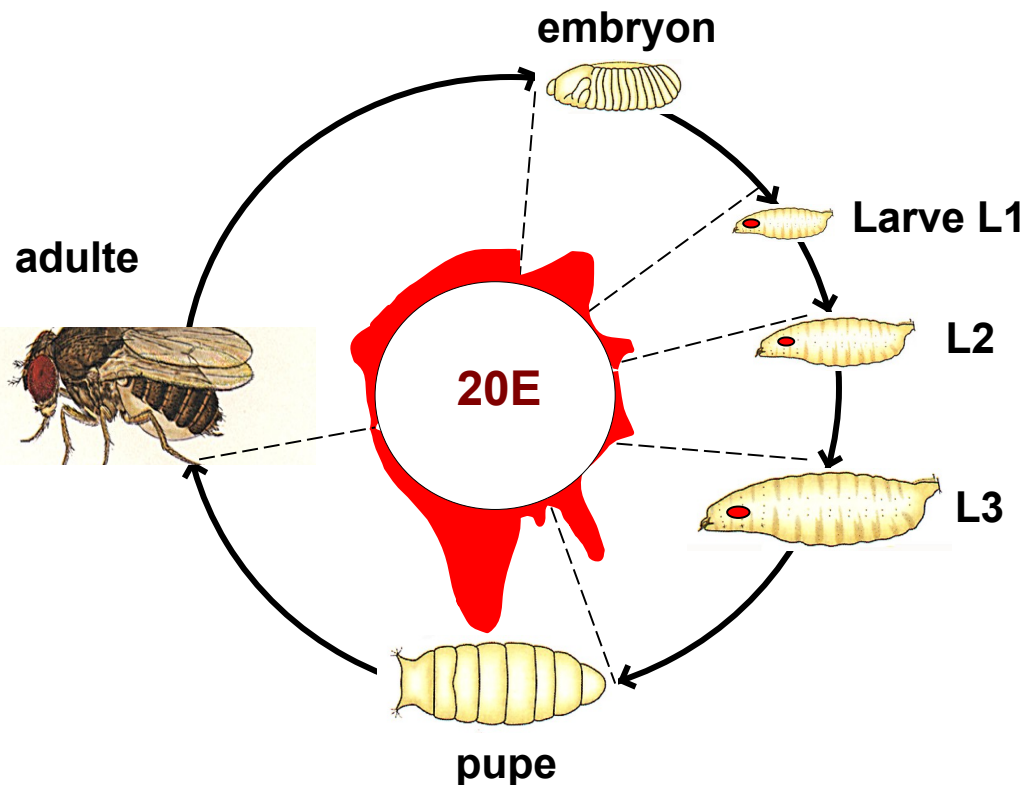
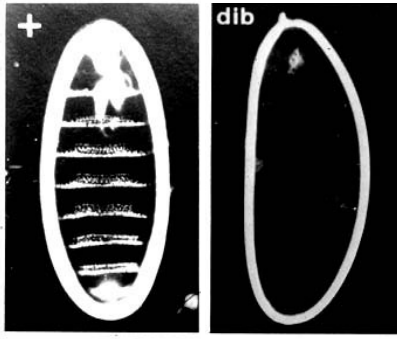


# Embryonic development of Drosophila



# Halloween mutants: steroid hormone biosynthesis

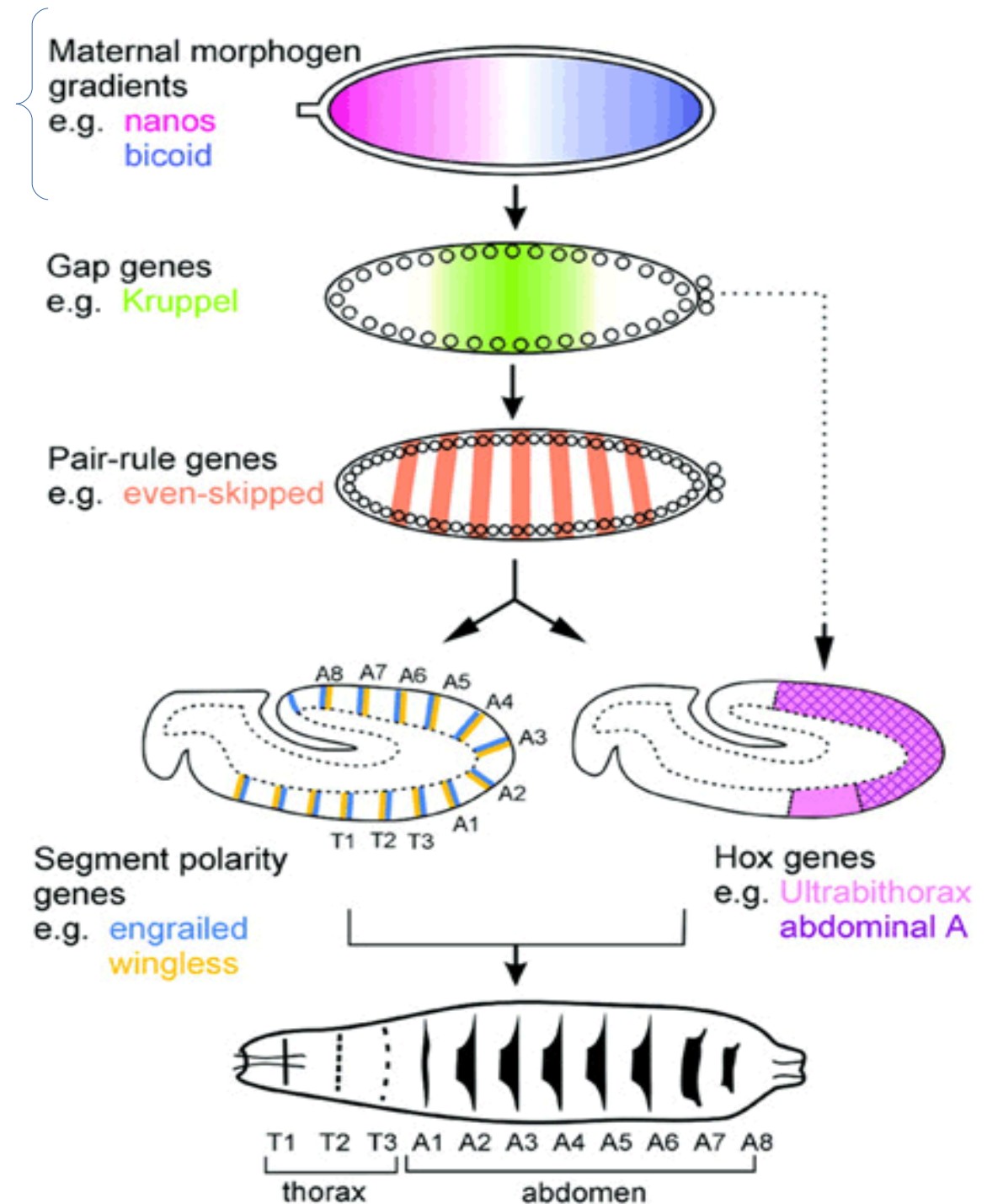
Mutants Halloween



(Chavez et al., 2000; Warren et al., 2002; Petryk et al., 2003; Niwa et al., 2004; Warren et al., 2004; Namiki et al., 2005; Yoshiyama et al., 2006)

# Maternal genes

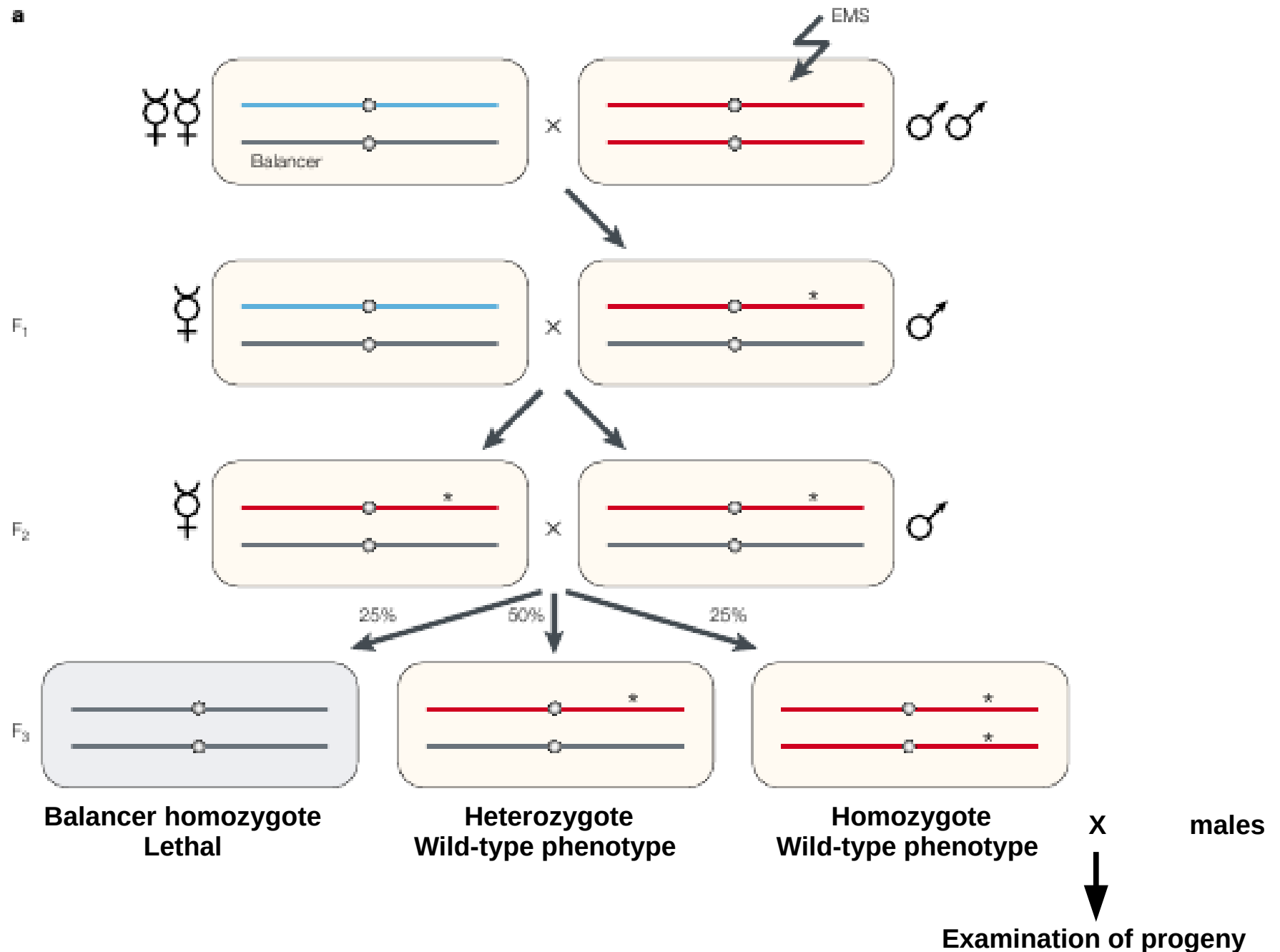
ARNm deposited in the egg before fecundation



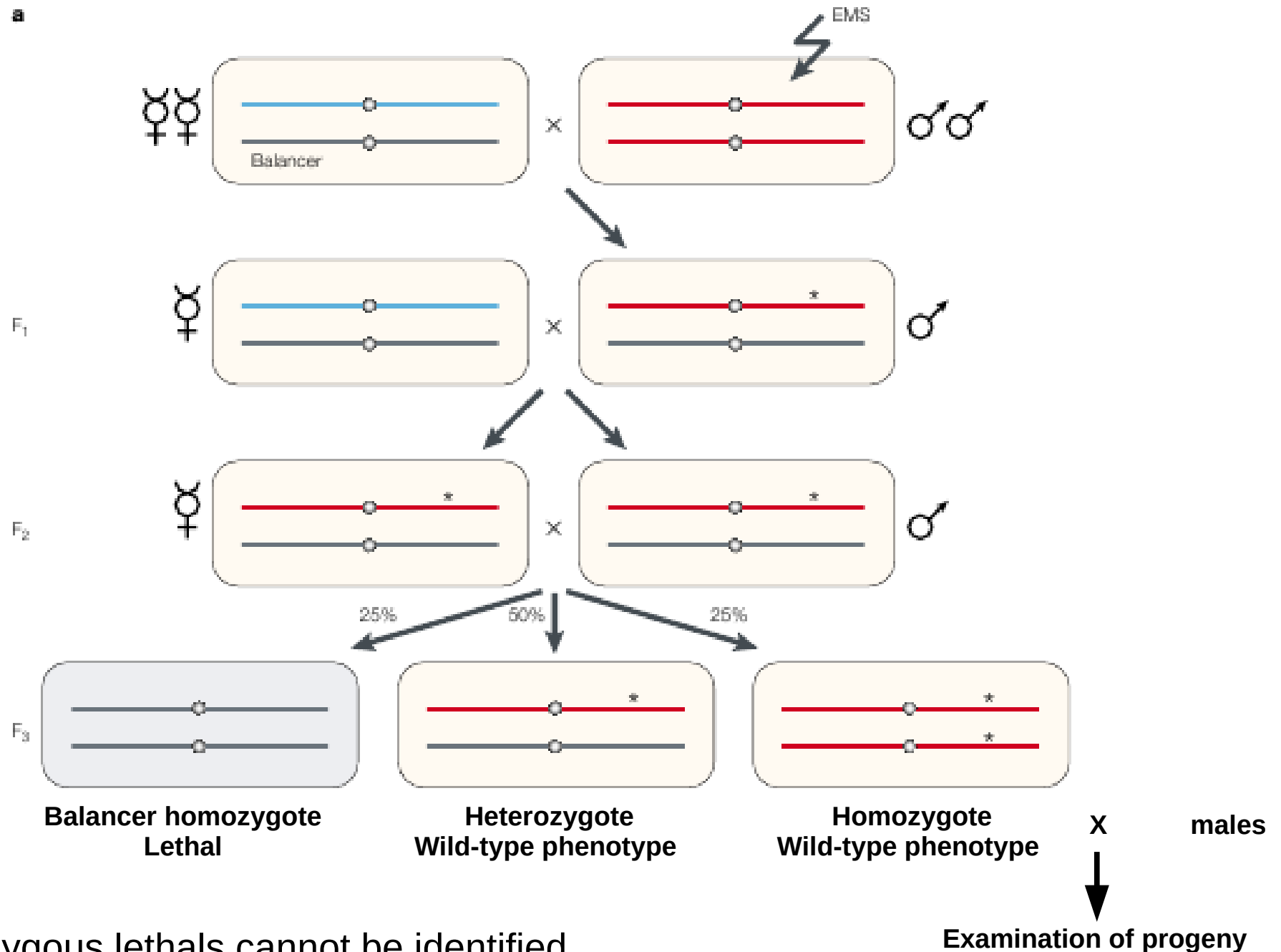


# Screen for maternal effect genes

a



# Screen for maternal effect genes



Homozygous lethals cannot be identified  
(ex : *Fz*, *Dsh*, *Apc*, *Nvd*)

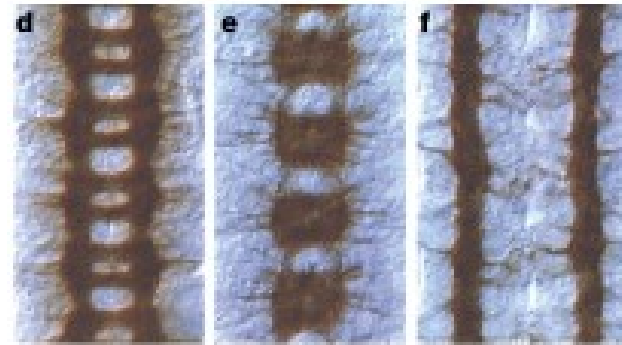
# Certain genes involved in embryonic development were not identified with this screen

Maternal effect genes whose mutation is recessive lethal

Genes involved in the development of internal structures (brain, gut, etc.)

Redundant genes

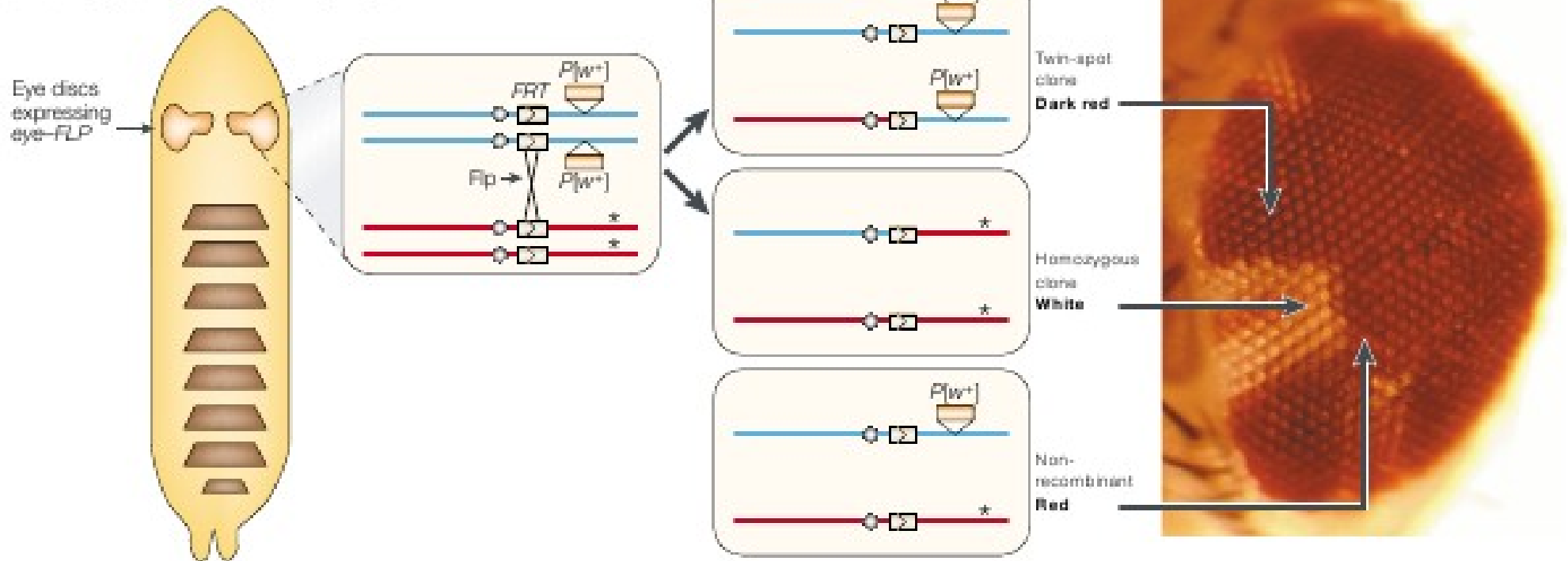
WT *roundabout* *commissureless*



**With such screens, only the first essential function of a gene can be identified.**

# Mitotic clones

**a** Genotype: *w eye-FLP; FRT P(w<sup>+</sup>) / FRT<sup>+</sup>*

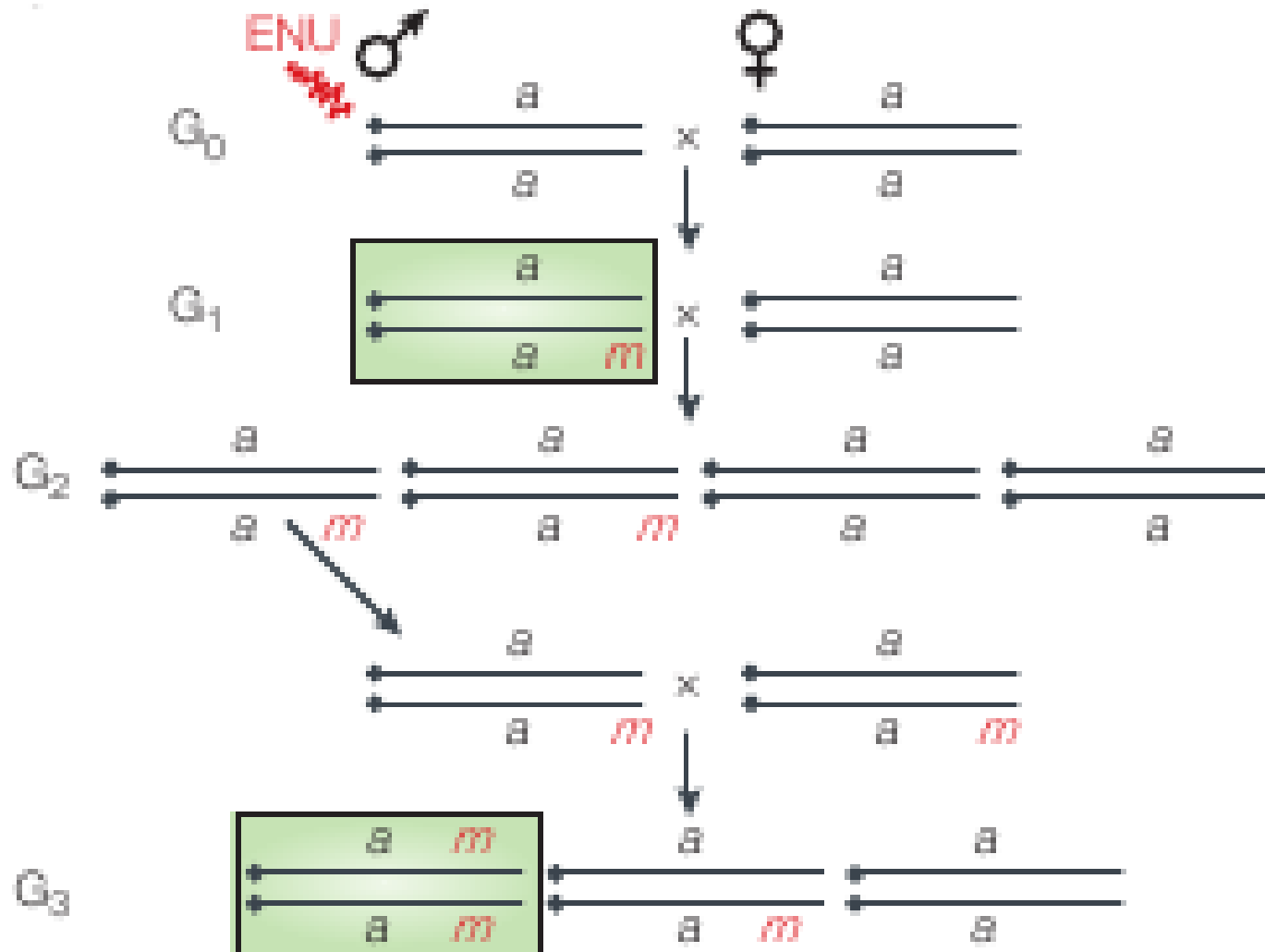


**Flp = Flippase**

**FRT = Flippase Recombination Target**



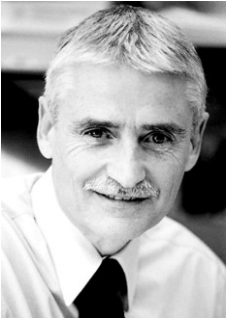
# Screen for suppressors or enhancers



$a/a$  individuals are viable and fertile.

Screen for enhancers or suppressors of the phenotype.

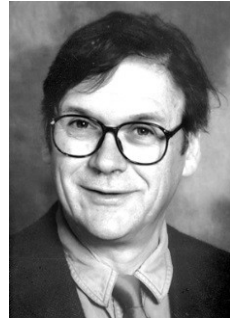
# Three Nobel prizes associated with genetic screens



Leland Hartwell



Paul Nurse



Tim Hunt

Eukaryotic cell division  
(Hartwell et al. 1974; Nurse et al. 1976)

## The Nobel Prize in Physiology or Medicine 2017



Seymour Benzer

Circadian rhythms  
(Konopka and Benzer 1971)



© Nobel Media. Ill. N.  
Elmehed  
Jeffrey C. Hall



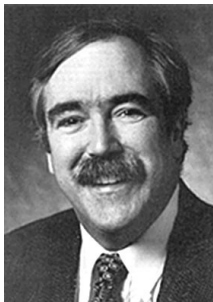
© Nobel Media. Ill. N.  
Elmehed  
Michael Rosbash



© Nobel Media. Ill. N.  
Elmehed  
Michael W. Young



C. Nusslein-Volhard



Eric Wieschaus

Development  
(Lewis 1978; Nusslein-Volhard and Wieschaus 1980)

# **General principles**

**Mutagens**

**Crosses**

**Types of phenotype**

**Identification and validation of causal mutations**

# Mutagens (1)

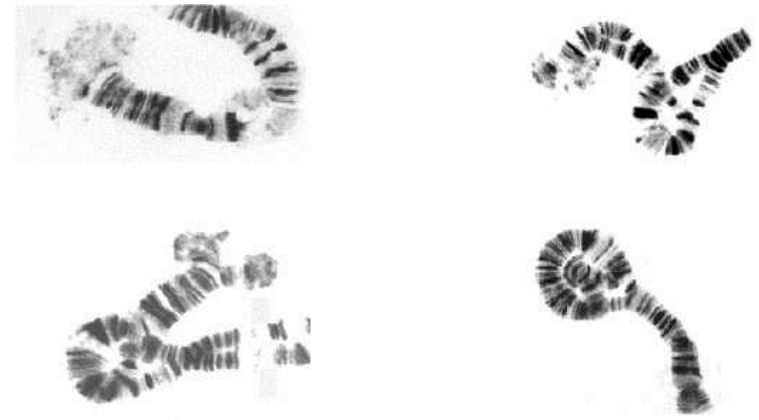
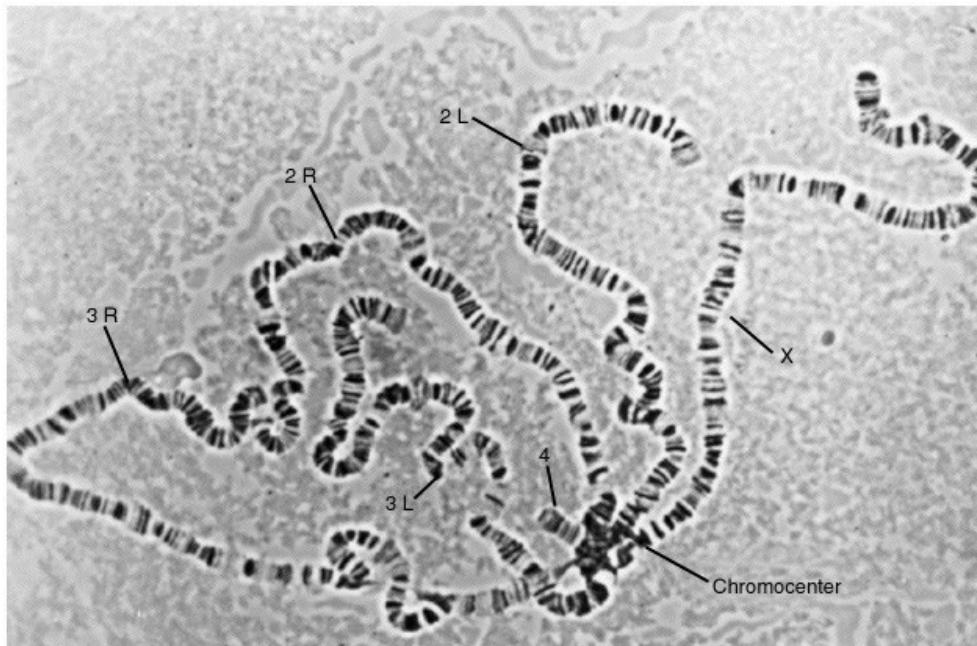
**X rays** : breaks in double-stranded DNA, resulting in large deletions of pieces of chromosome or chromosomal re-arrangements. → good to map by cytological examination of chromosomes, but often not limited to single genes

## **Chemical:**

**Ethylmethane sulfonate (EMS)** : very efficient, alkylation agent ( GC to AT), point mutations

In *Drosophila*, EMS can produce  $\sim 10^{-3}$  mutations per gene  
→ how many mutated genes on average on one chromosome containing 5000 genes?  
→ if 6000 such EMS-treated chromosomes are generated, how many alleles per gene can be expected from the screen?

# X rays



Photomicrographs of polytene chromosomes; name the mutation, if any

# Mutagens (2)

## Chemical:

**Methylmethane sulfonate (MMS)** : agent alkylant, moins efficace que l'EMS pour la drosophile, induit un peu plus de délétions que l'EMS.

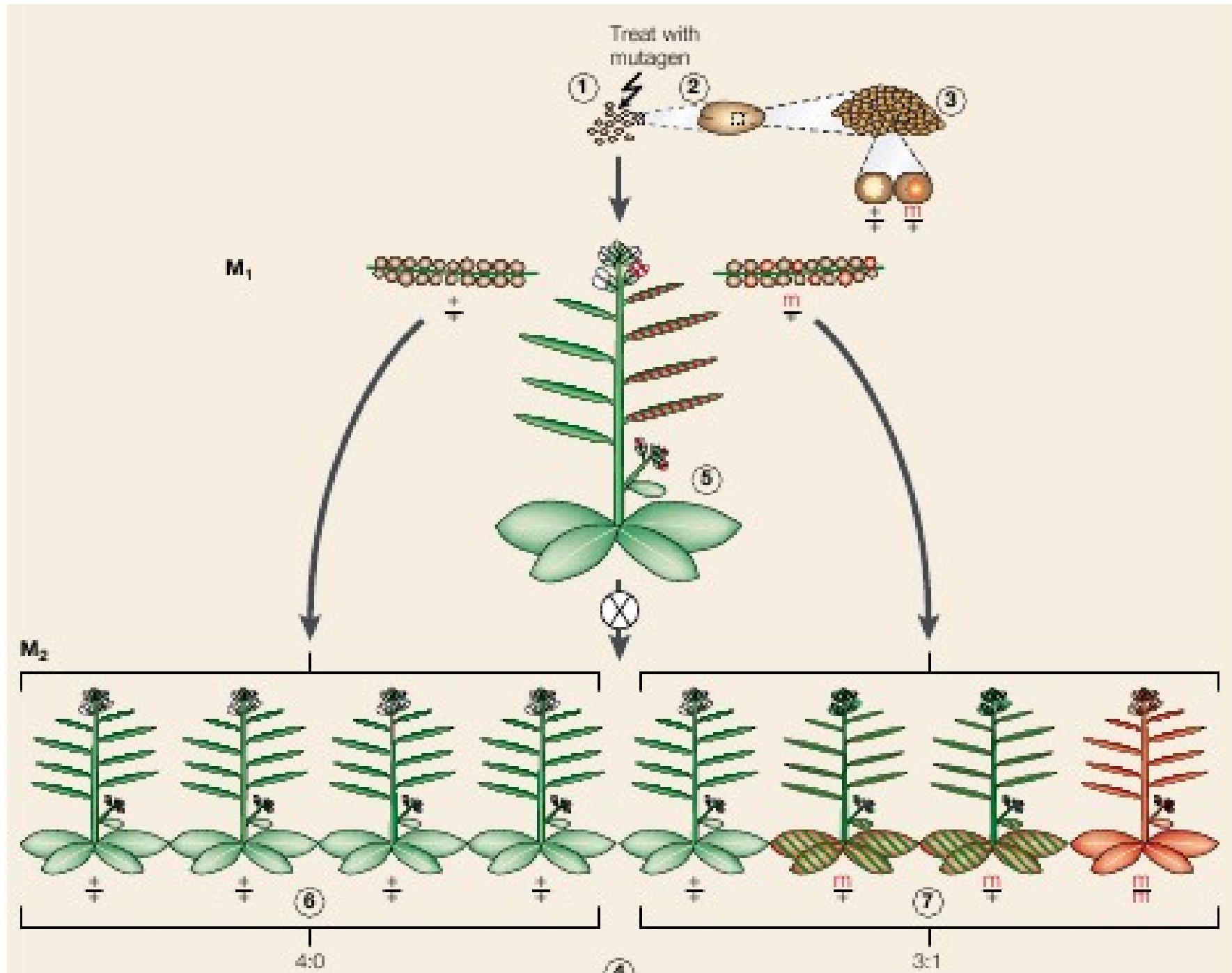
**N-nitroso-N-ethylurea (ENU)** : ethyle oxygen atoms (O2 and O4 of T, AT to GC, O6 of G, GC to AT), fewer aberrations than EMS

**Triethylmelanine (TEM)** : deletions

**Formaldehyde** : deletions



# Chemical mutagens create mosaic individuals



# Mutagens (2)

## Chemical:

**Methylmethane sulfonate (MMS)** : agent alkylant, moins efficace que l'EMS pour la drosophile, induit un peu plus de délétions que l'EMS.

**N-nitroso-N-ethylurea (ENU)** : ethyle oxygen atoms (O2 and O4 of T, AT to GC, O6 of G, GC to AT), fewer aberrations than EMS

**Triethylmelanine (TEM)** : deletions

**Formaldehyde** : deletions

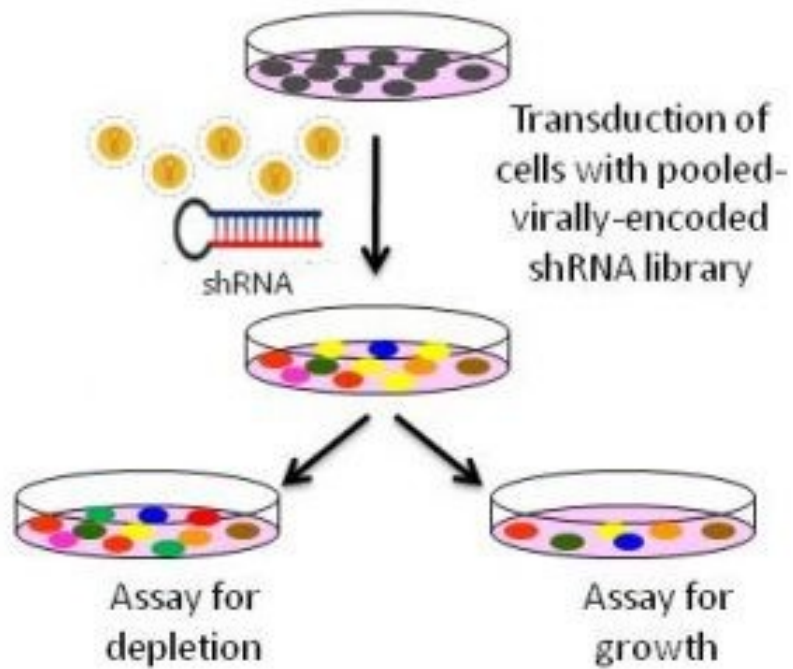
## **Insertational:**

**Transposable elements without transposase**: integrate into the genome, facilitates identification of the mutation

**RNAi**

**CRISPR-Cas-9**

## Pooled screens

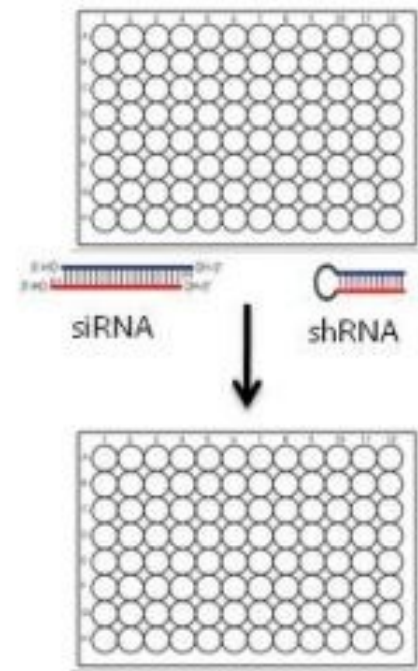


Sequence colonies or analyze barcodes on microarrays



## Arrayed screens

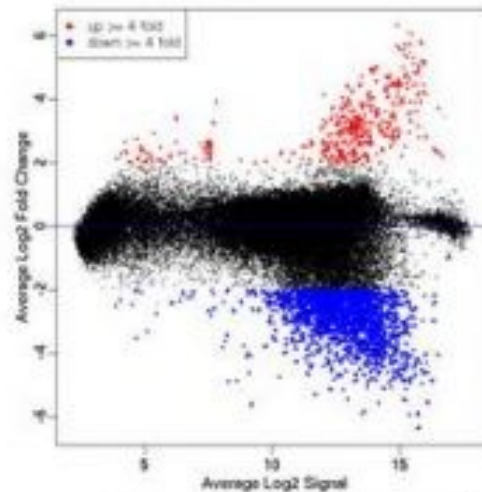
Transfection of cells with RNAi reagents



Assay for phenotype



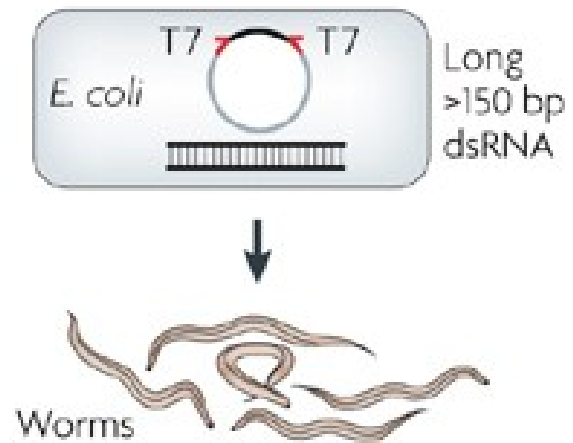
## Selection



Data analysis and "hit"

# RNAi screens

*C. elegans*



*Drosophila*

Long dsRNA  
>100 bp

Bathing



Humans

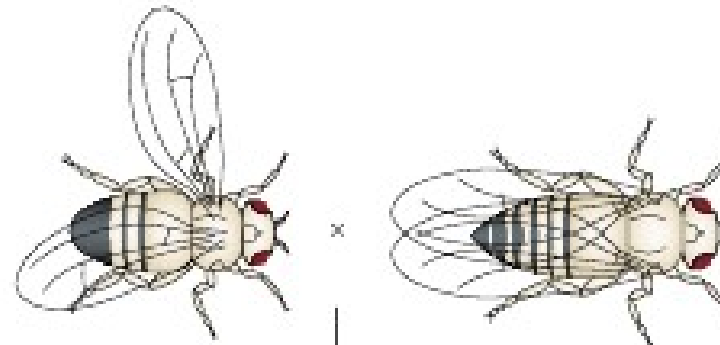
siRNA  
21 bp

Transfection



Enhancer-trap GAL4

UAS-gene X



Genomic enhancer — GAL4  
Tissue-specific expression of GAL4

GAL4  
UAS — gene X  
Transcriptional activation of gene X

# Forward genetics

Genetic screen for a phenotype of interest, identification of the mutated gene etc...

# Reverse genetics

Start from a gene of interest knockout, transgenesis, etc...

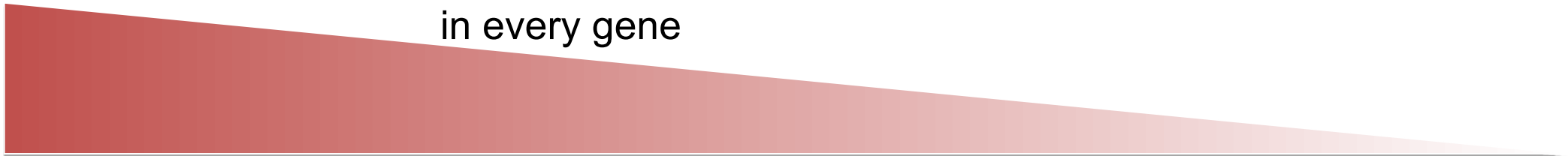
The distinction is fuzzy when one starts from a subset of a library of mutants

Random screens  
no *a priori* bias

Screen of a library  
with mutants  
in every gene

Screen  
of a subset

Study  
of a single gene



# Crosses

Since chemical mutagens create mosaic individuals, the progeny must be screened

F1 screen: screen for suppressors and enhancers

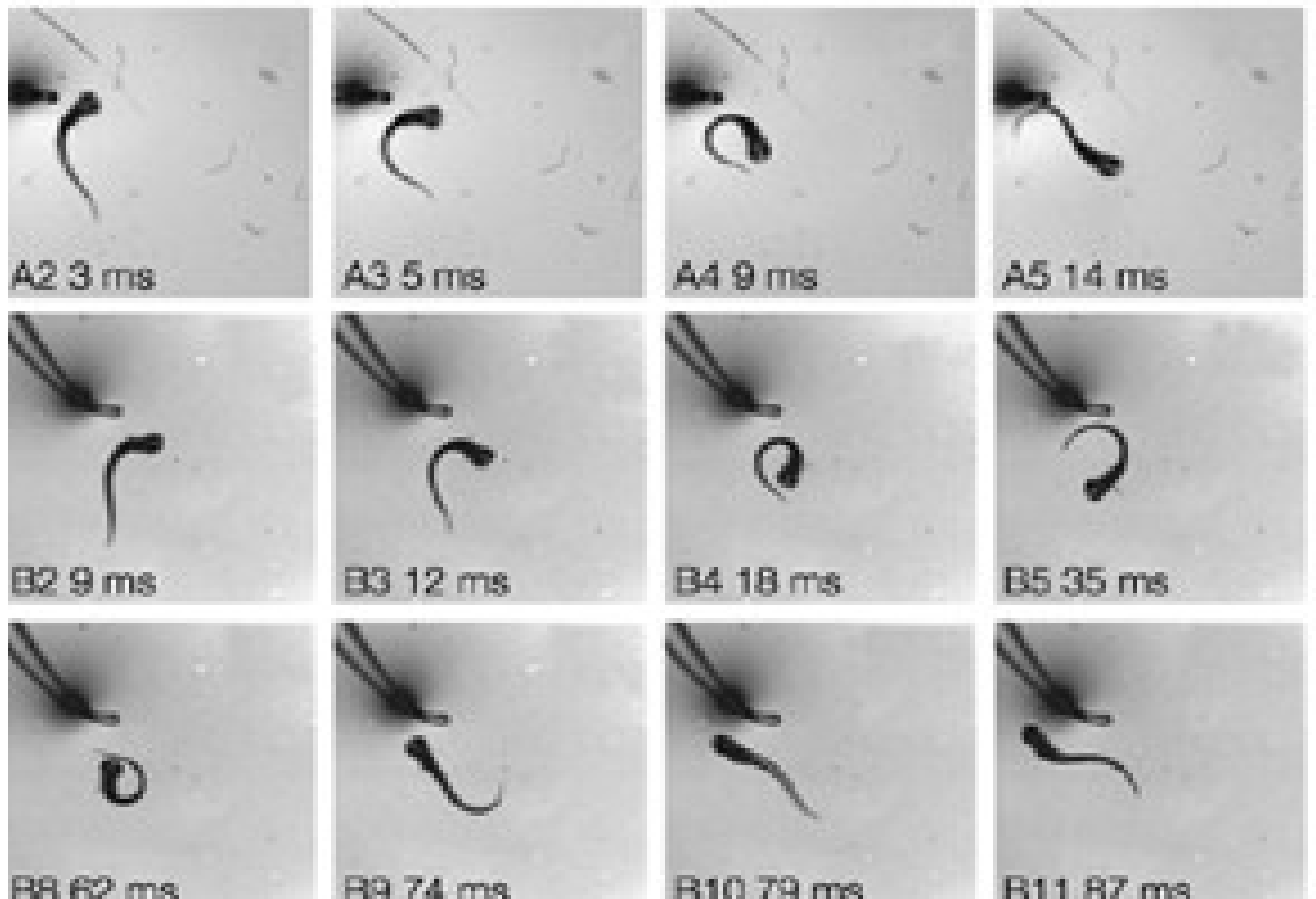
F2 screen: screen for recessive mutations

F3 screen: screen for maternal effect genes



# Phenotypes

Morphology, Physiology, Behavior



*space cadet* Zebrafish mutant

# Phenotypes

Direct observation

WT



*agamous-1*

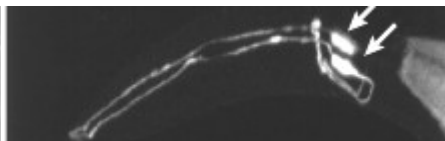


Staining (GFP, antibodies)

WT



mutant 1

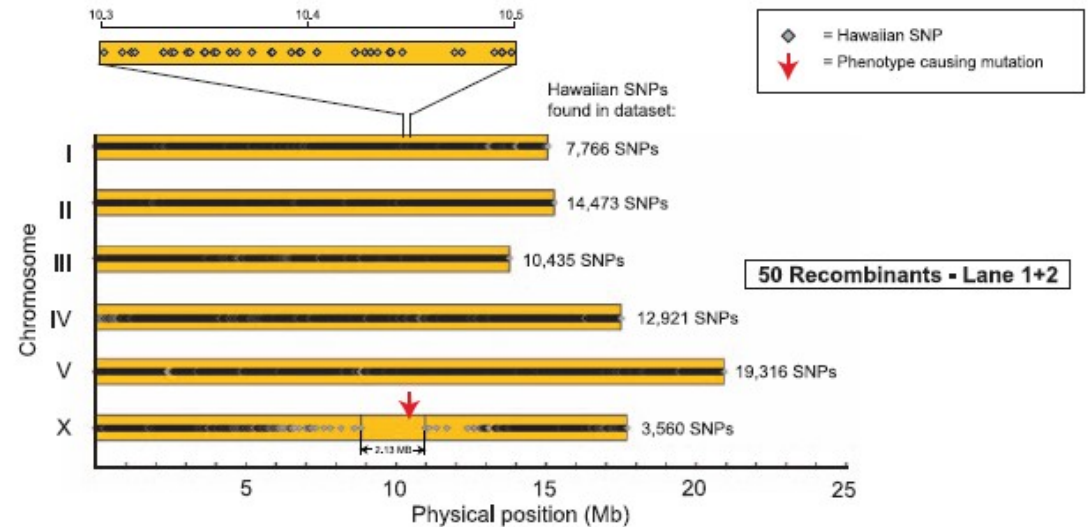
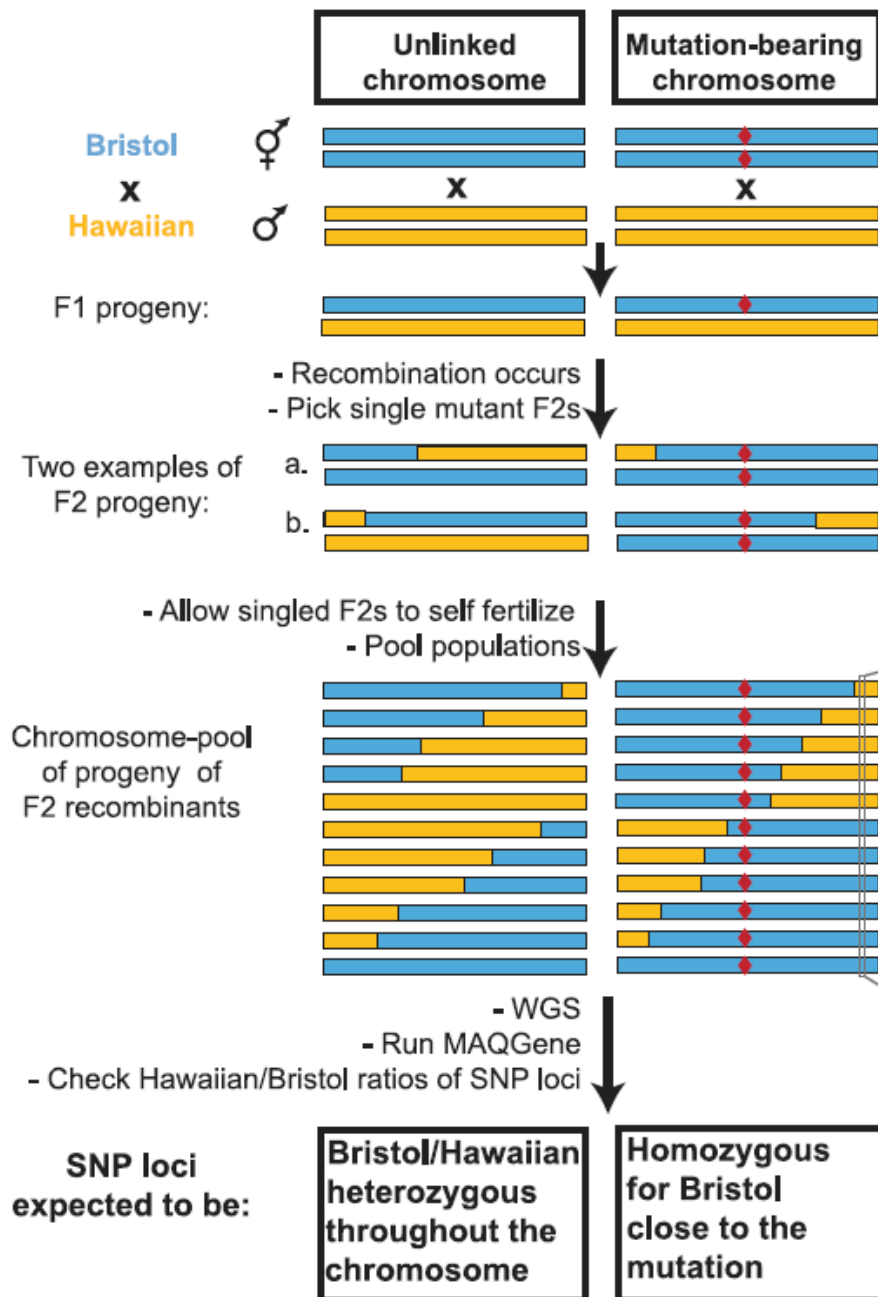


mutant 2



*str2::GFP*

# Identification of the mutation



Requires nucleotide divergence and many recombinants

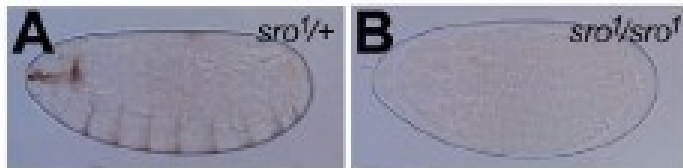
# Identification of the mutation

Once a small region is identified

Complementation test with deletions/mutants already available

Analysis of candidate gene expression

**Rescue of the mutant phenotype with transgenes**



Niwa et al. (2010) Development 137, 1991

**Table 1. *sro*<sup>1</sup> lethality was rescued by *sro/nm-g* overexpression**

Genotype	Number of adults
<i>+/+; 2-286-GAL4 sro<sup>1</sup>/+ sro<sup>1</sup></i>	0 (308)
<i>UAS-sro/+; sro<sup>1</sup>/sro<sup>1</sup></i>	0 (170)
<i>UAS-nm-g/+; sro<sup>1</sup>/sro<sup>1</sup></i>	0 (138)
<i>UAS-sro/+; 2-286-GAL4 sro<sup>1</sup>/+ sro<sup>1</sup></i>	128 (286)
<i>UAS-nm-g/+; 2-286-GAL4 sro<sup>1</sup>/+ sro<sup>1</sup></i>	57 (270)

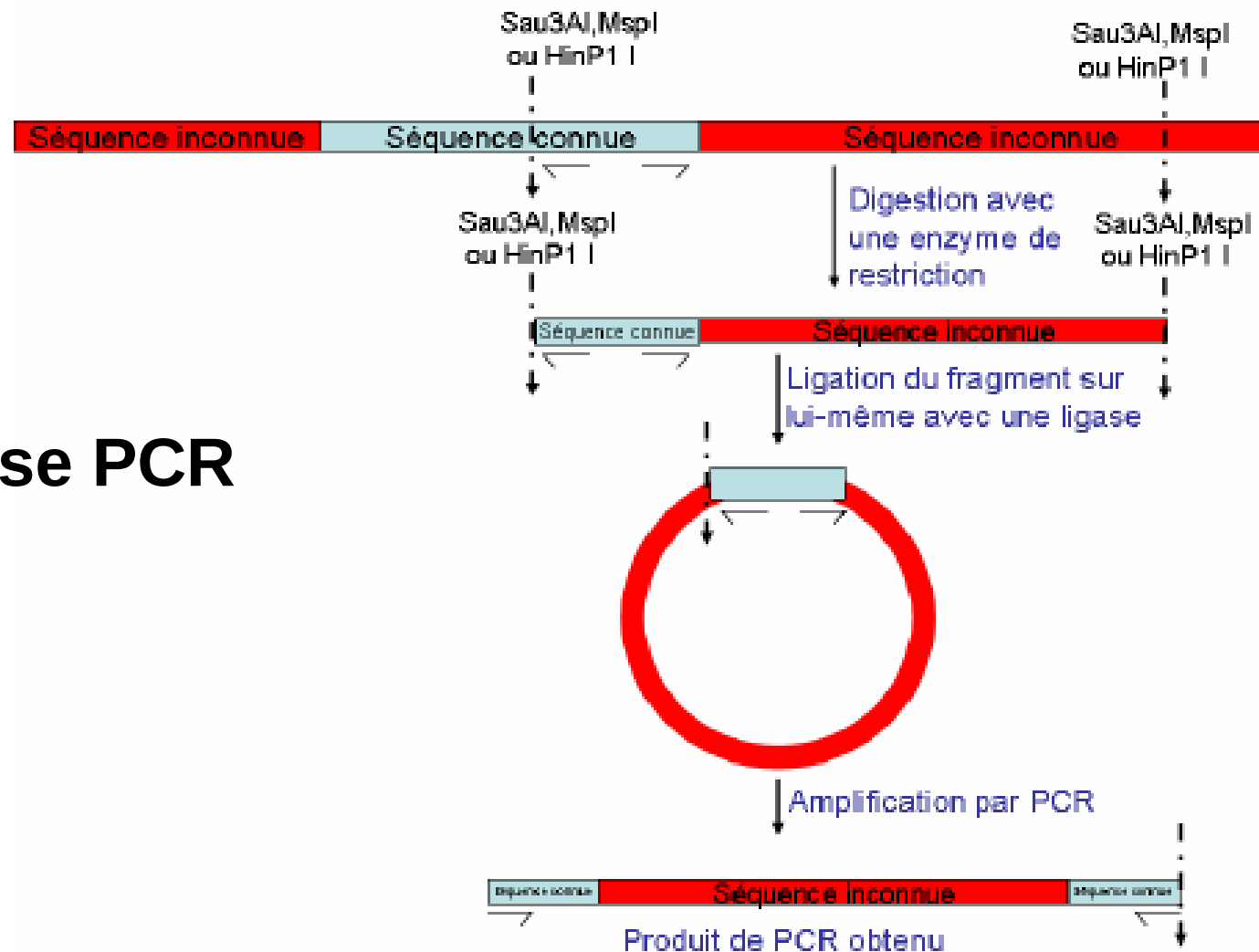
The numbers of viable adults were scored. Parentheses indicate the number of viable progeny with the presence of balancer markers from the parental strains.

Sequencing and search for mutations (nonsense, deletions, etc.)

# Identification of the mutation

For transposable elements

## Inverse PCR



# Other types of screens

## Gene expression screens

RNAseq

In situ hybridization of all genes

## Screen of DNA sequences

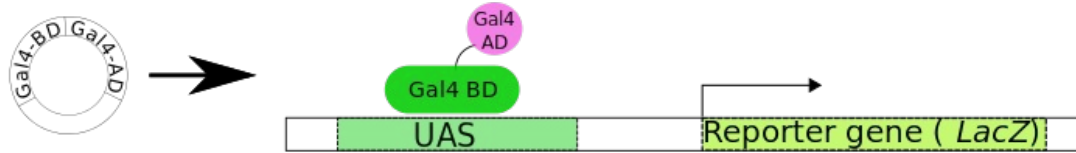
Library with all the genes coding for transcription factors

Two-hybrid screen

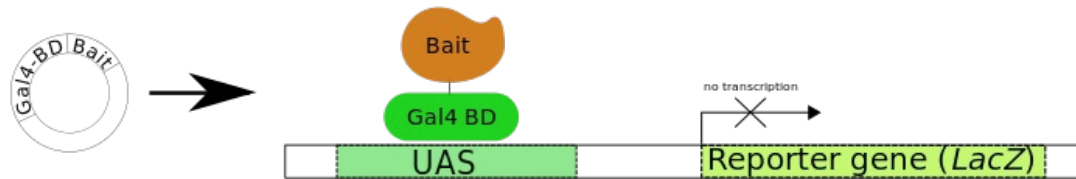
etc.



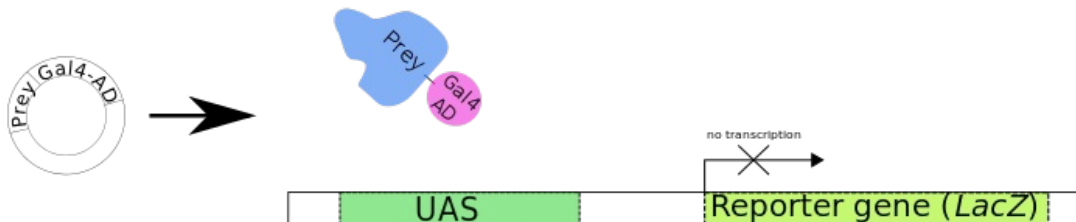
# Yeast two-hybrid screen



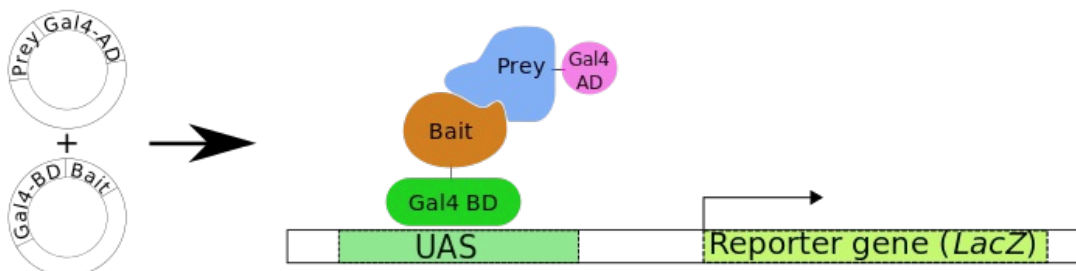
A. Regular transcription of the reporter gene



B. One fusion protein only (Gal4-BD + Bait) - no transcription



C. One fusion protein only (Gal4-AD + Prey) - no transcription



D. Two fusion proteins with interacting Bait and Prey

# **From laboratory to “real-life” data**

---

# Knock out



# Natural variation

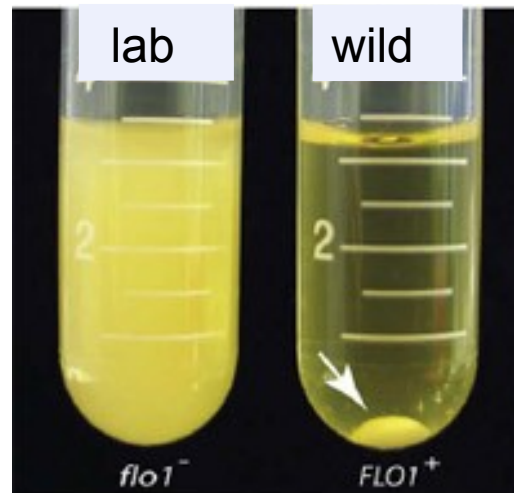


# Domestication of laboratory strains

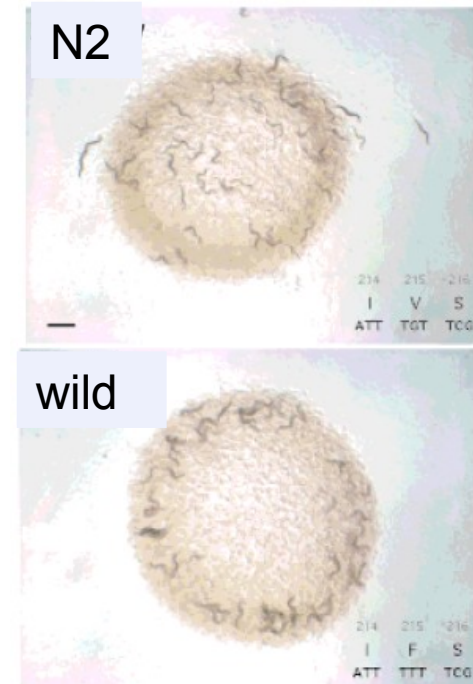
*Arabidopsis thaliana*



*Saccharomyces cerevisiae*



*Caenorhabditis elegans*



Domestication of laboratory strains  
results in extreme phenotypic values  
for many traits:  
artificial selection and pleiotropy

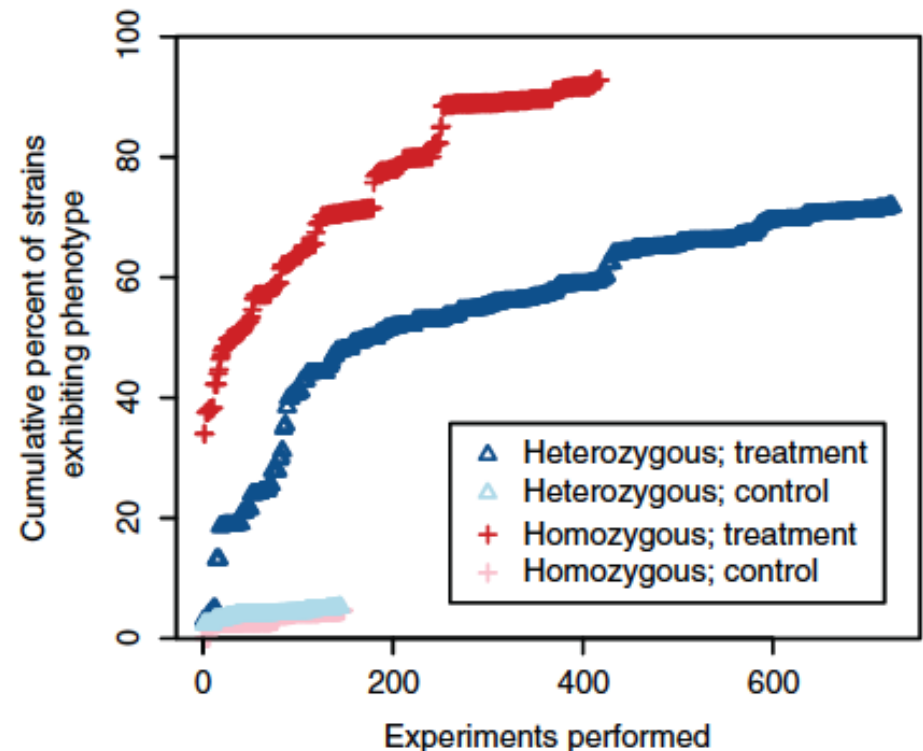
# Choice of laboratory environment

ca. 10-20 years ago: surprise at not finding phenotypes in gene knockouts

## The Chemical Genomic Portrait of Yeast: Uncovering a Phenotype for All Genes

Maureen E. Hillenmeyer, *et al.*  
*Science* **320**, 362 (2008);

1144 growth environments  
for *S. cerevisiae*



# **Genetic Screens**

## **Laboratory mutations**

- Not in nature
  - Extreme effects
  - Would likely be lost under selection
  - Must be induced
- 
- Interrogates (nearly) all regions
  - Readily cloned
  - Strong effects

# **Linkage/Association**

## **mapping**

## **Natural mutations**

- Representative of nature
  - Variants with small effects
  - Sustained under selection
  - Readily available
- 
- Interrogates only variable regions
  - Difficult to map
  - Small effects



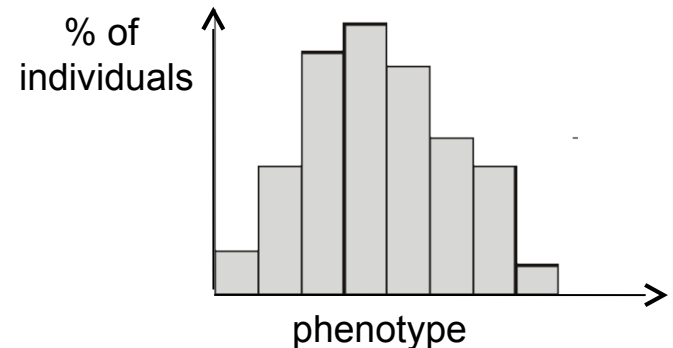
# Quantitative genetics

---

# Quantitative genetics

- If to each genotype corresponds a distribution of phenotypes  
= variable expressivity  
the character itself is quantitative

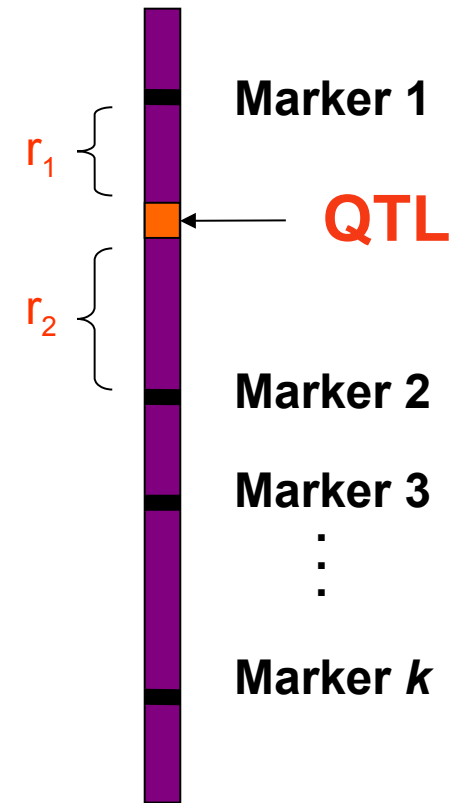
and/or



- If the variation of many genes is involved in the phenotypic difference between two strains/individuals  
the segregation of the character is quantitative

# Quantitative Trait Loci (QTL) mapping

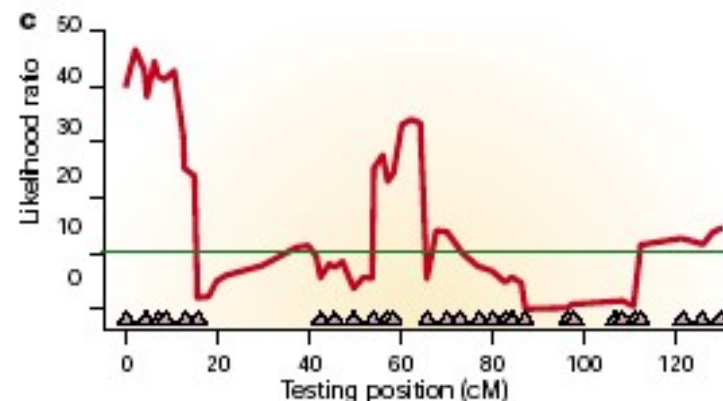
- QTL are specific **genetic loci** that affect quantitative traits.
- QTL can be detected by markers that are linked with it.



## Two goals:

Identify the location of the QTL

Estimate the genetic effects of the QTL



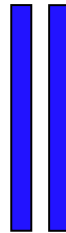
# Epigenetics

---



Wild-type

*Linaria vulgaris*



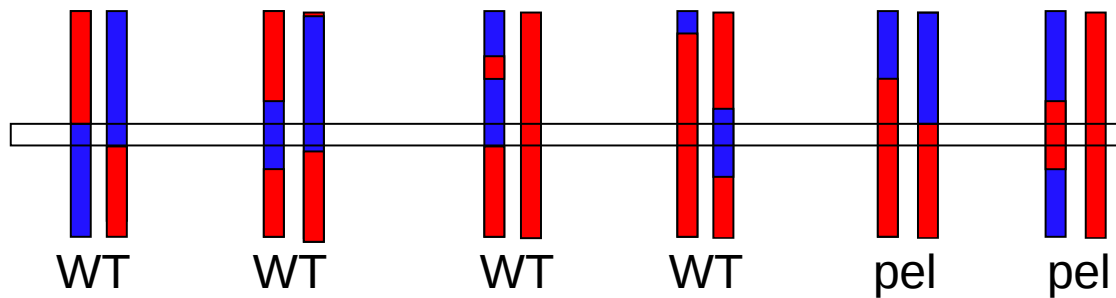
X



Peloric



X

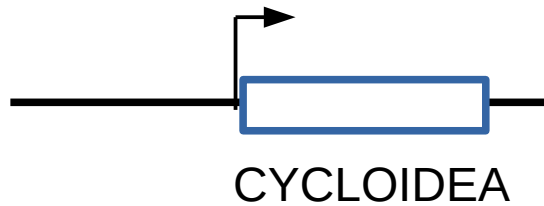


Cubas 1999 Nature

# An epimutation



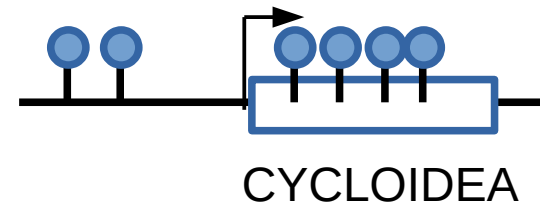
**Wild-type**



Presence of  
CYCLOIDEA  
proteins



**Peloric**



Methylated DNA

Absence of  
CYCLOIDEA  
proteins

# Noise





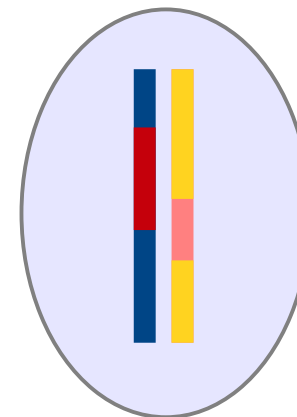
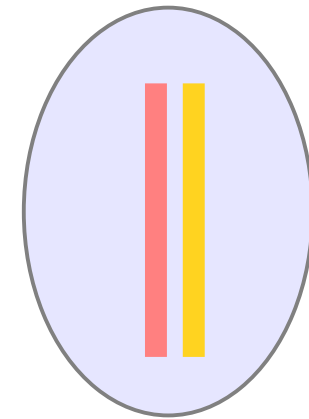
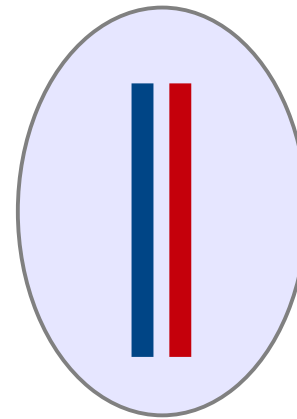
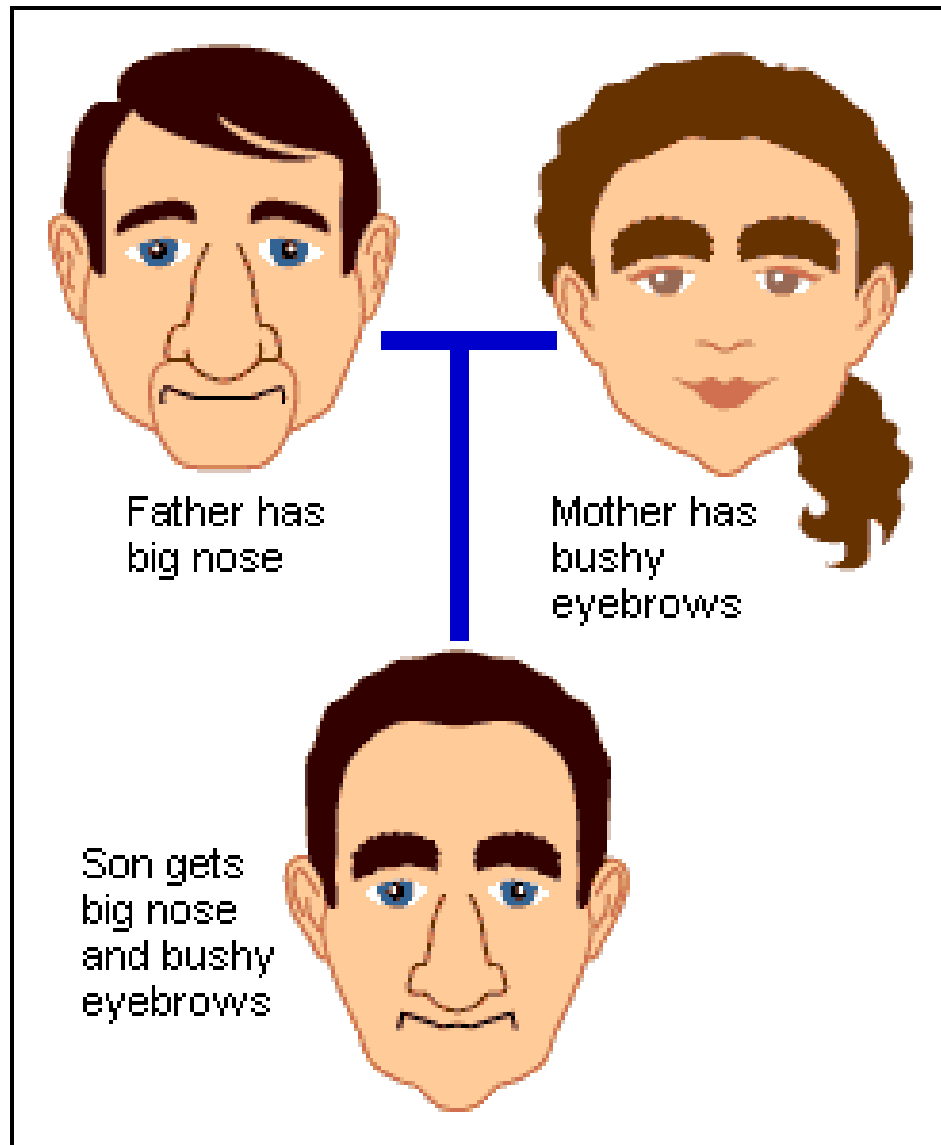
# **Various concepts of chance/randomness in biology**

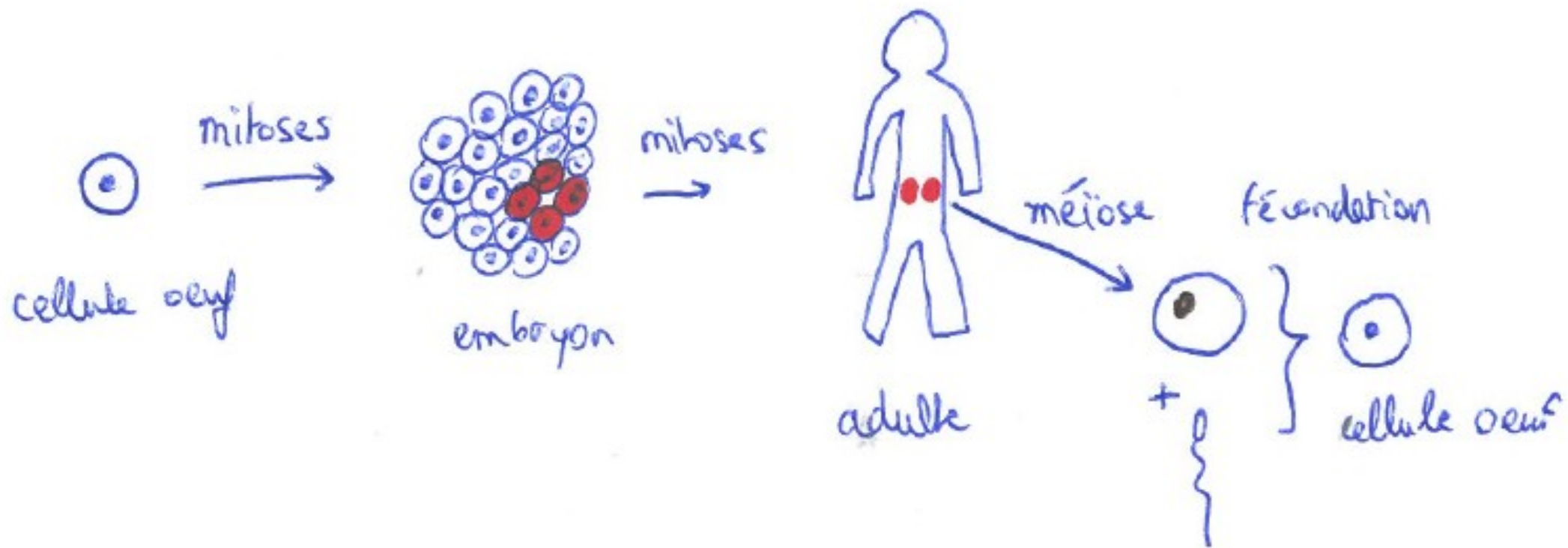
Are not explained within the framework of our current theory  
(no theory, initial conditions not known with sufficient precision,  
or because calculations are too complex)

Cannot be predicted to occur: probabilistic events

No finality/purpose: an end is achieved without having been the  
cause of the accomplishment of the effect

# Assortment of chromosomes from father and mother

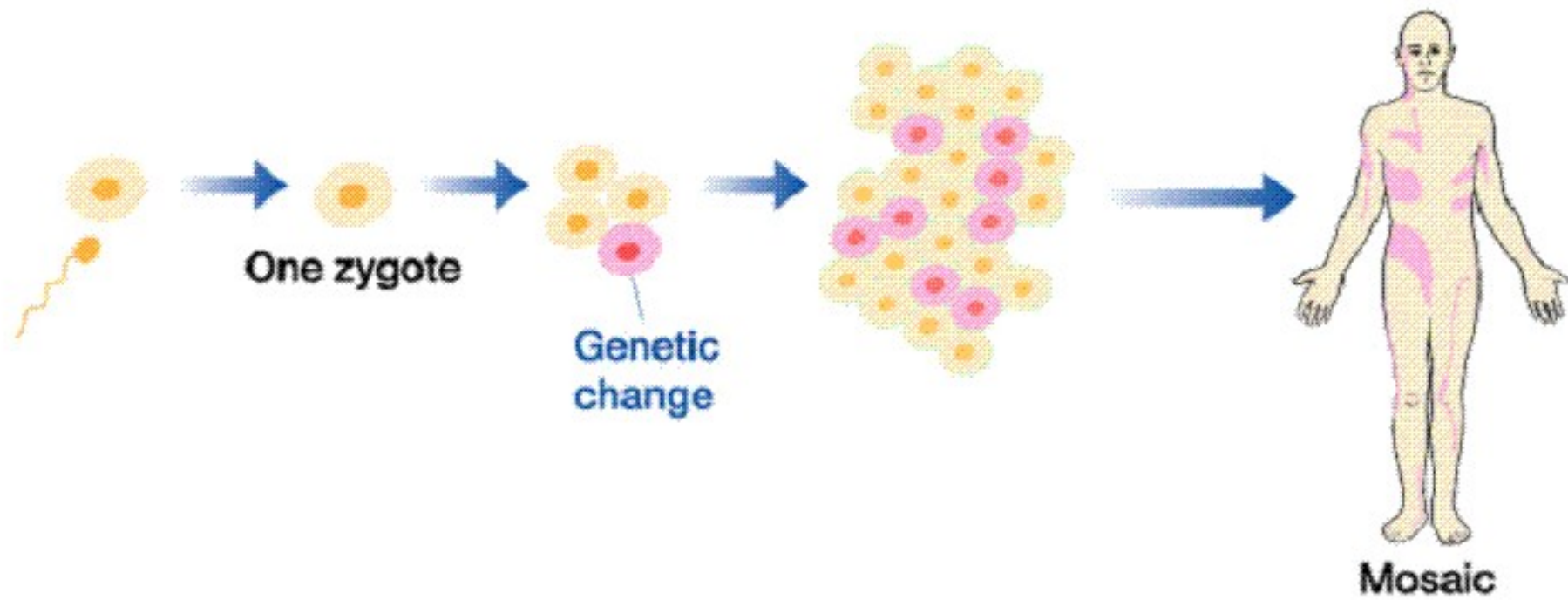




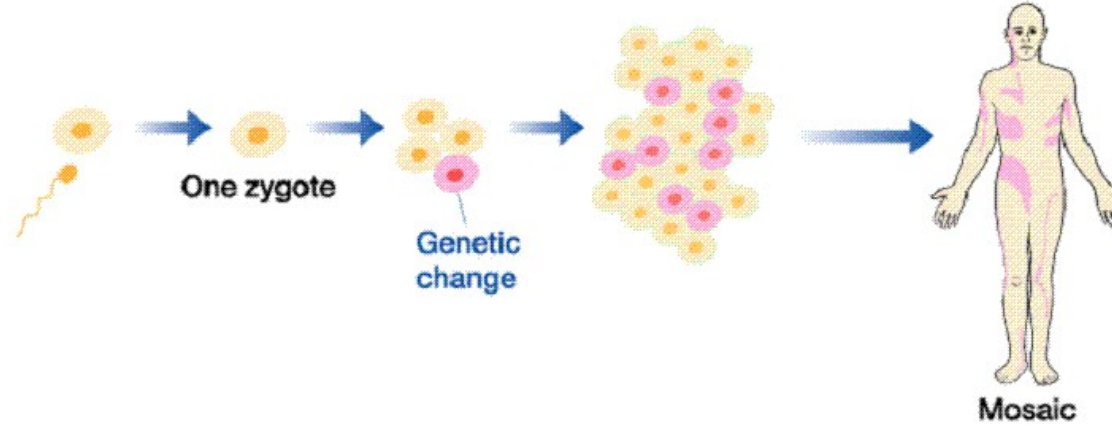
Germline



Somatic cells

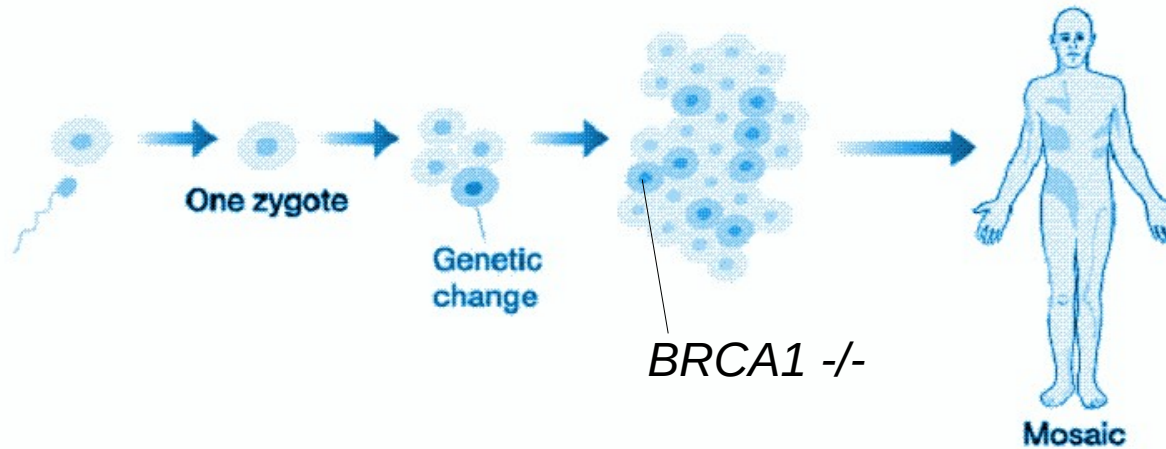


woman  
*BRCA1* +/+



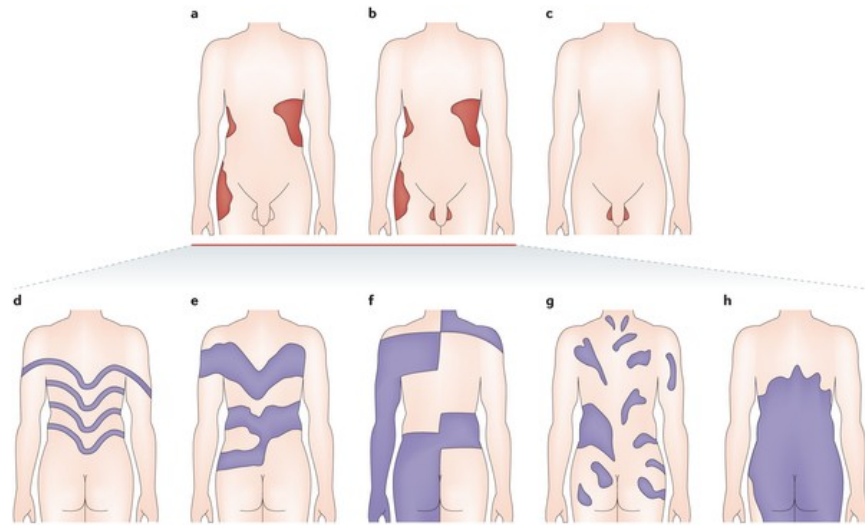
**10% chance to develop breast cancer**

woman  
*BRCA1* +/-

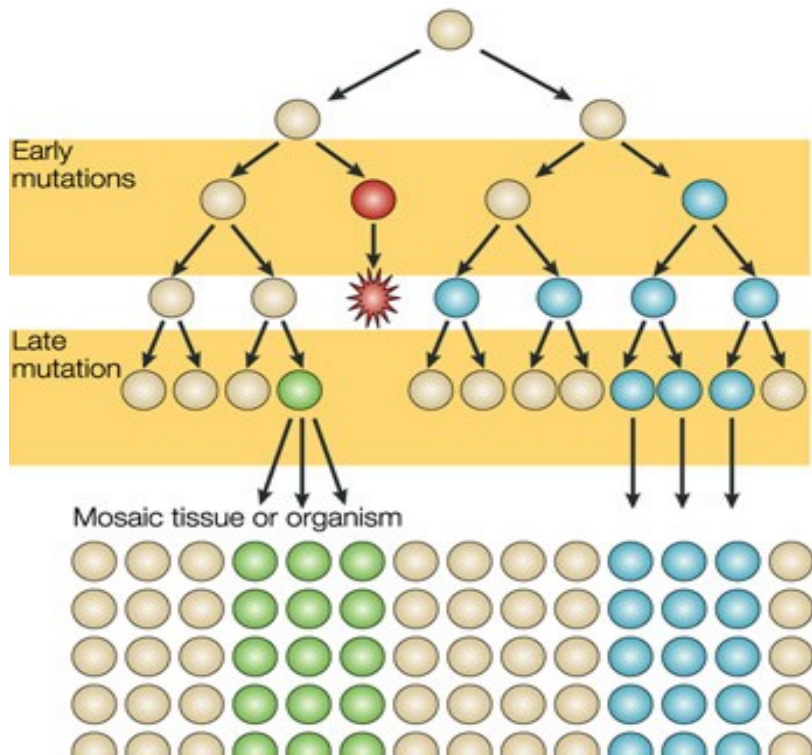


**45% chance to develop breast cancer before 70 years old**  
Cancer cells will be *BRCA1* -/-

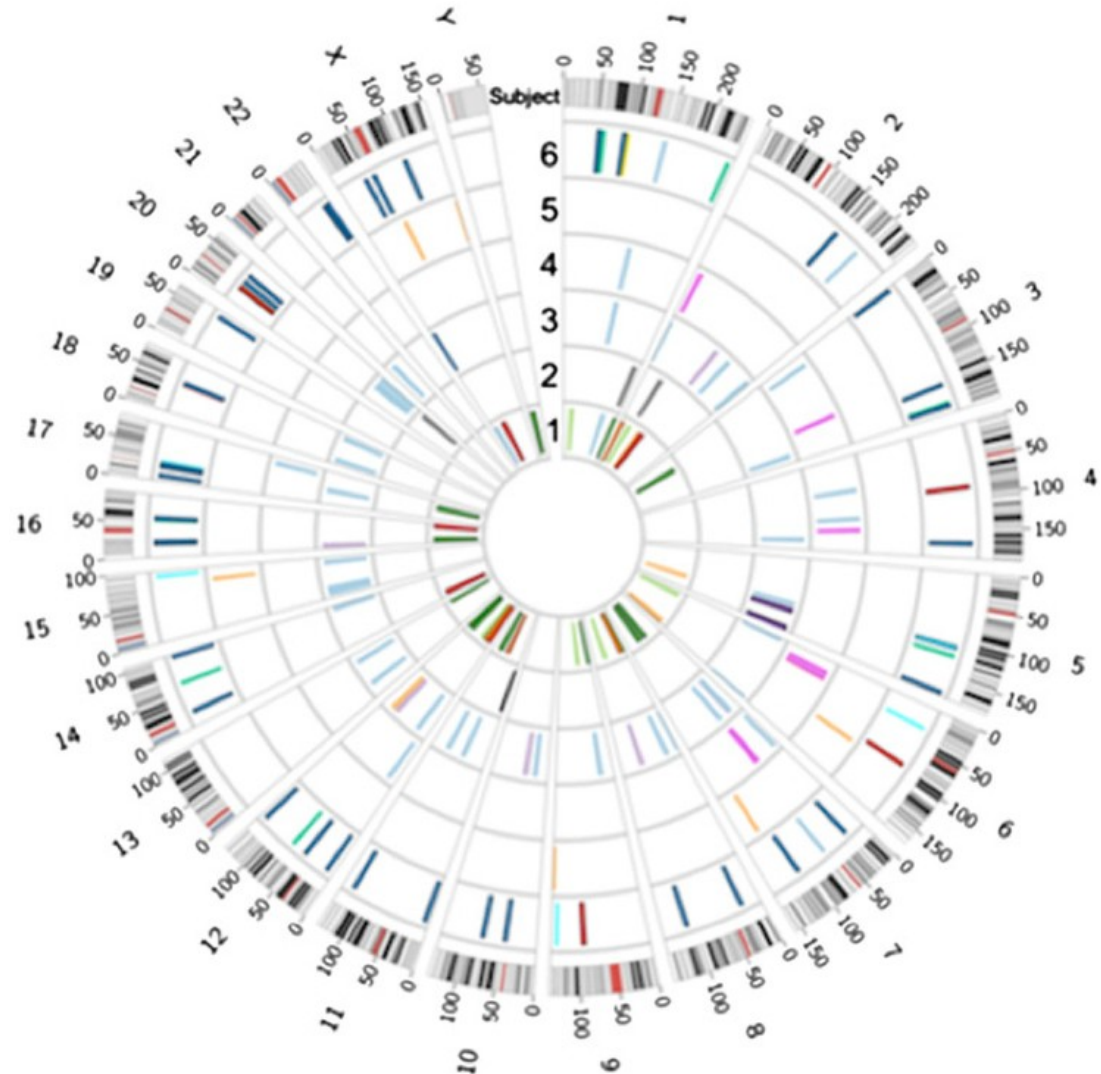
# Somatic mosaicism



Nature Reviews | Genetics



73 somatic CNVs in 11 tissues of six persons

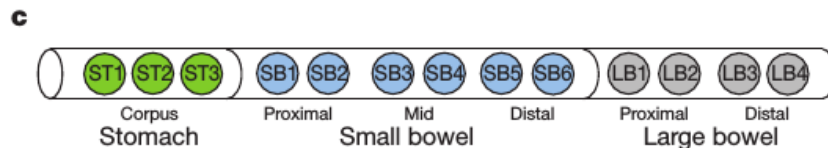
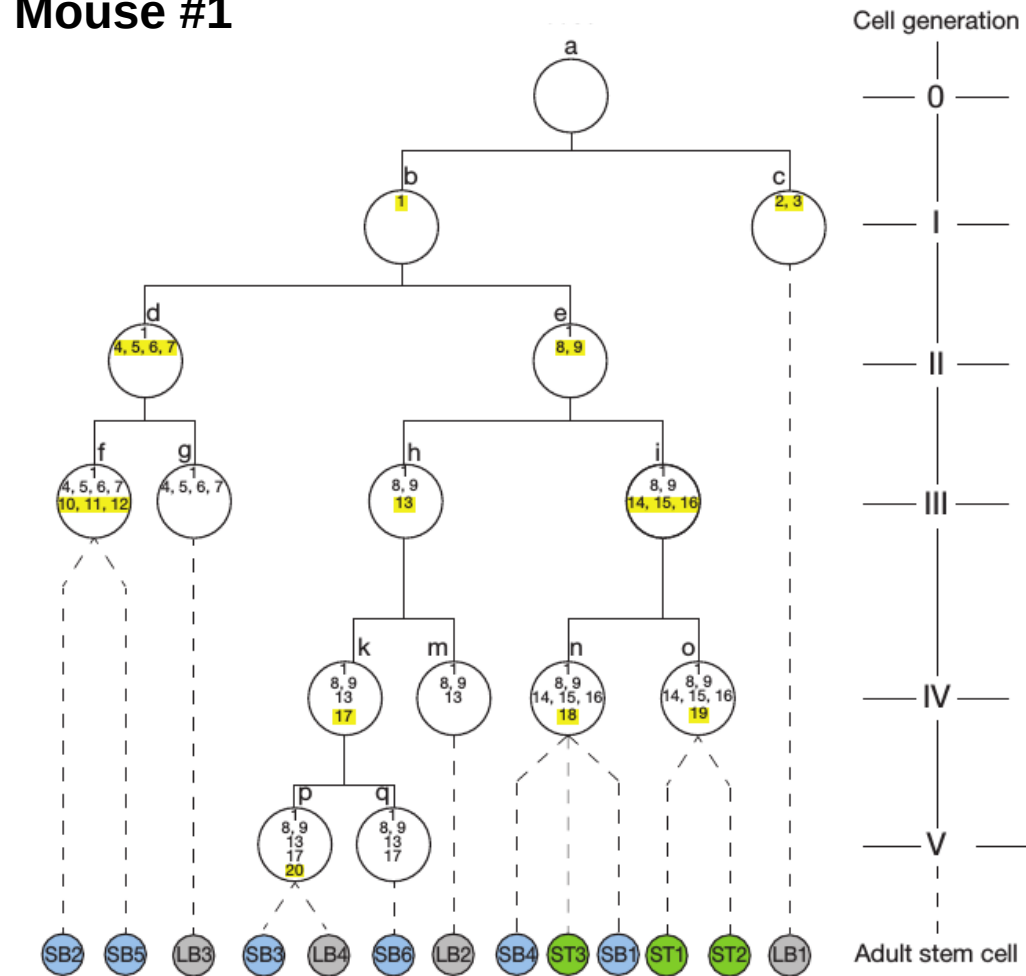


- O'Huallachain 2012 PNAS

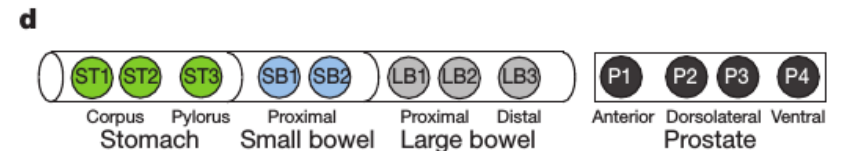
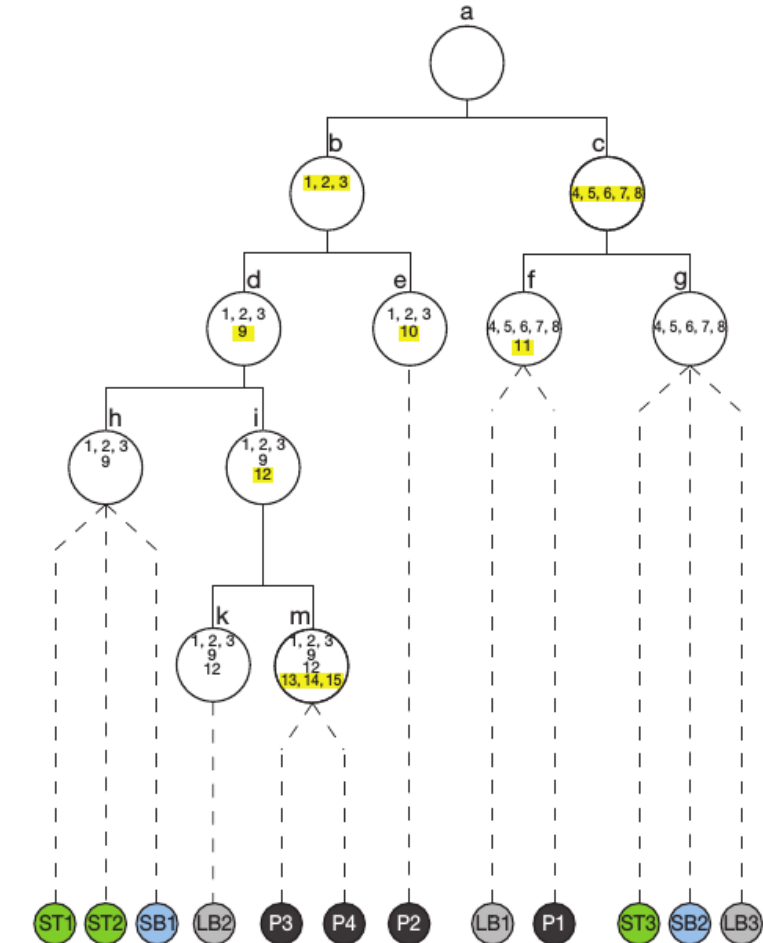


# Somatic mosaicism used to reconstruct cell lineages

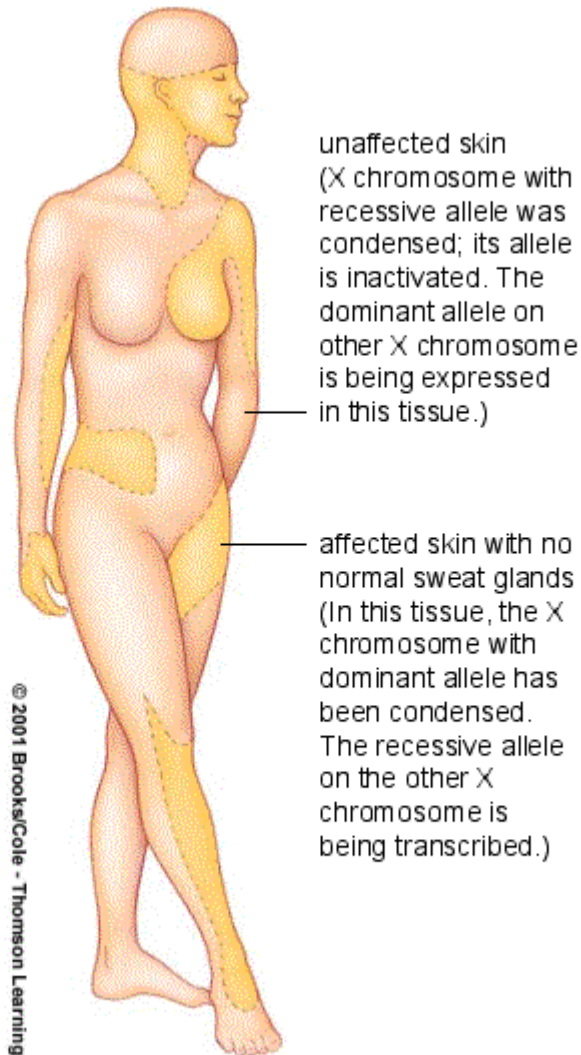
Mouse #1



Mouse #2

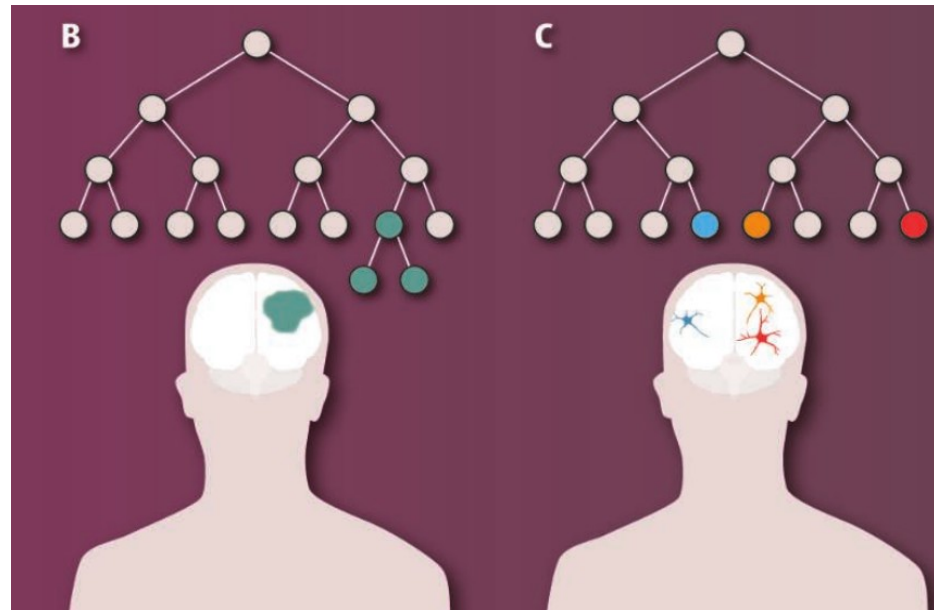


# Female mosaicism X inactivation pattern





# Somatic transposition in human brain



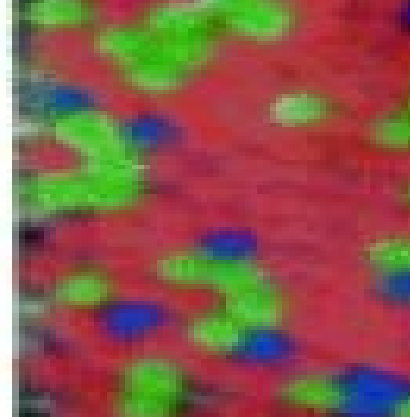
In three individuals:

in the hippocampus and caudate nucleus

7,743 somatic L1 insertions, 13,692 somatic Alu insertions and 1,350 SVA insertions

# Developmental noise

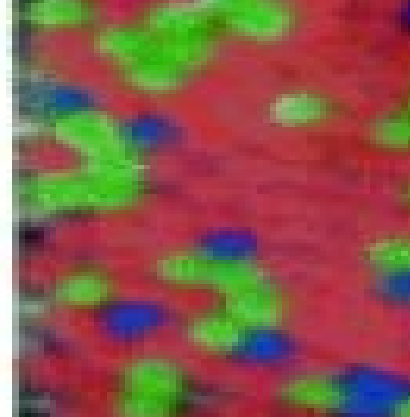
Differences between left and right sides of the body



ear shape, neuron connectivity, olfactory receptor gene expression, X inactivation pattern, organ cell number and size...

# Developmental noise

## Differences between left and right sides of the body



ear shape, neuron connectivity, olfactory receptor gene expression, X inactivation pattern, organ cell number and size...

## Differences between twins

immune system cells, gait, arms crossing, voice, heart beat, brain waves...

## Some can be attributed to variation in the number of determinant molecules

During terminal differentiation of mouse 3T3-L1 pre-adipocytes, individual TF abundance differs dramatically (from ~250 to >300,000 copies per nucleus) and the dynamic range can vary up to fivefold during differentiation.

Simicevic 2013 Nature

# Causes of phenotypic differences ?

**Genetic**



**Epigenetic Environment**



**Stochasticity**



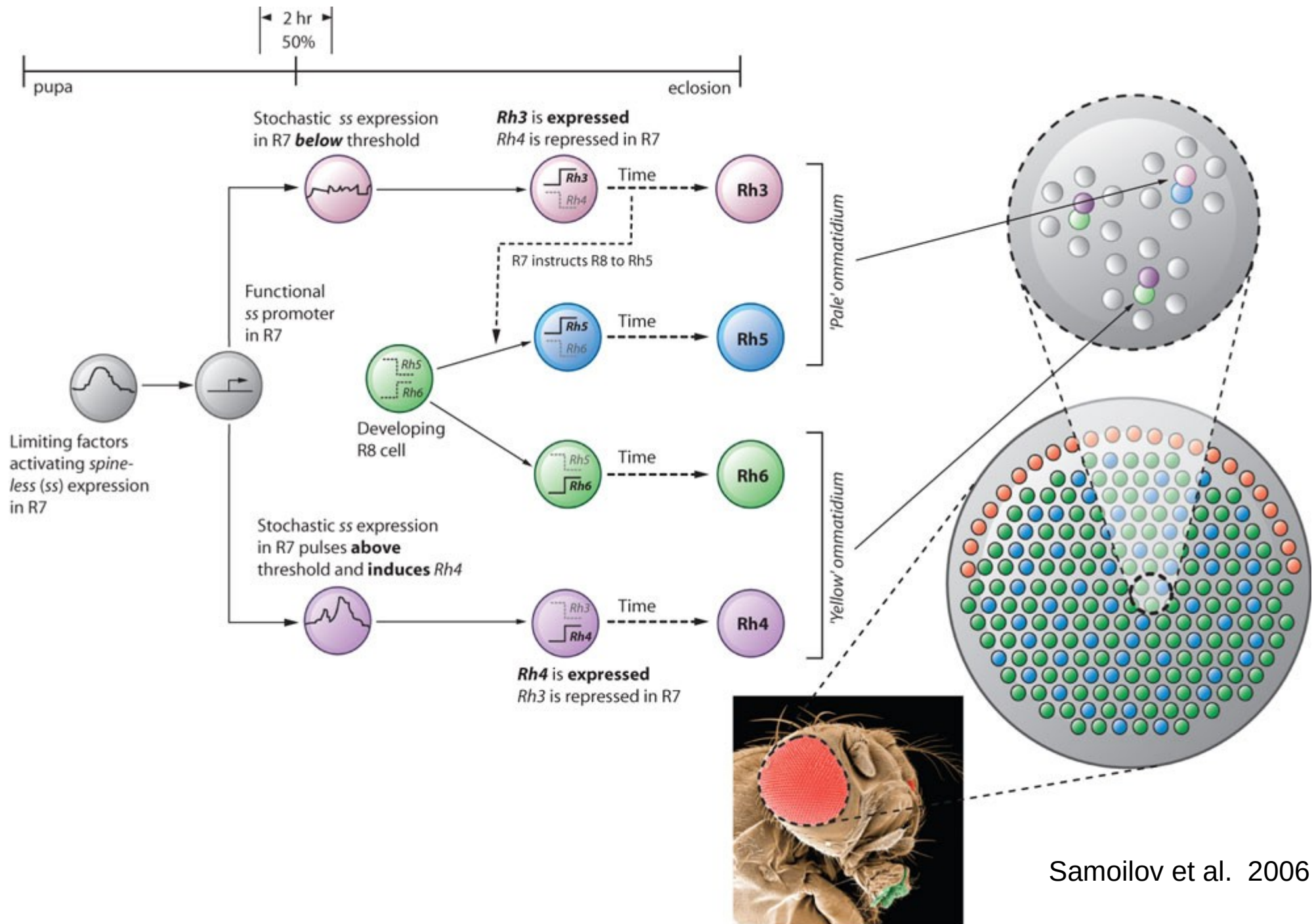
**Transmitted**

**Deterministic**

**Interactions**

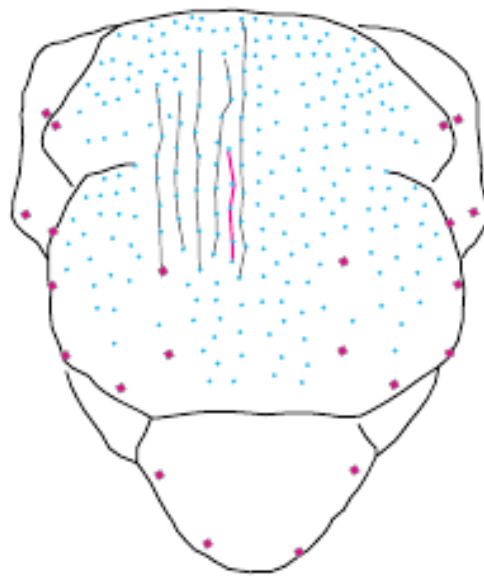


# Developmental noise can be “good”

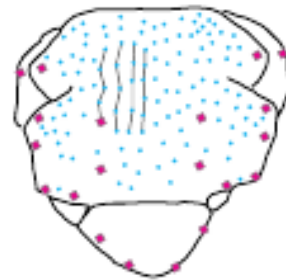


# Robustness

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Well-fed



Semi-starved

# Robustness

**Absence or low variation of a phenotype  
when faced with an incoming variation**

**1) Of what?**

**2) To what?** To either:  
- stochastic variation  
- environmental variation: specify  
- genetic variation: specify

**3) How much?**

Different phenotypic metrics

Coefficient of variation: standard deviation/mean

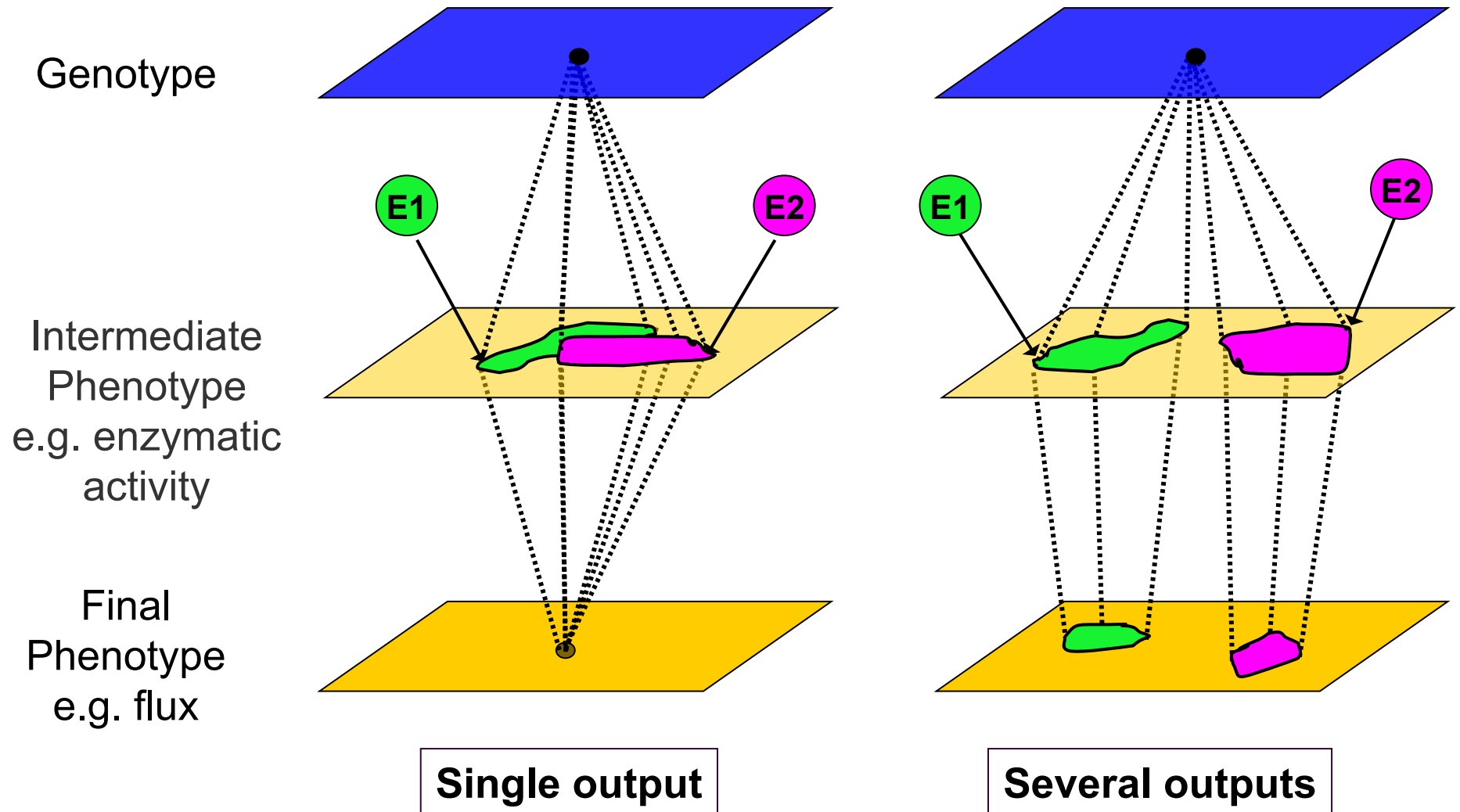
Historically:

quantitative genetics (low variance, canalization)

physics/chemistry/engineering (robustness, buffering)

**Canalization:** mechanisms that make the system follow a certain trajectory

# Trait plasticity versus invariance (robustness) at different levels of the genotype-phenotype map

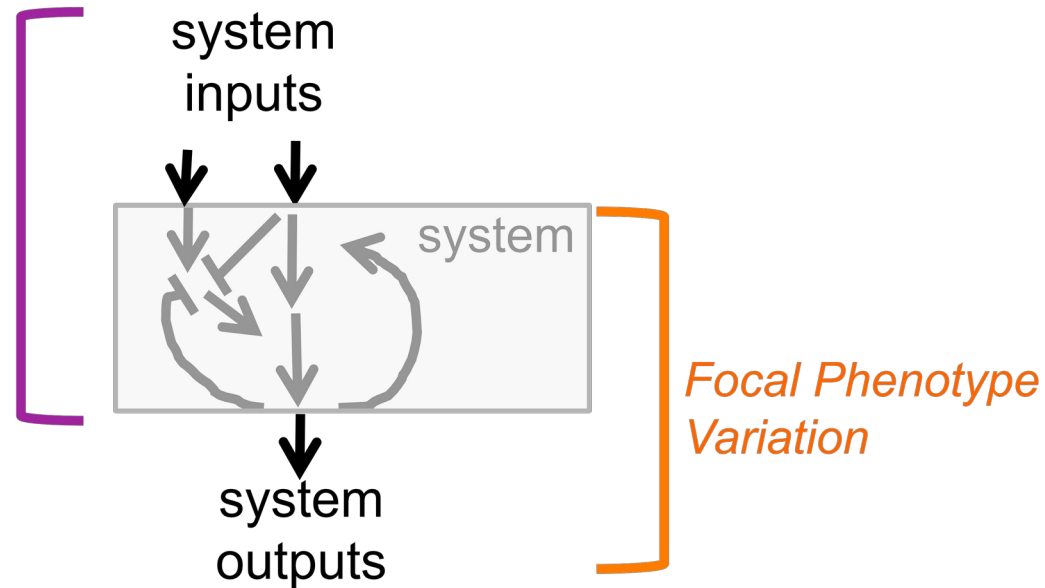




## *Propagation of variation*

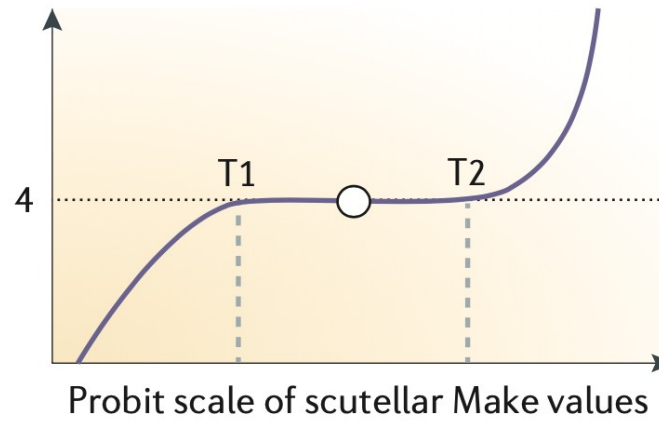
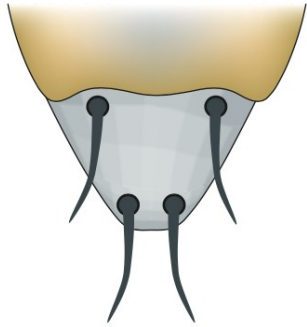
*Incoming Variation:*

- Noise
- Environmental
- Genetic

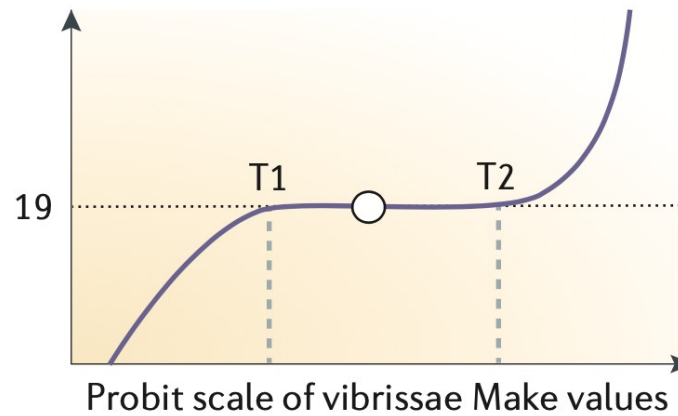
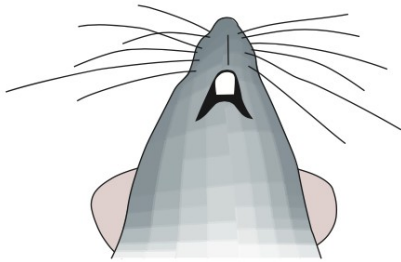


## b Experiments

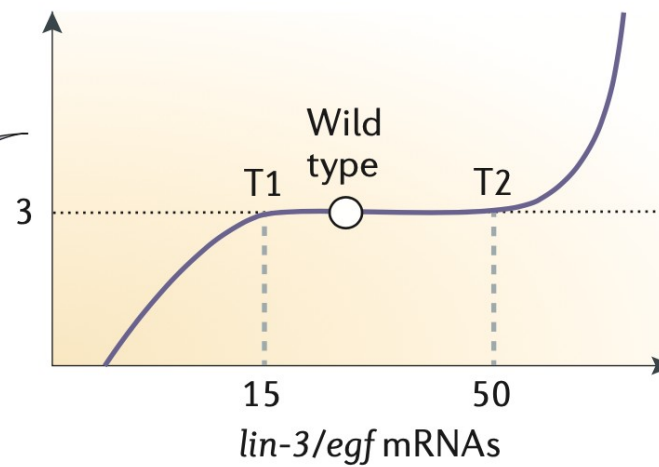
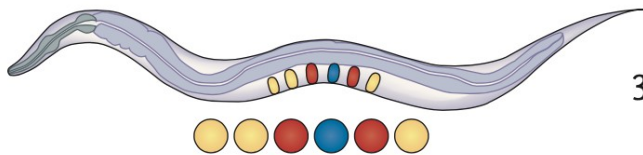
Number of scutellar bristles



Number of vibrissae



Vulval induction index



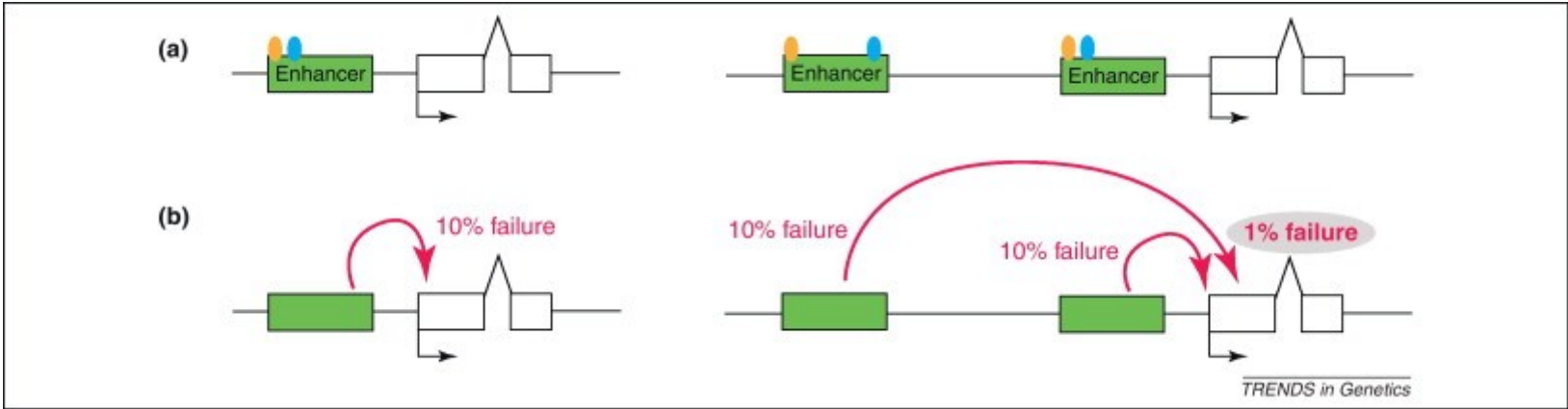
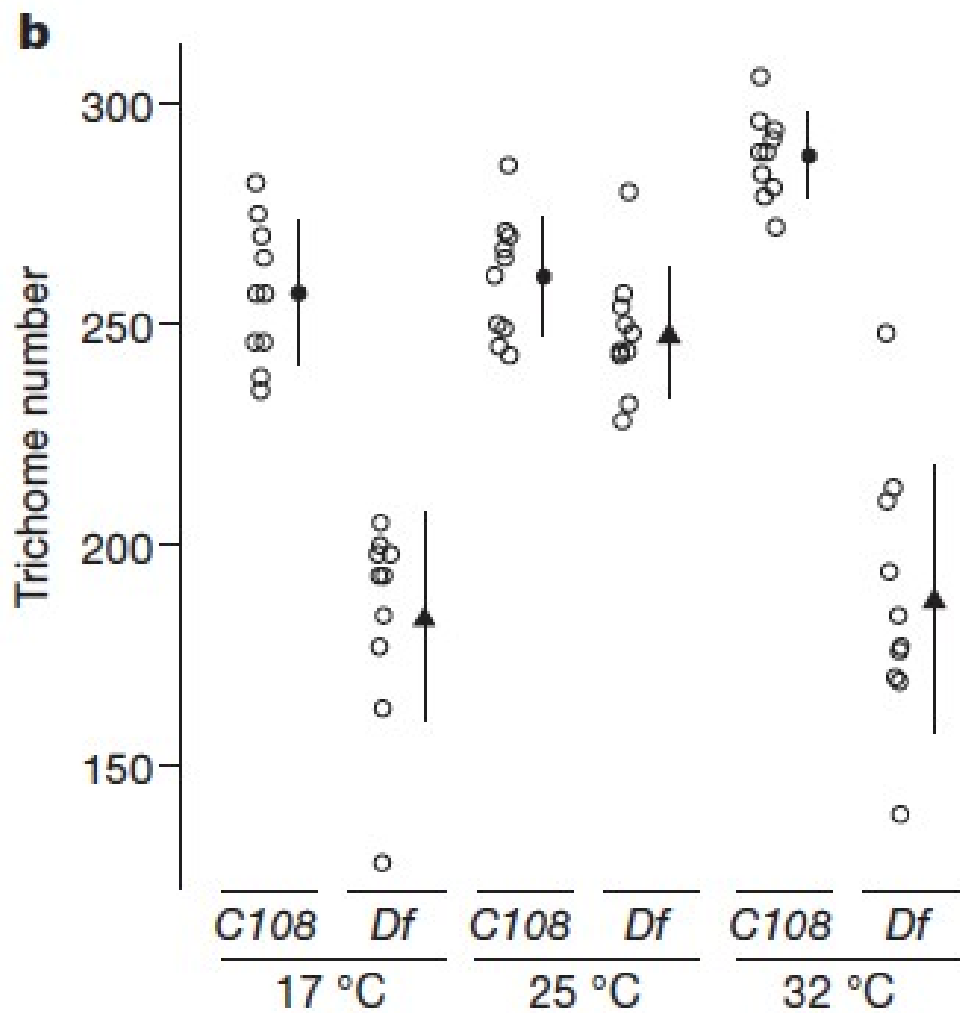
# Causes of robustness

Non-linearity

Redundancy

LETTERS

Phenotypic robustness conferred by apparently redundant transcriptional enhancers



# **Cryptic genetic variation**

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# Cryptic genetic variation

First requires defining the *phenotype of interest*

**Genetic variation that has no effect on phenotype of interest**

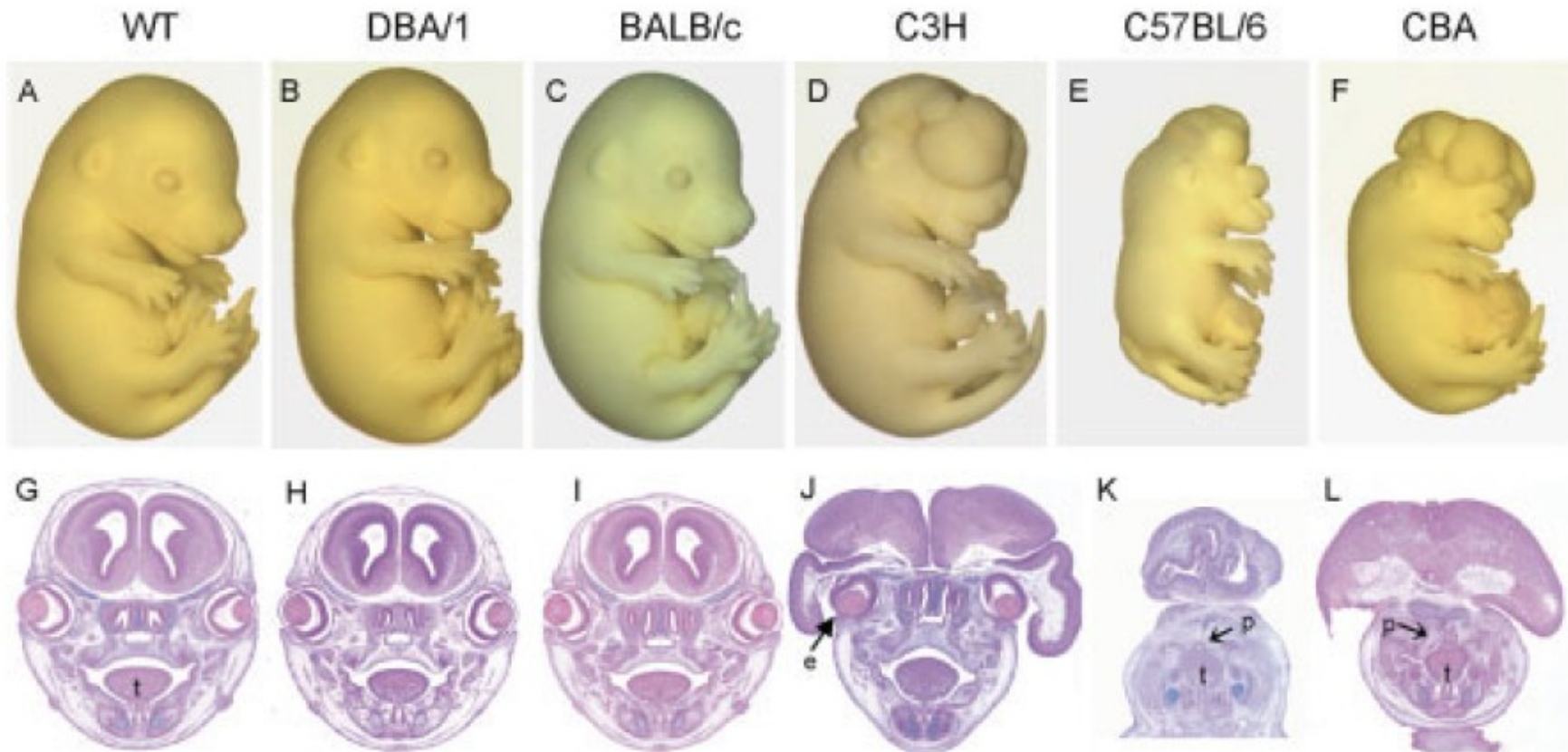
**... but may be revealed *under some circumstances*  
by its effect on this phenotype**

Cryptic genetic variation (CGV) is defined as standing genetic variation that does not contribute to the normal range of phenotypes observed in a population, but that is available to modify a phenotype that arises after environmental change or the introduction of novel alleles.

Gibson & Dworkin *Nat Rev Gen* 2004

# Expressivity of one mutation varies with wild genetic background

*Tcof1*<sup>-/-</sup> heterozygote mice

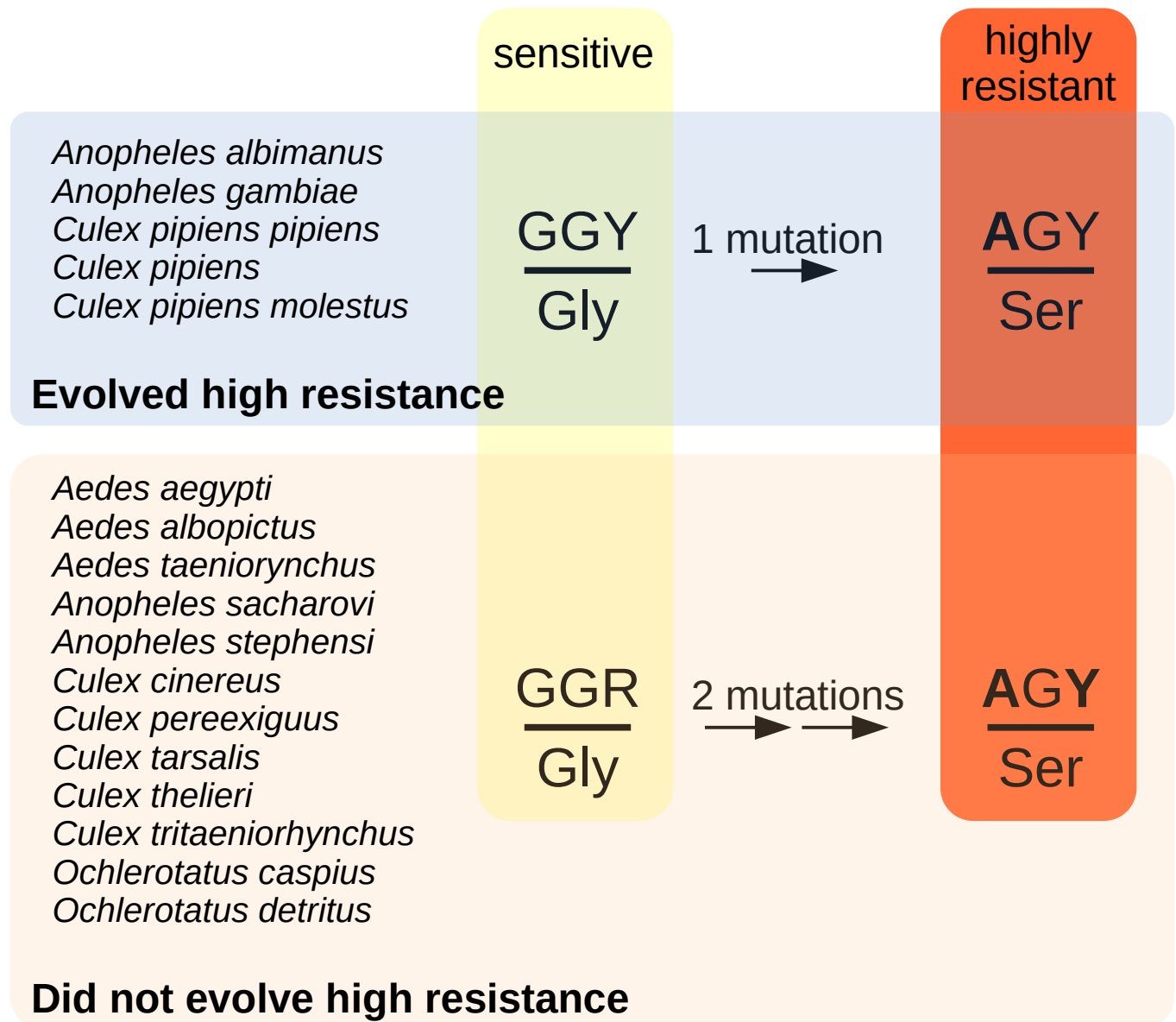


# Influence of contingency

Resistance  
to carbamates and  
organophosphates



position 119  
in *AchE1* gene



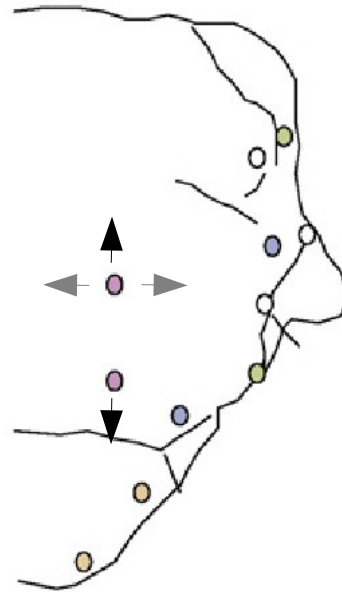
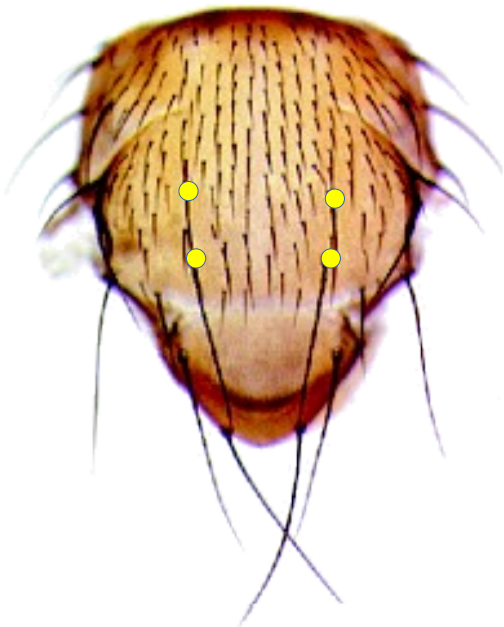
.. But might if more time is allowed



# The genome constrains evolution

## Standing variation

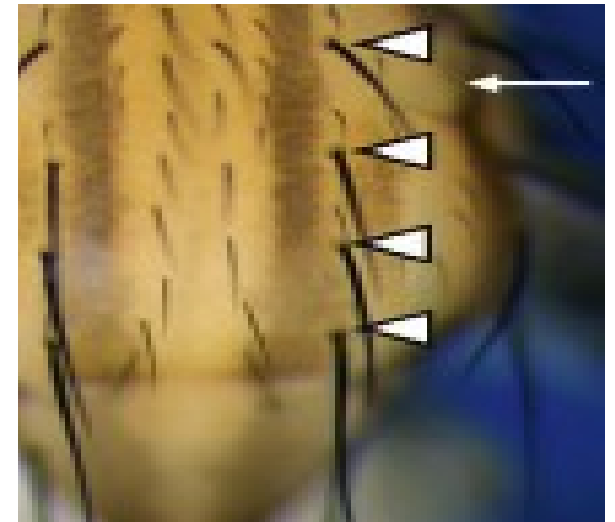
*D. melanogaster*



► variation  
► no variation

## Natural evolution

*D. quadrilineata*



Marcellini et al 2006 PLoS Biol

# The Genotype-Phenotype Map

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# The first genotype-phenotype map

Here a dot represents  
the mean state of a  
**population**

## Genotype space

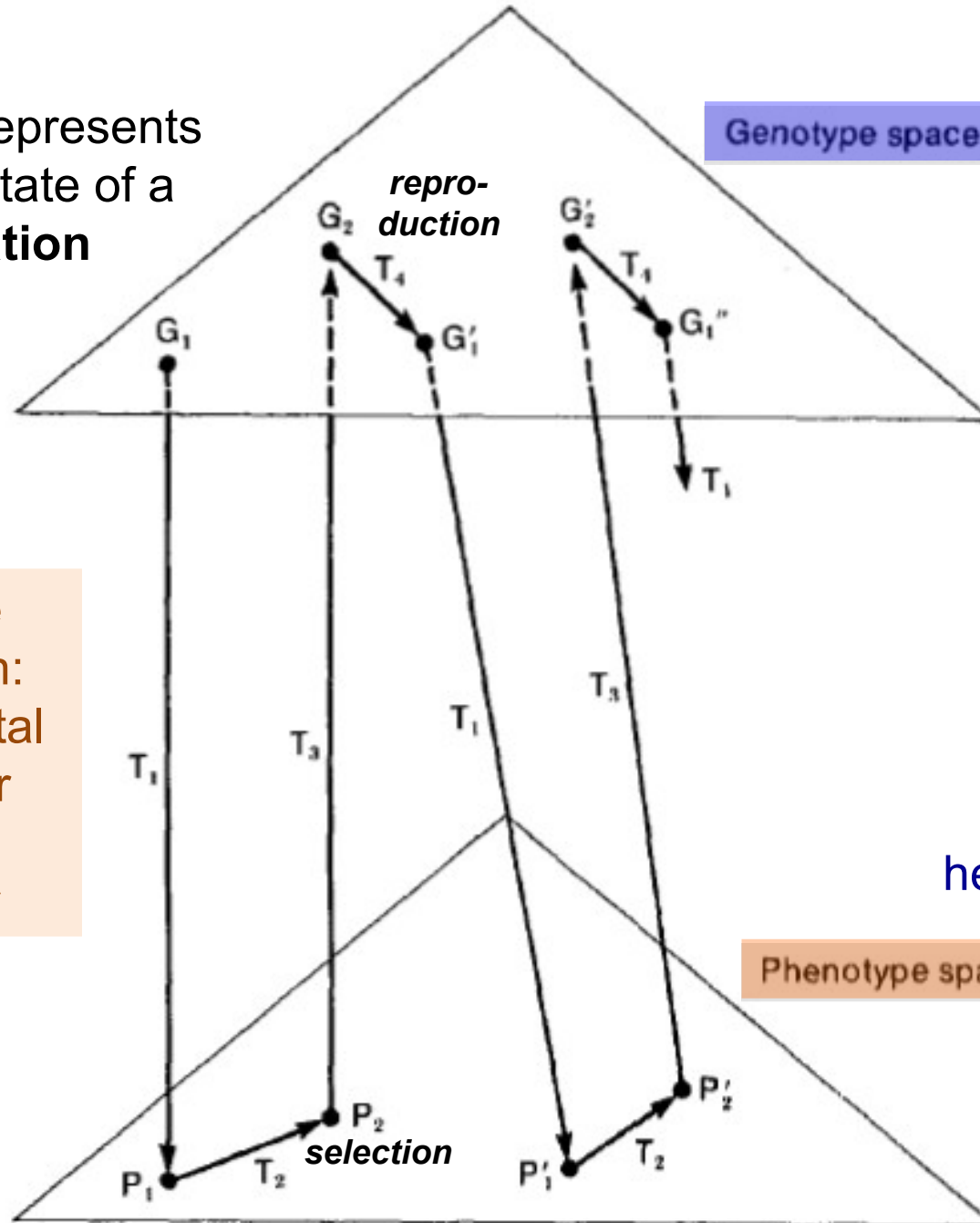
# Population genetics: stochastic processes and selection coefficient

Phenotype  
construction:  
developmental  
and cellular  
biology  
physiology

# Quantitative genetics: heritability of phenotypes

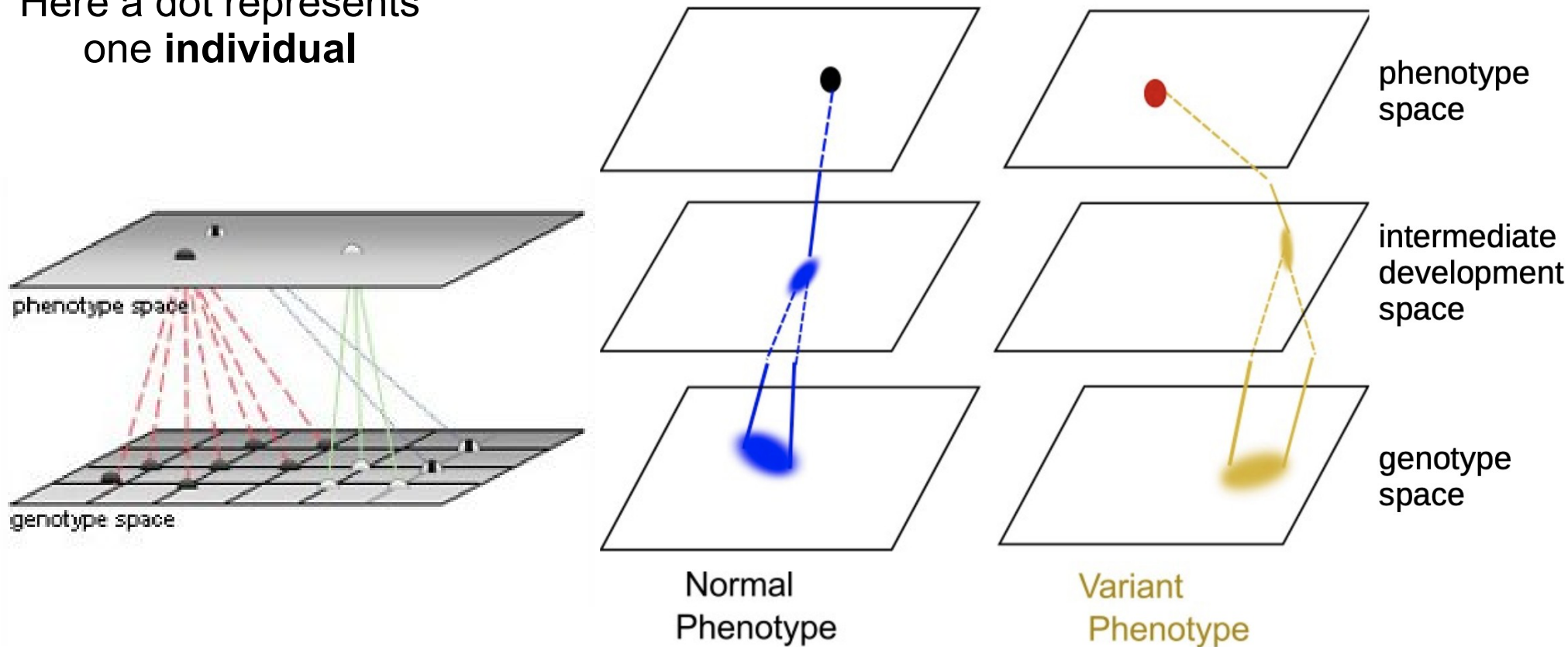
## Phenotype space

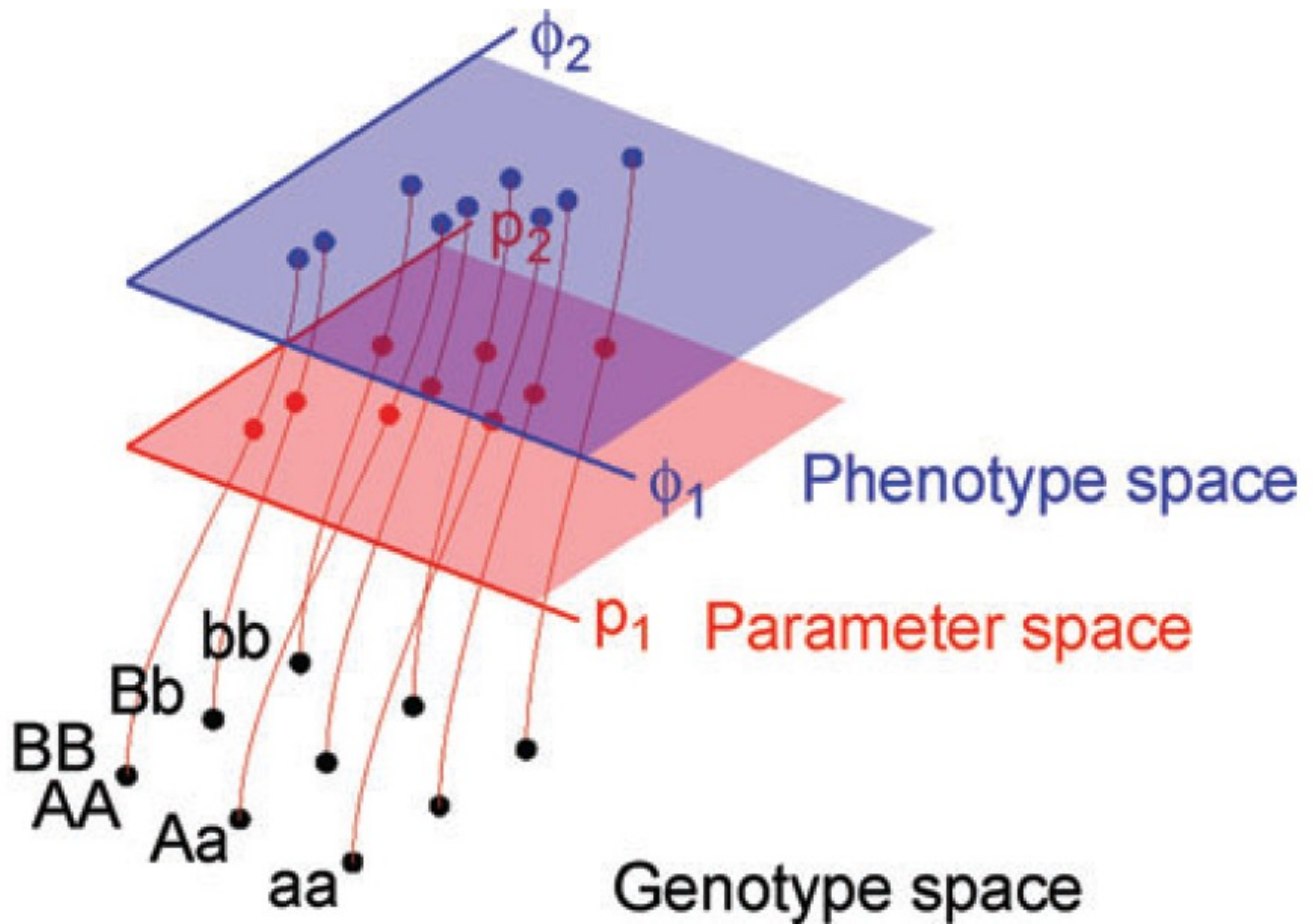
Evolutionary  
biology of phenotypes,  
evolutionary ecology



# Intermediate steps in the genotype-phenotype map

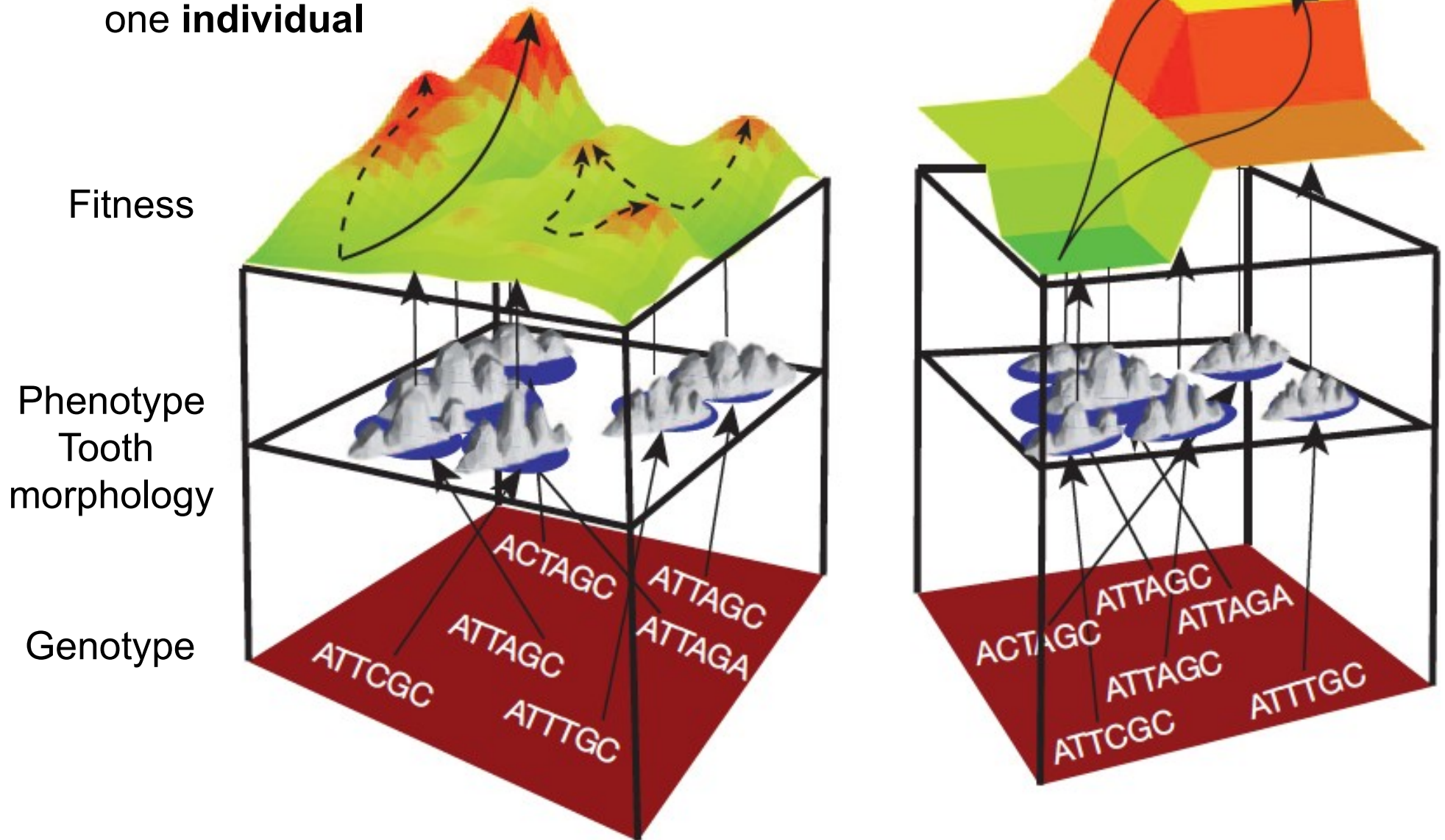
Here a dot represents one **individual**





# The genotype-phenotype-fitness map

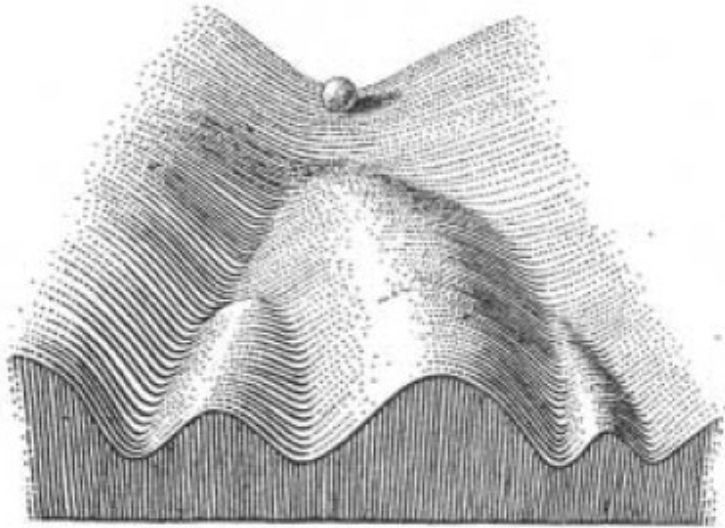
Here a dot represents  
one **individual**





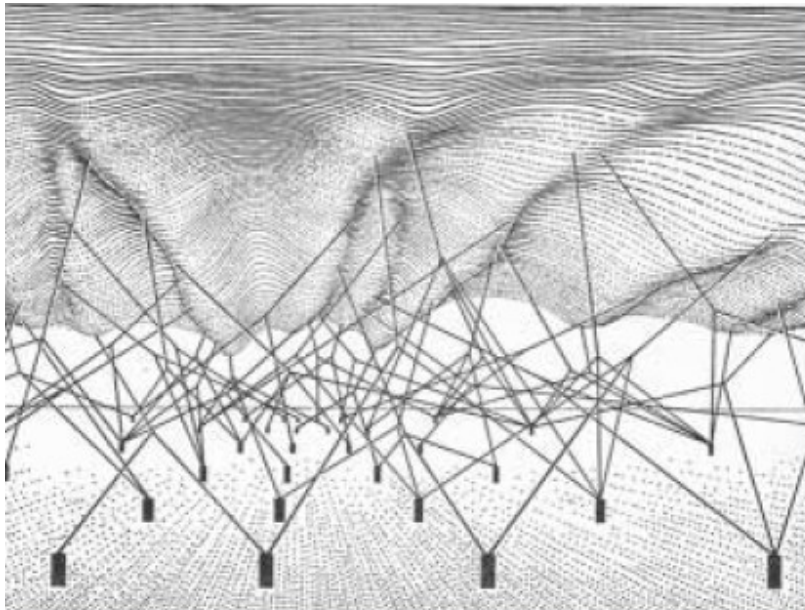
# The Epigenetic Landscape

## A metaphor for the G-P relationship

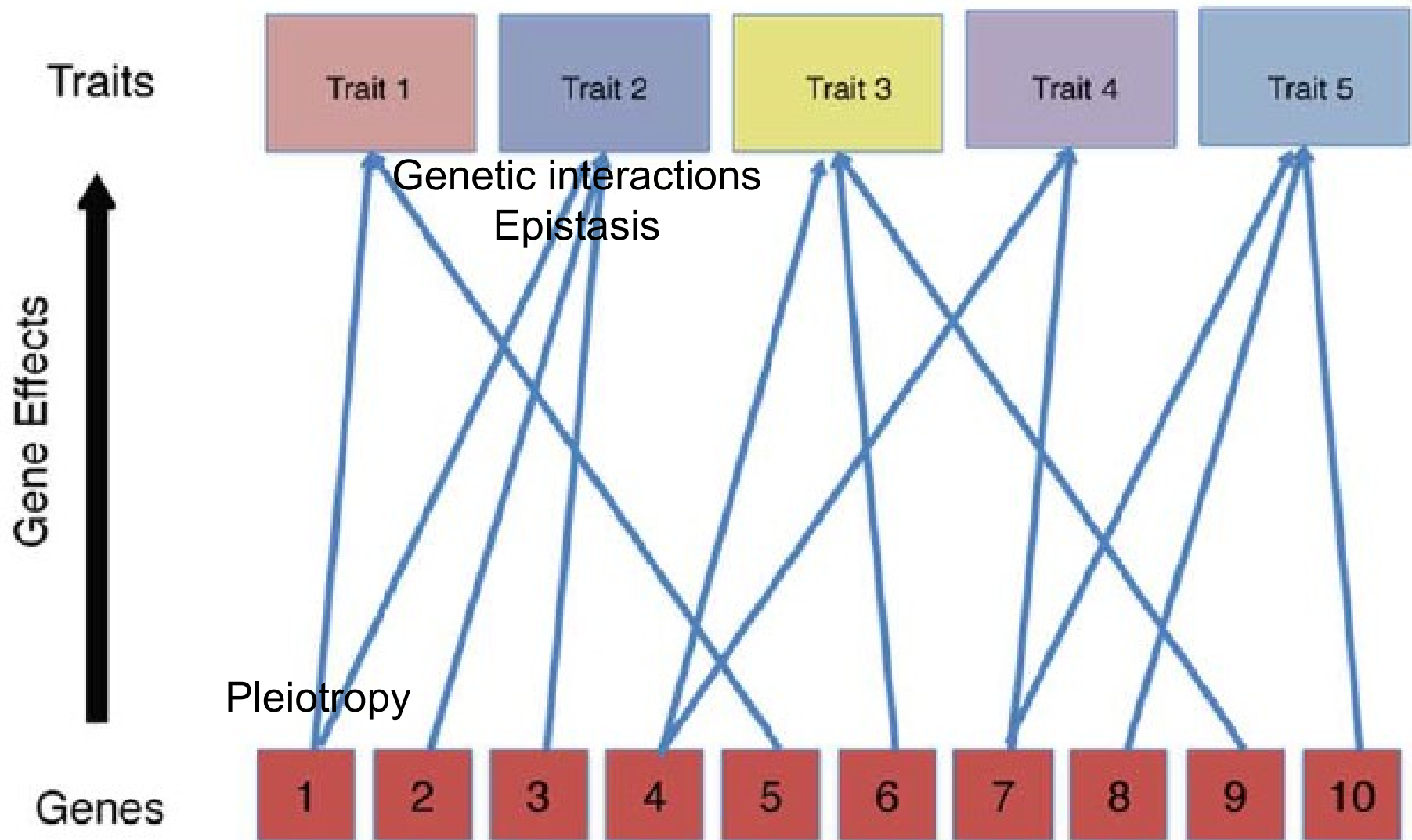


Development

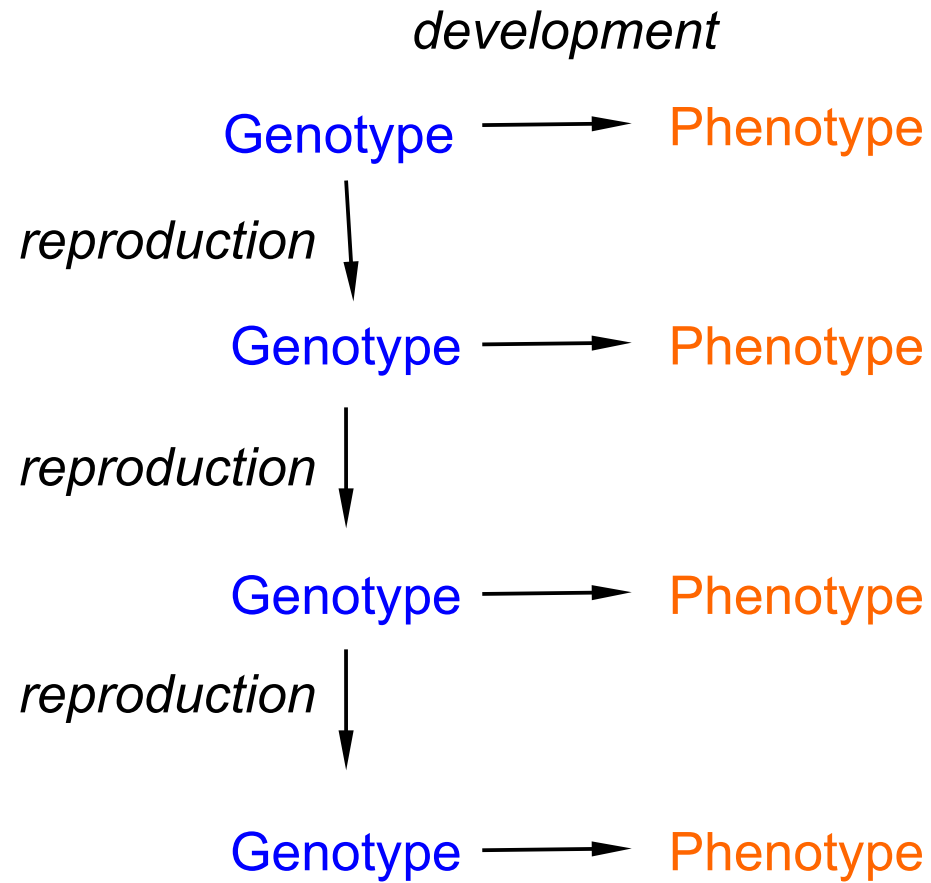
Canalization



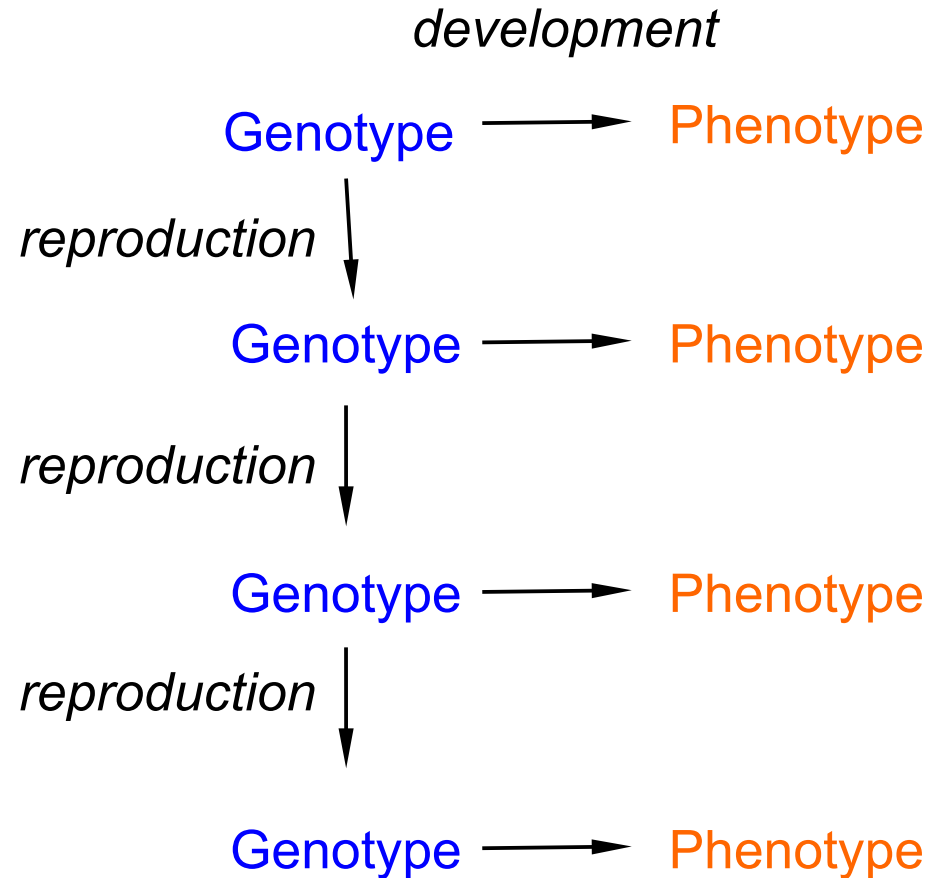
Genes underlying  
the landscape







# A simplistic view



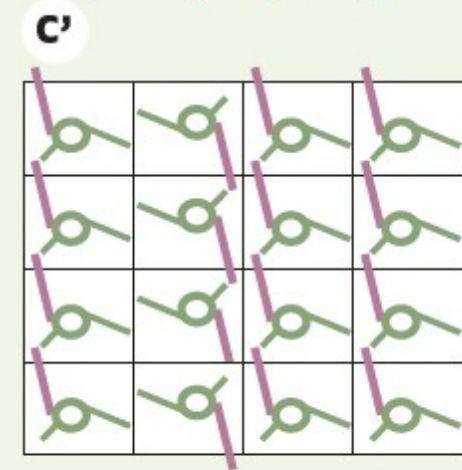
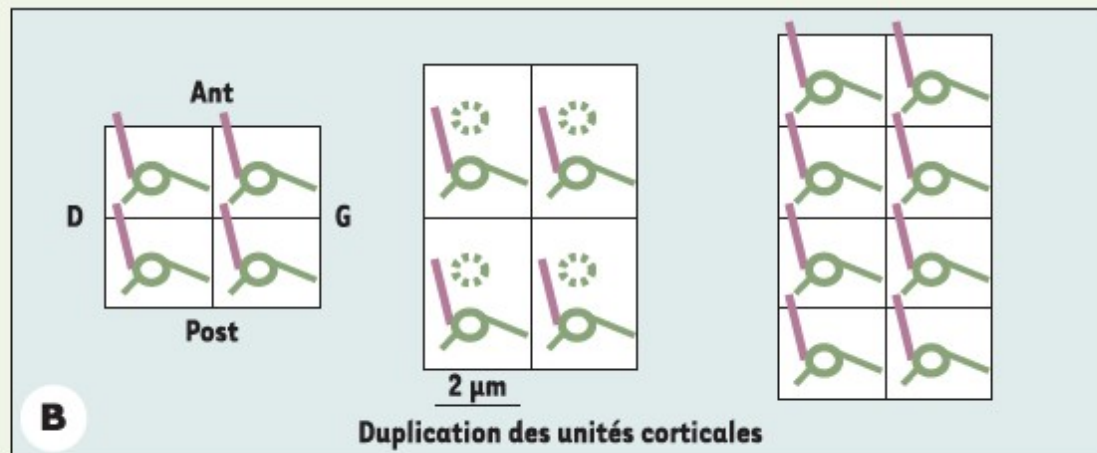
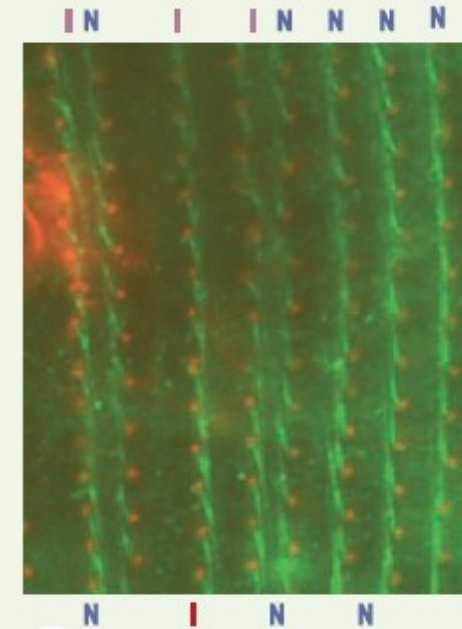
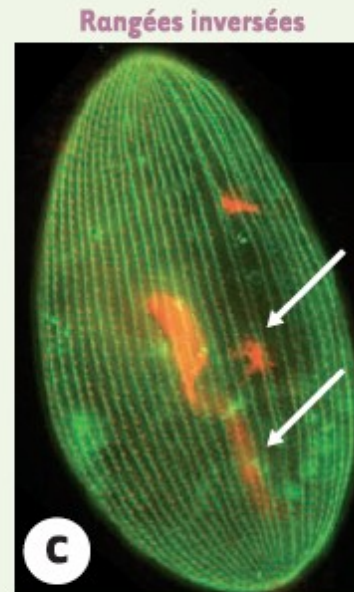
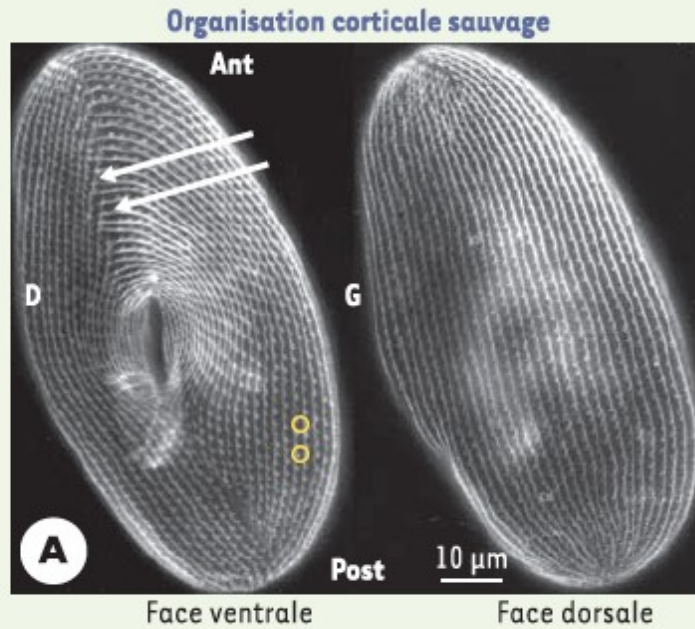
Heritable traits are not always due to genes

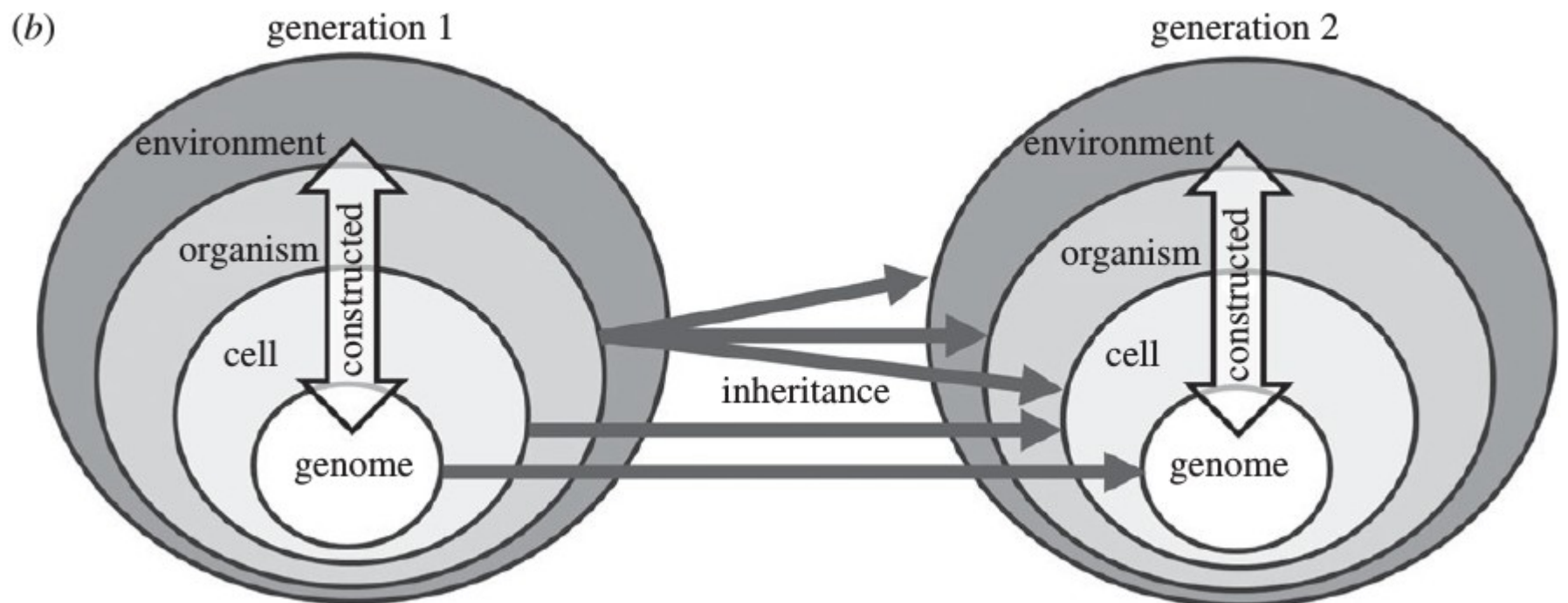
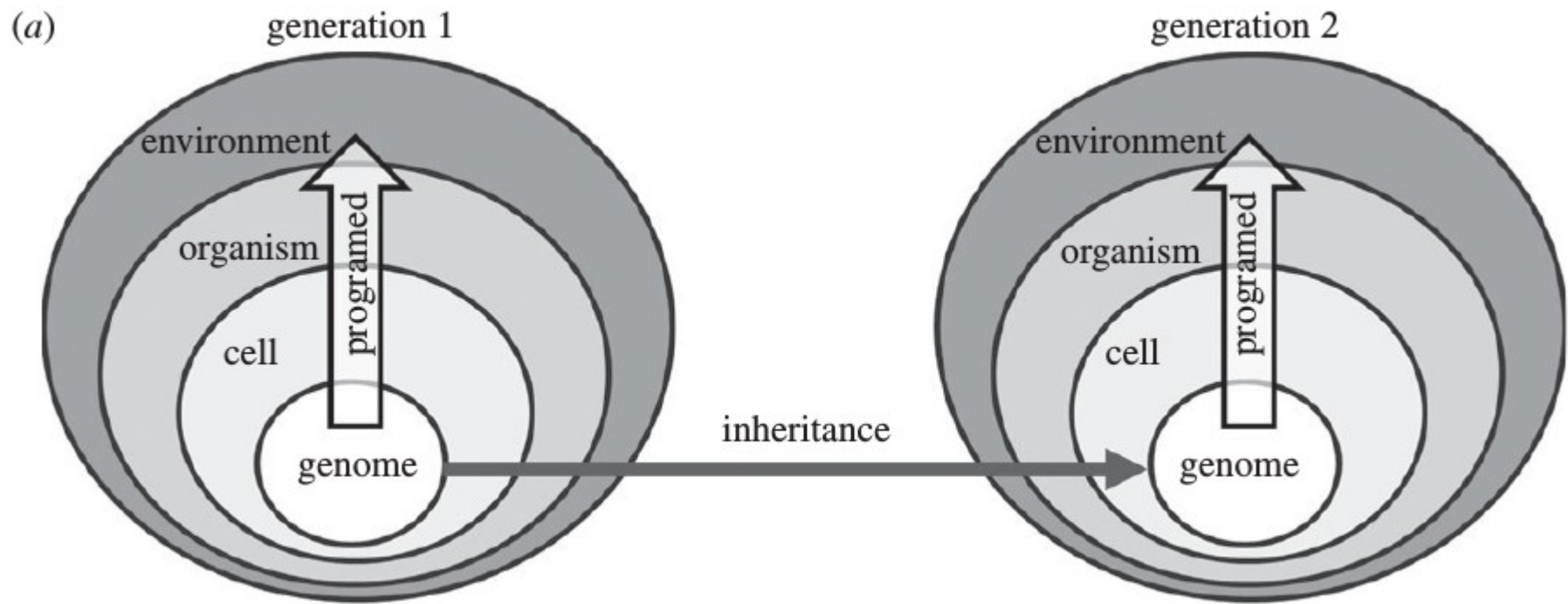
The genotype does not determine entirely the phenotype

The genotype cannot replicate by itself

Genotype and phenotype imply variation

# Cortical heredity in *Paramecium*

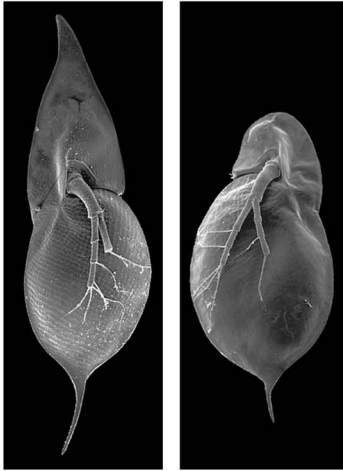






# Plasticity: one genotype → several phenotypes

Daphnia



with  
helmet

without  
helmet

Nemoria arizonaria caterpillars



spring: caterpillars feed  
on catkins



summer: caterpillars feed  
on leaves

Water crowfoot plant



leaves growing  
above water

leaves growing  
below water

Desert locusts



solitary



gregarious

Commodore butterfly



winter



summer

Commodore butterfly: Michael Wild, CC-BY-SA-3.0 (winter), Svdmlen, CC-BY-SA-3.0 (summer)

Daphnia: Agrawal *et al* (1999)

Nemoria arizonaria caterpillars: Sadava *et al* (2014)

Water crowfoot plant: J R Crellin, CC BY-NC-ND 3.0

# **Complexifications of the G-P map**

**Genetic Linkage**

**Large number of alleles**

**Epistasis**

**Noise**

**Supergene**

**Robustness**

**Pleiotropy**

**Cryptic genetic variation**

**GxE**

**Epigenetics**

**Plasticity**