

# **Manipulating DNA**

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**What can we do with DNA ?**

# What can we do with DNA ?

## **Extract, purify**

## **Make more**

Amplify

Clone

Synthesize

## **Examine**

Quantify

Examine length

Stain, probe

Sequence

Examine 3D structure

Measure physical properties of DNA molecules

## **Modify**

Cut

Ligate

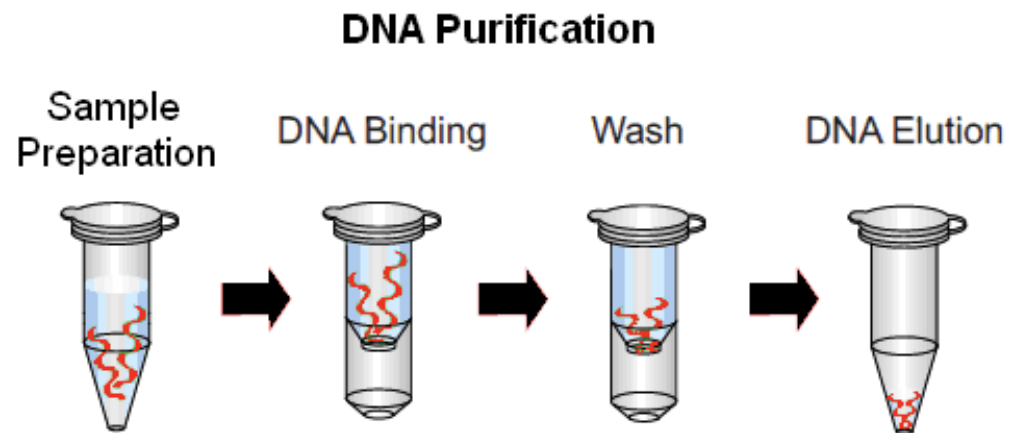
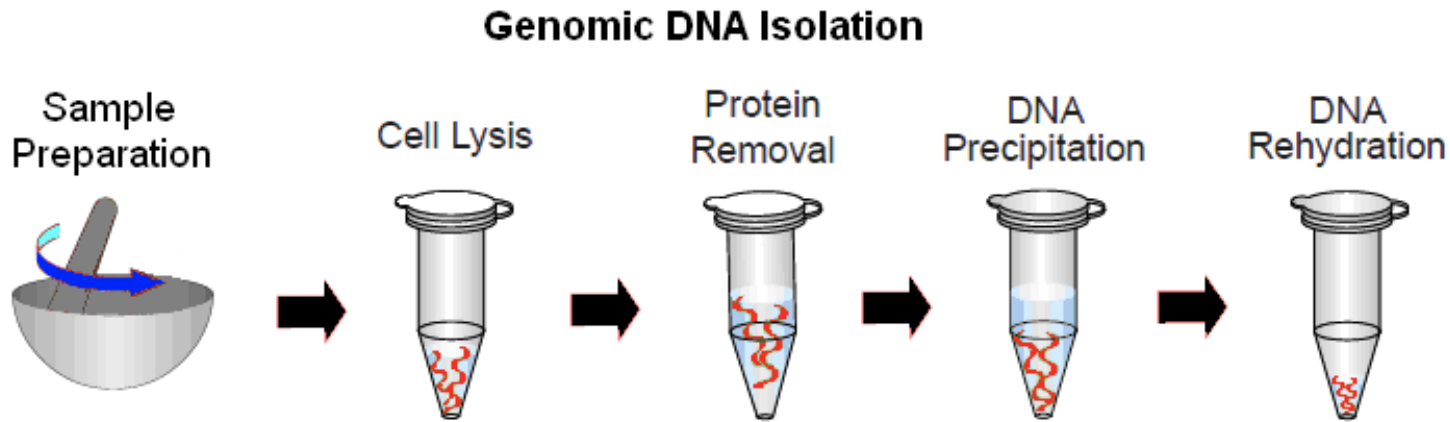
Recombine fragments

Introduce foreign DNA

Mutate

# Extract DNA

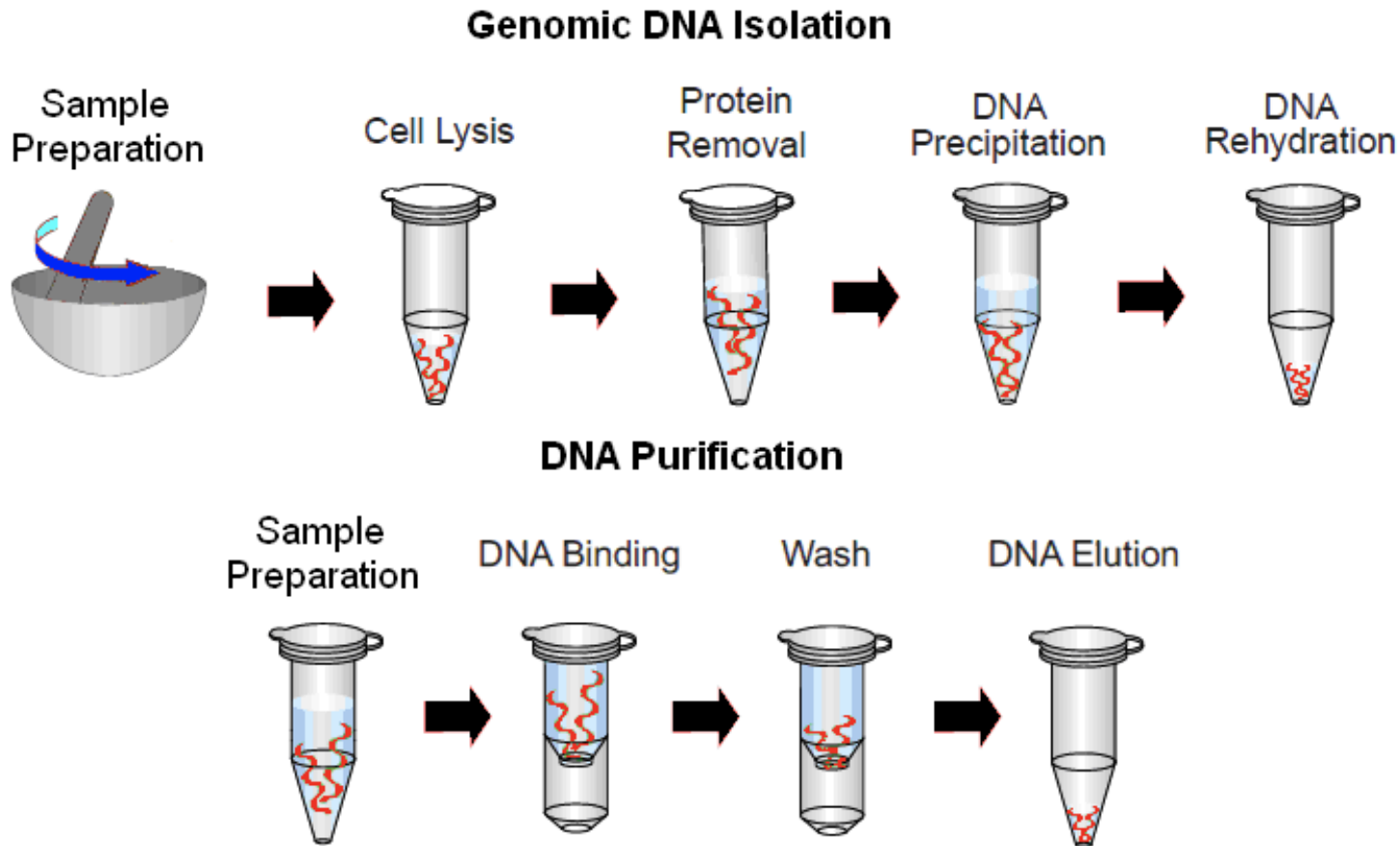
Break cells, remove lipids and proteins,  
precipitate DNA, remove liquid, resuspend in aqueous solution



***Be aware of contaminants!***

# Extract DNA

Break cells, remove lipids and proteins,  
precipitate DNA, remove liquid, resuspend in aqueous solution



***Be aware of contaminants!***

(DNA from mitochondria, viruses, bacteria, researcher, symbionts...)

# Amplify DNA



Mix:  
Genomic DNA  
Probes (oligonucleotides)  
Nucleotides  
Taq polymerase  
Ions ( $MgCl_2$ )

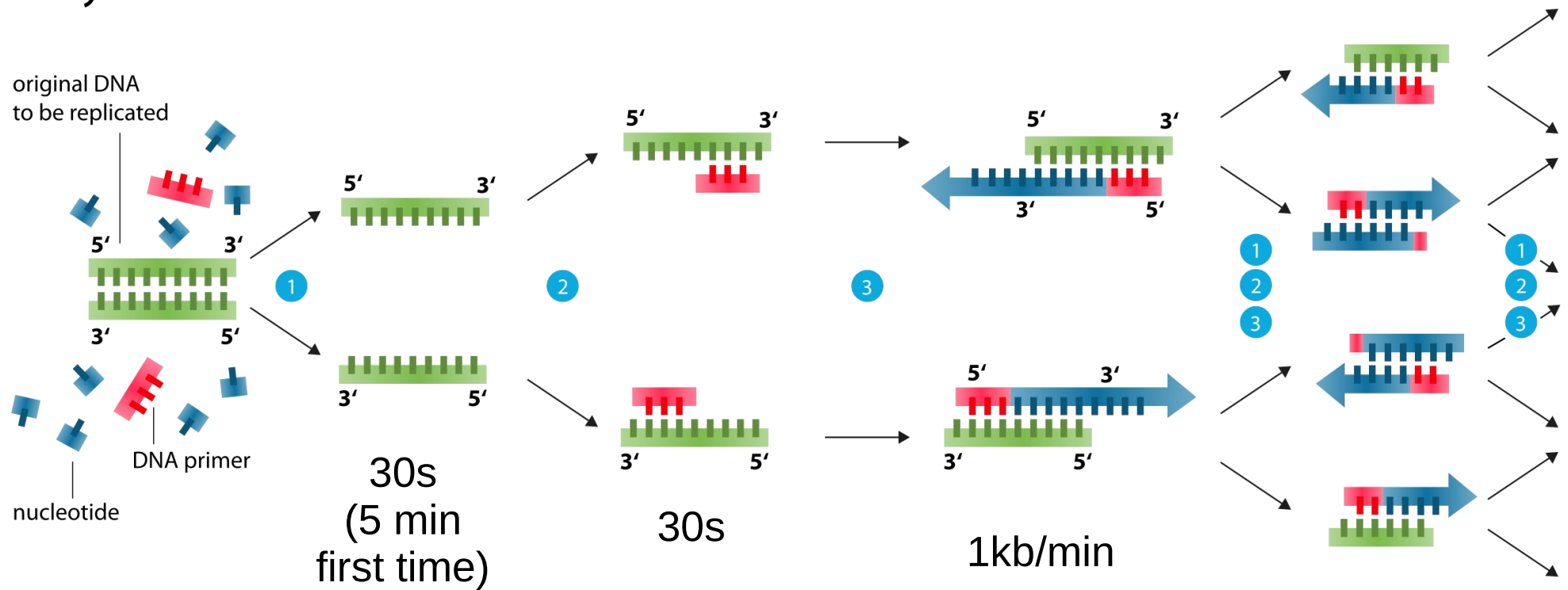
Cycles of Denaturation, Annealing,  
Elongation

## PCR: Polymerase Chain Reaction

Amplifies DNA fragments of between 0.1 and 10 kb (up to 40 kb)

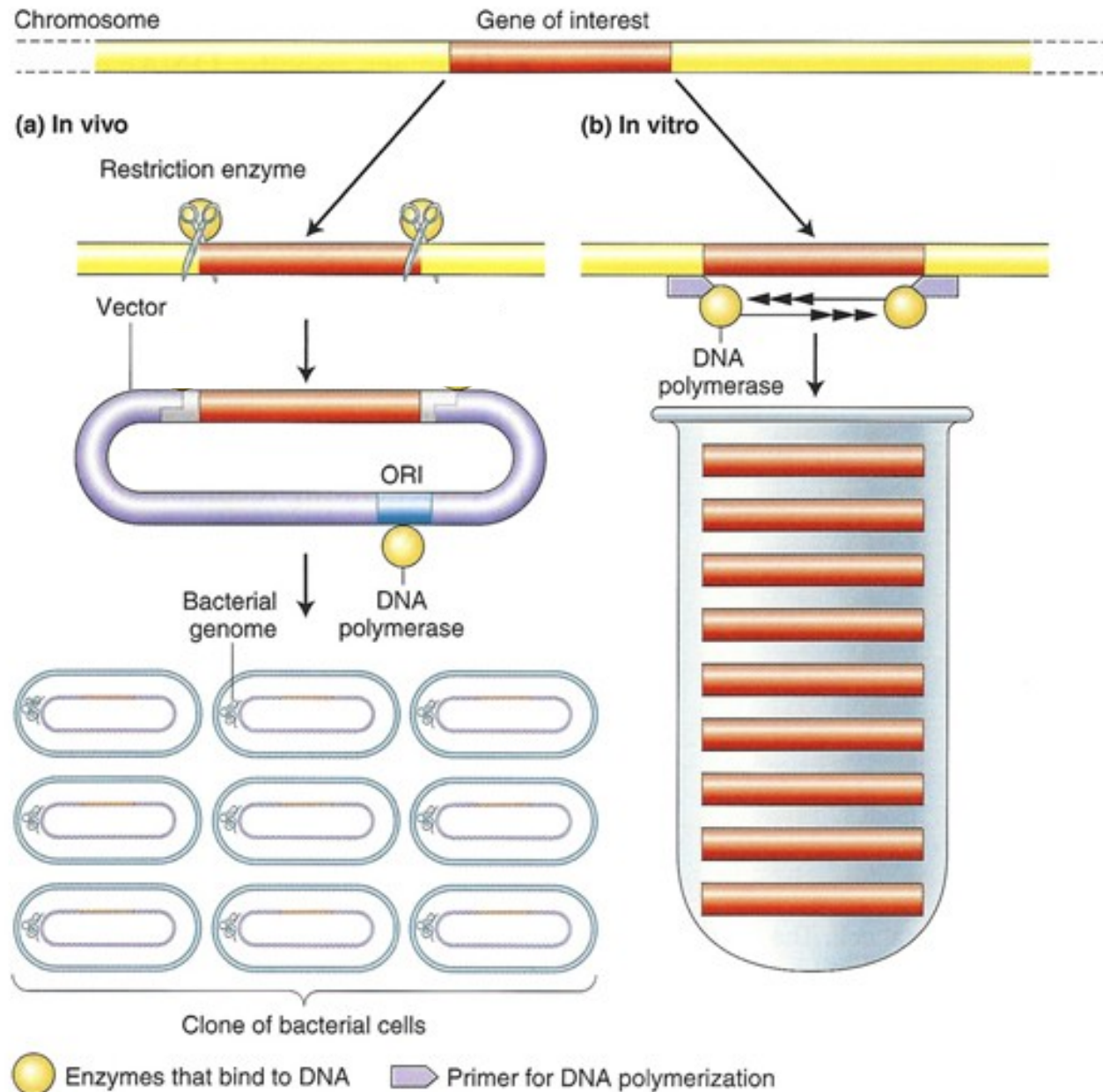
# Amplify DNA

## Polymerase chain reaction - PCR



- 1 Denaturation** at 94-96°C
- 2 Annealing** at ~68°C
- 3 Elongation** at ca. 72 °C

# Cloning vs. PCR



# Amplify DNA

DNA fragments

5 kb-15 kb: plasmids in bacteria

~10 kb: lambda phage-based vectors

Up to 40 kb: fosmids in bacteria

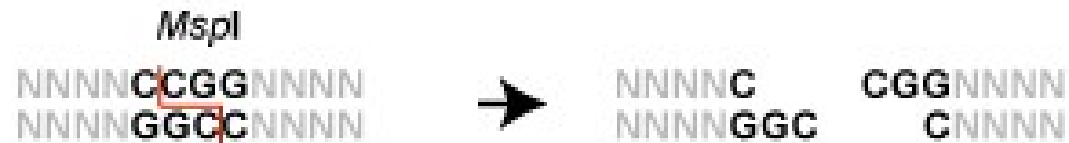
~100-300 kb: bacterial artificial chromosomes (BAC)



# Cut DNA with restriction enzymes

Sites de restriction

Résultats après coupure

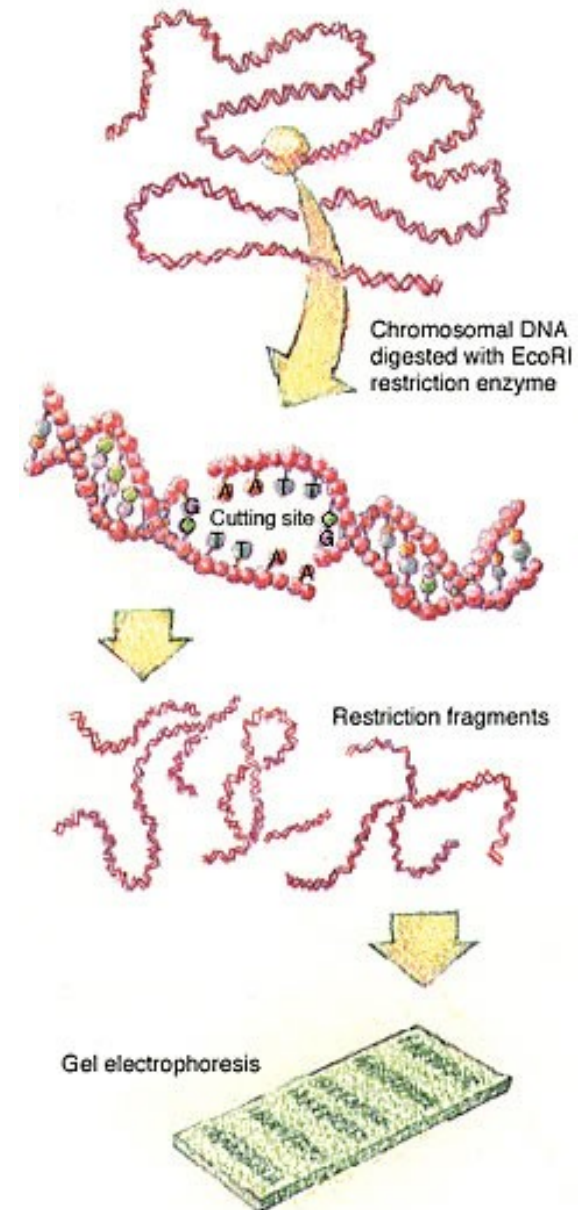
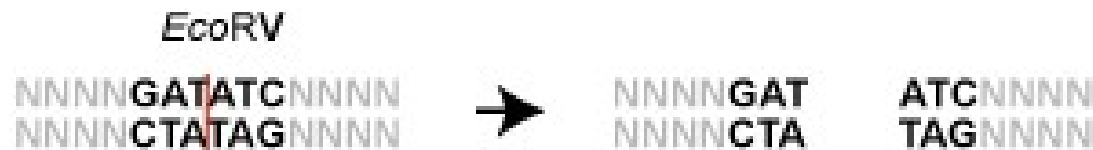
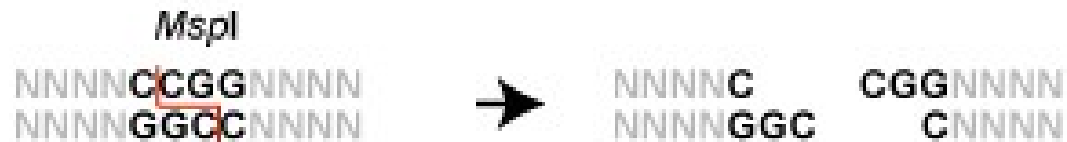


Blunt ends, 3' protruding ends, 5' protruding ends

# Cut DNA with restriction enzymes

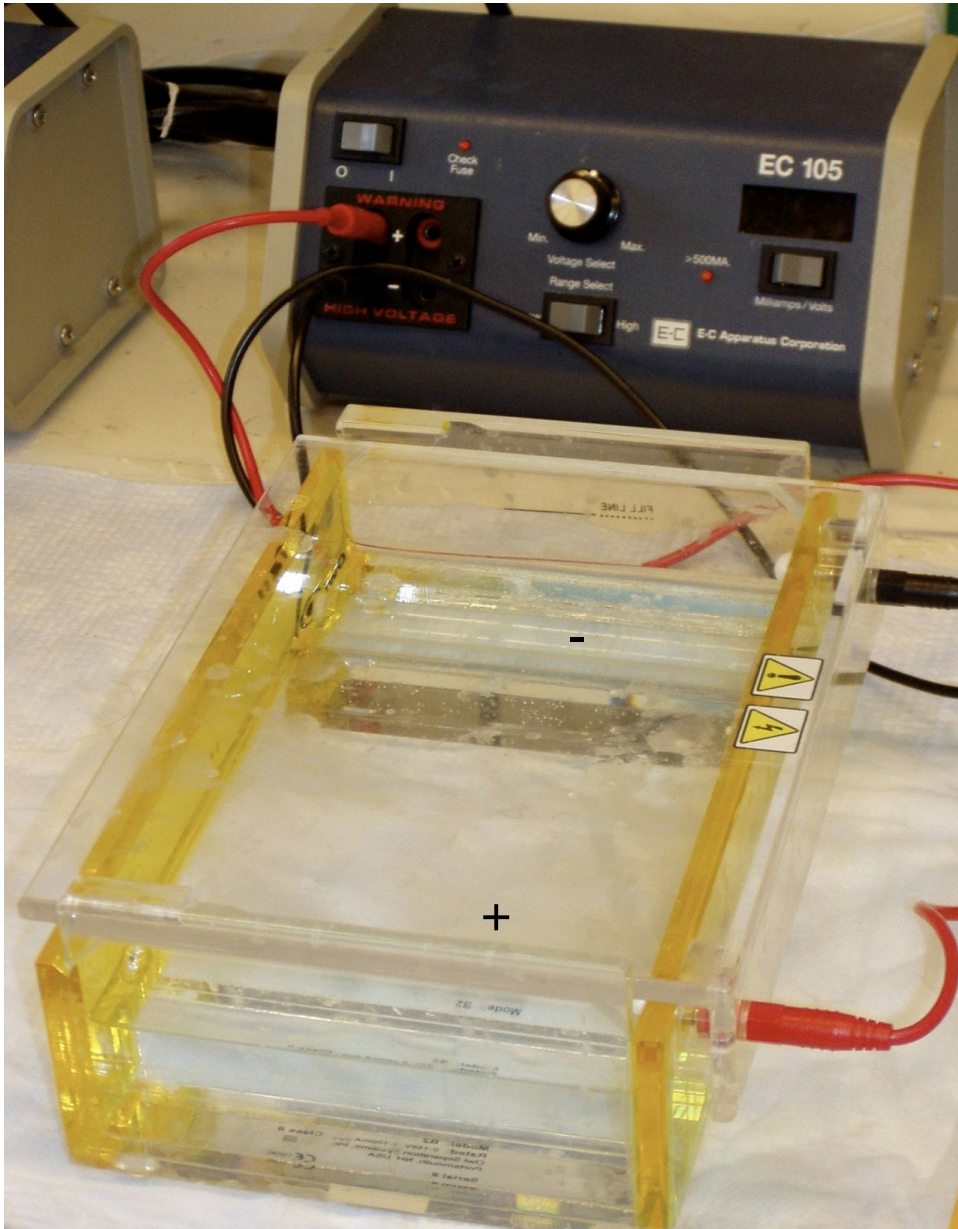
Sites de restriction

Résultats après coupure

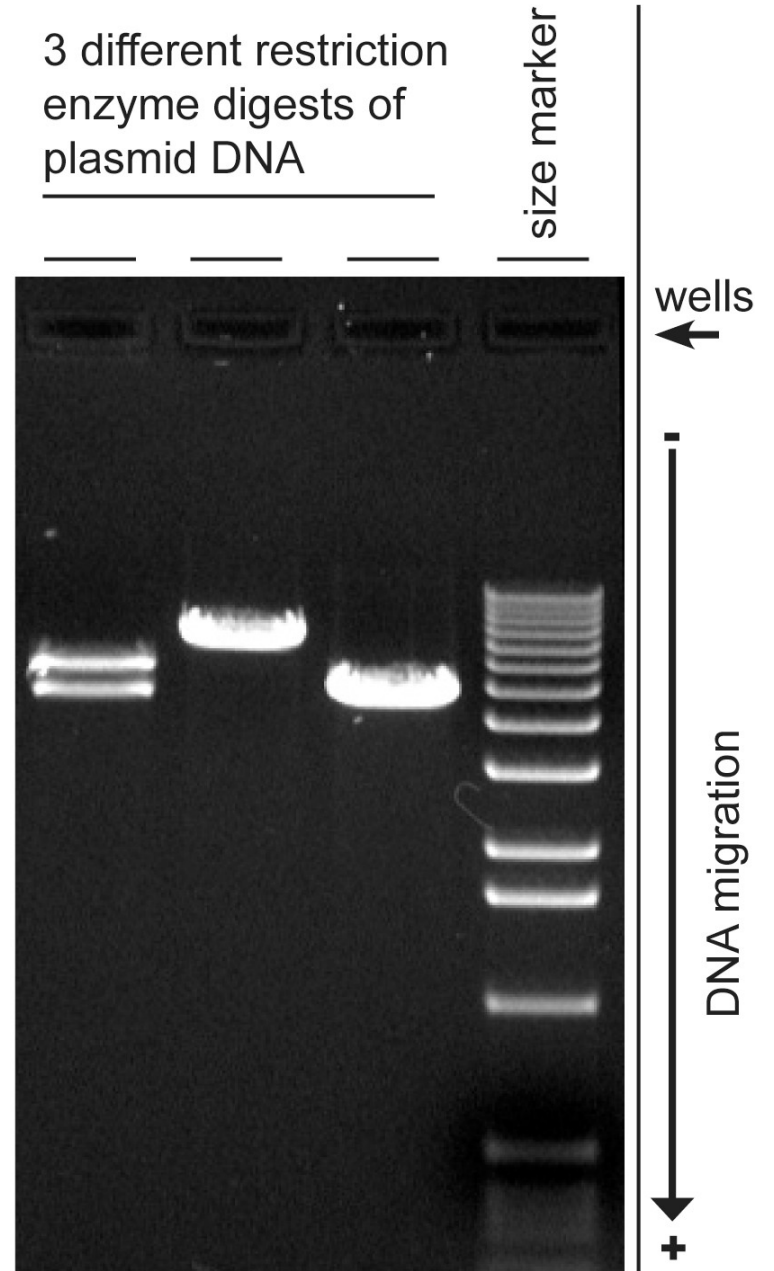


Blunt ends, 3' protruding ends, 5' protruding ends

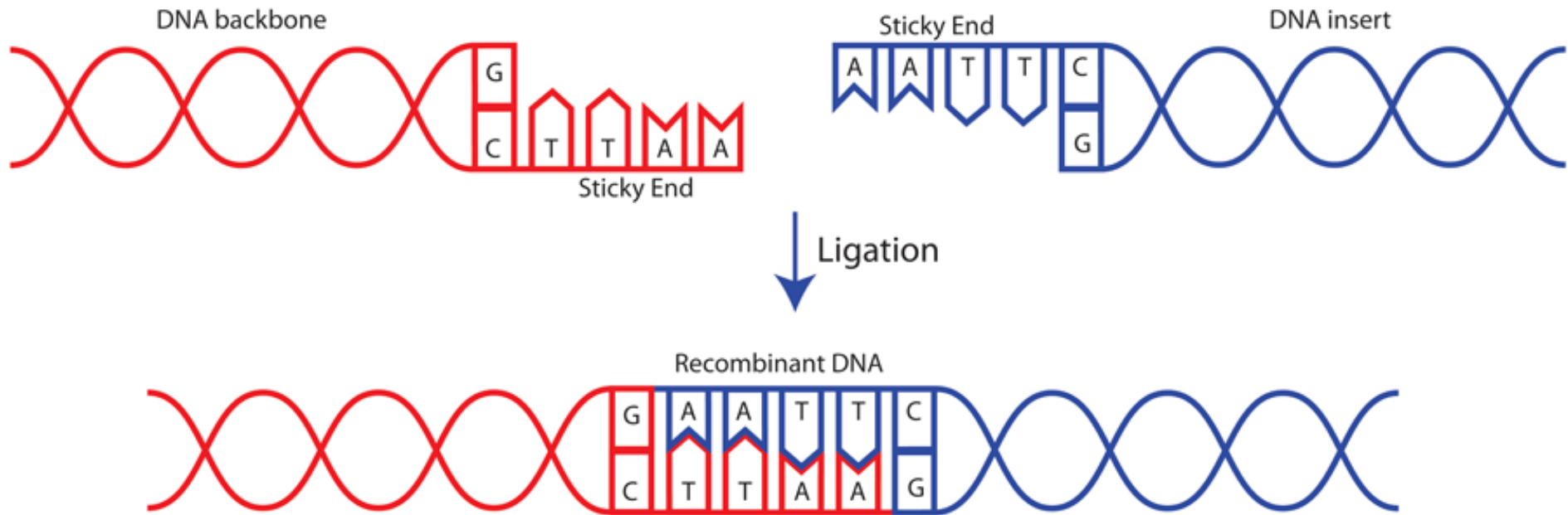
# Examine length of DNA



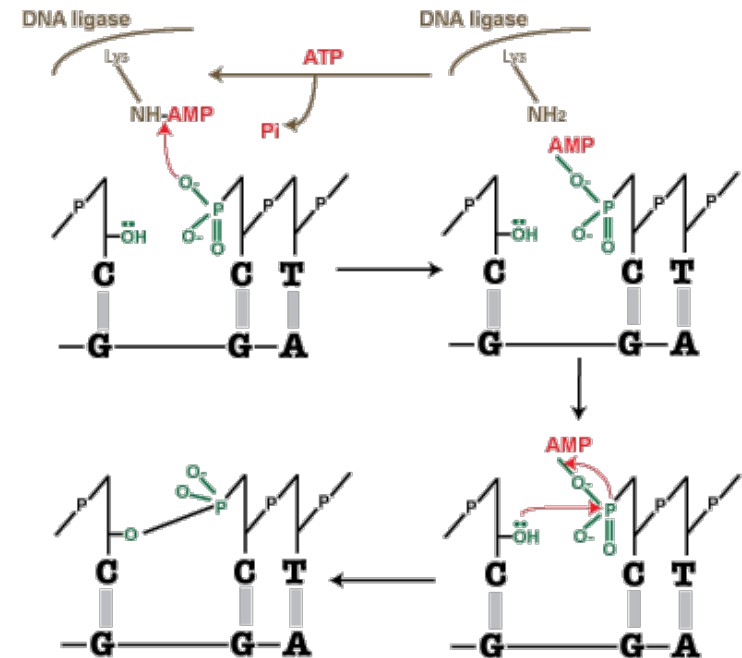
TAE (Tris-acetate-EDTA) buffer



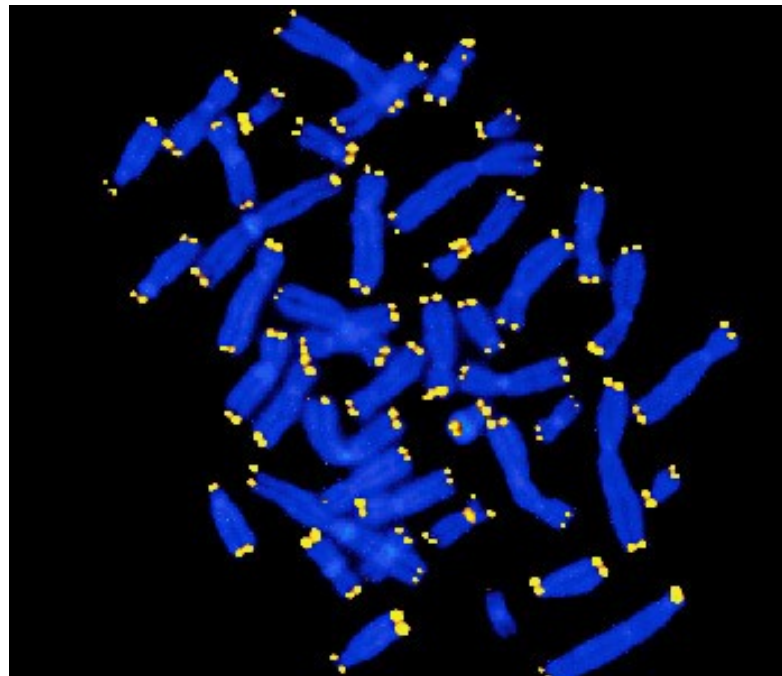
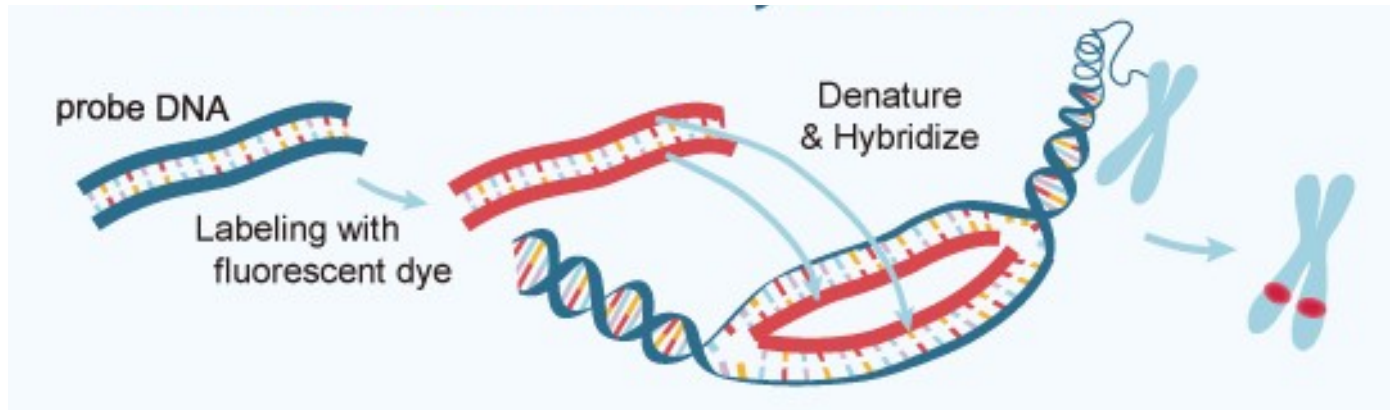
# Ligate DNA



Fragments have to be phosphorylated but only on one strand  
Dephosphorylate the vector to inhibit self-circularization



# Probe DNA: Fluorescent In Situ Hybridization

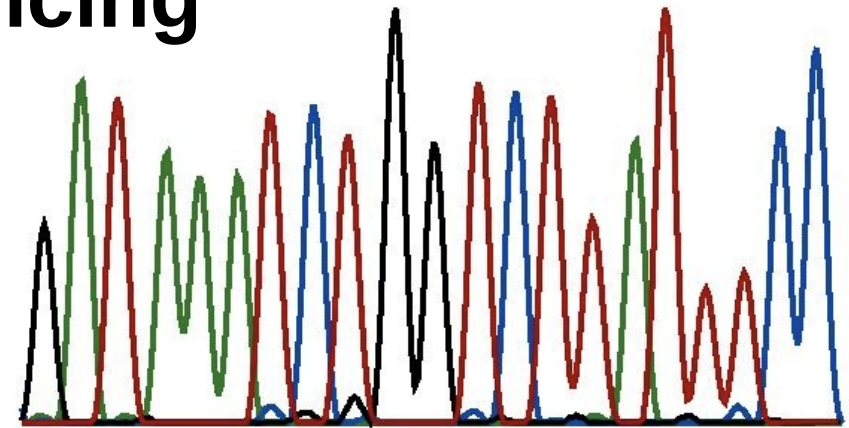


Probes for telomere sequences

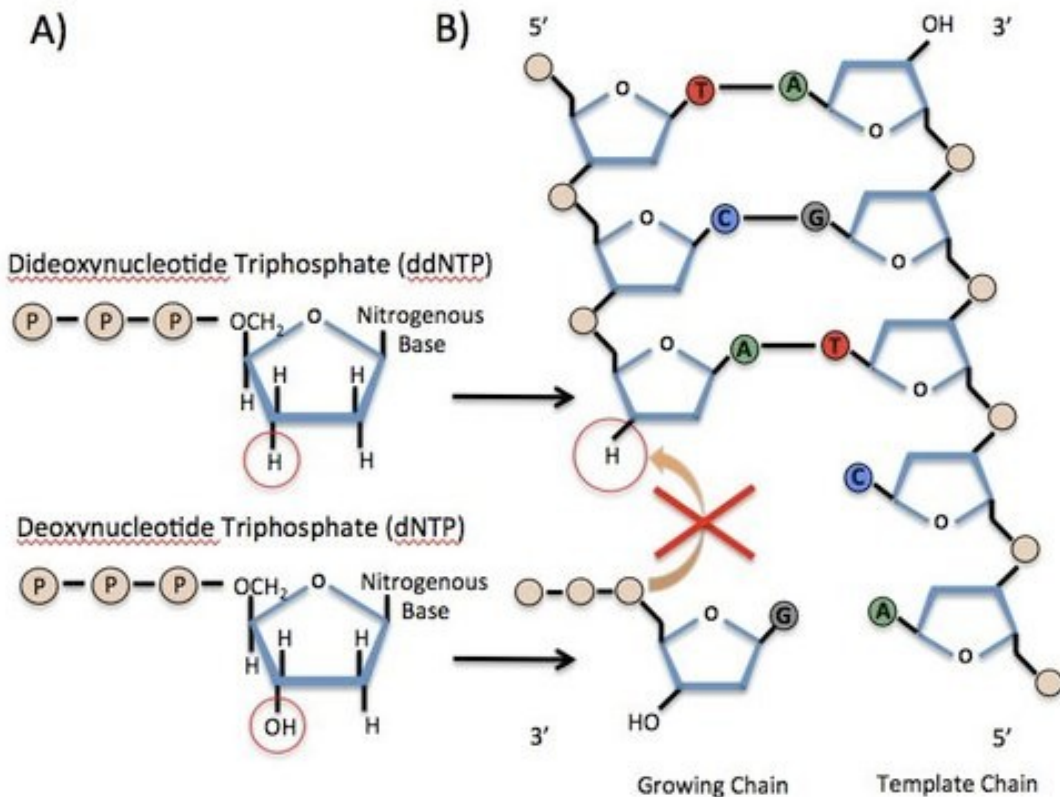
# Sanger sequencing

800 bp long  
 Starts based on oligonucleotide (primer)  
 ~4 euros per reaction

## Dye terminator sequencing

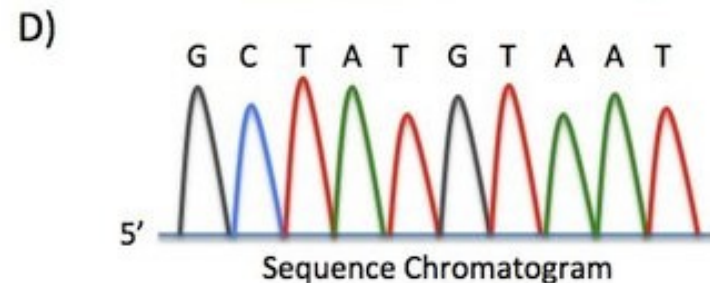


120 130  
 G A T A A A T C T G G T C T T A T T T C C

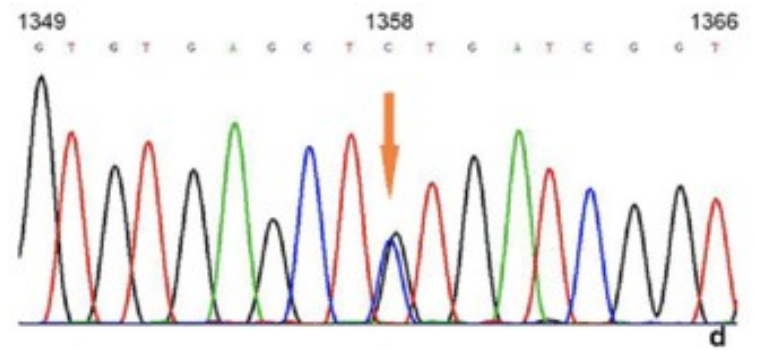
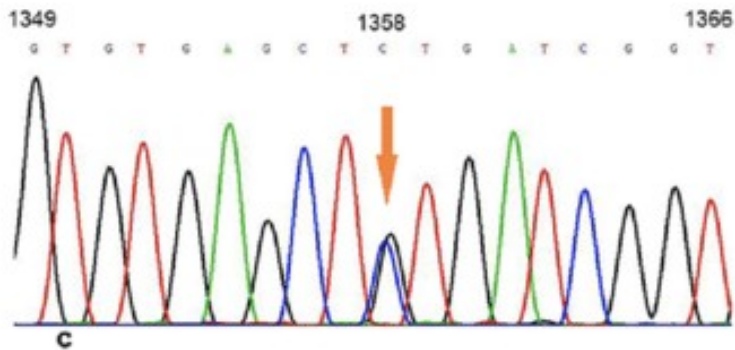
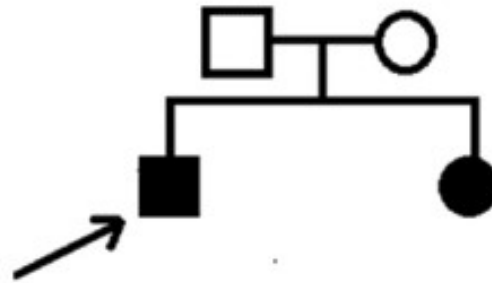
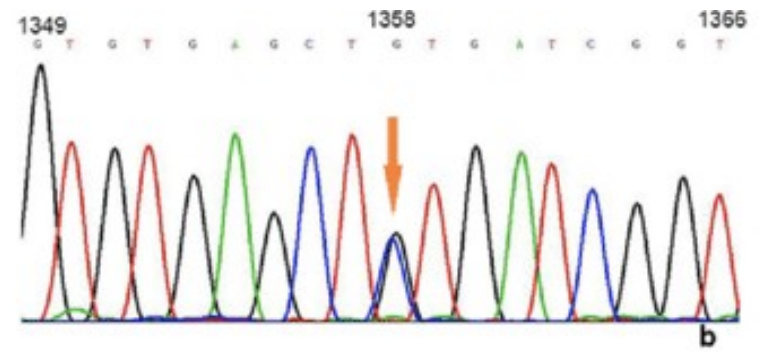
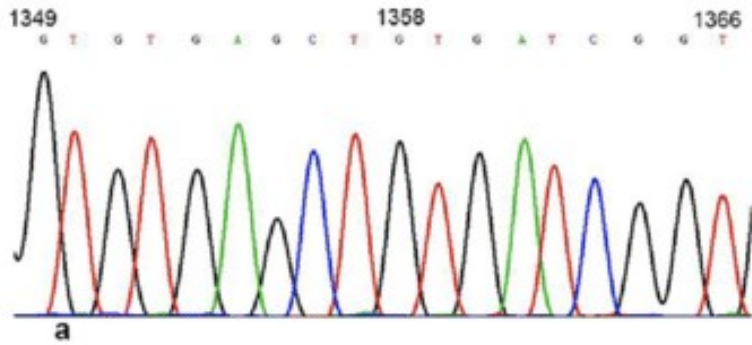


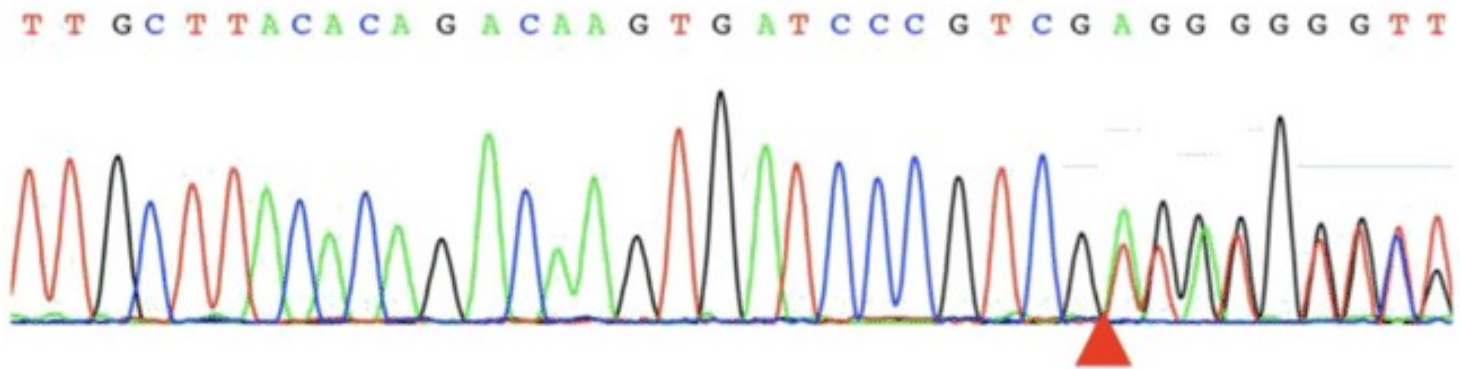
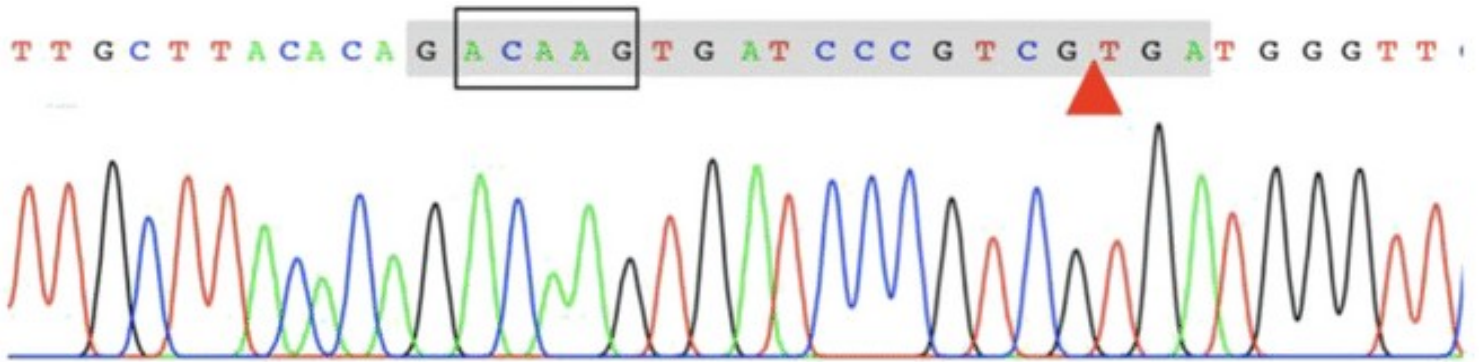
**C)** Template Sequence  
 3' GAGCAAATTCGATACATTATTGT... 5'  
 Primer  
 5' CTCGTTTAAG... 3'

CTCGTTTAAGG — G  
 CTCGTTTAAGGC — C  
 CTCGTTTAAGGGT — T  
 CTCGTTTAAGGGTA — A  
 CTCGTTTAAGGGTAT — T  
 CTCGTTTAAGGGTATG — G  
 CTCGTTTAAGGGTATGT — T  
 CTCGTTTAAGGGTATGTA — A  
 CTCGTTTAAGGGTATGTAA — A  
 CTCGTTTAAGGGTATGTAAT — T



GTGTGAGCTGTGATCGGT

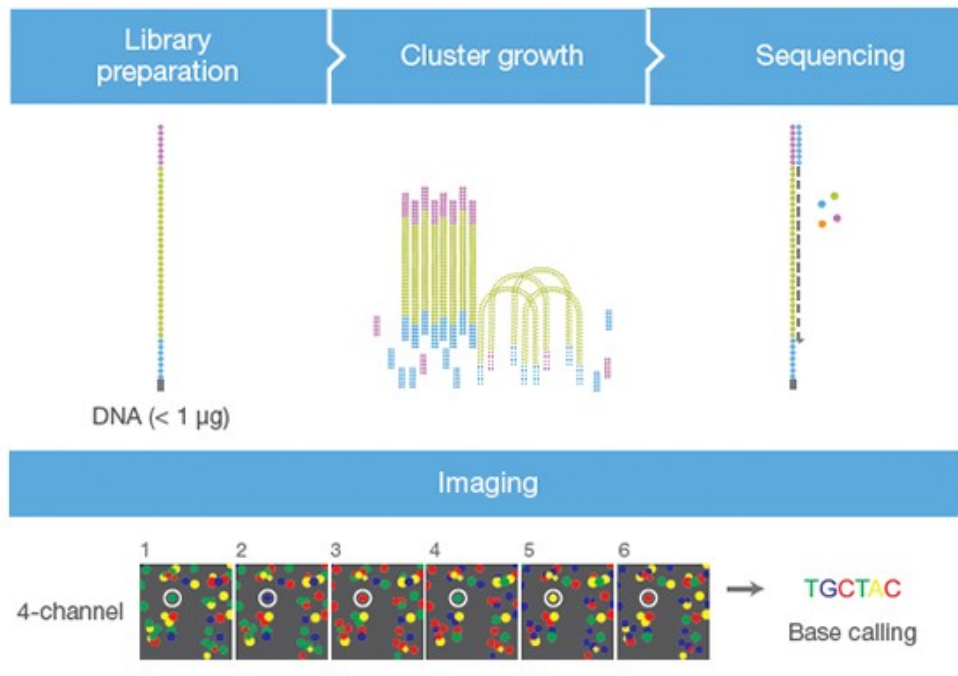






# Illumina sequencing

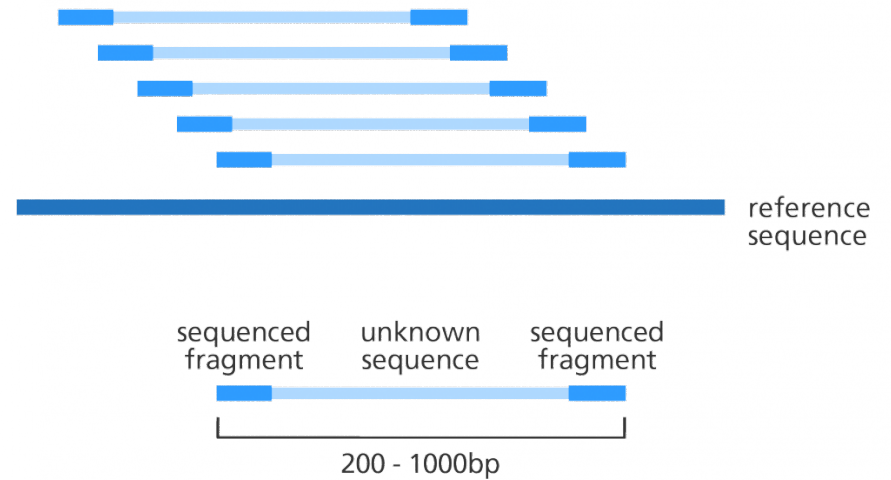
Millions of reads, each ~100 bp long  
Starts at all possible positions  
~500 euros per run



Single-end reads



Paired-end reads



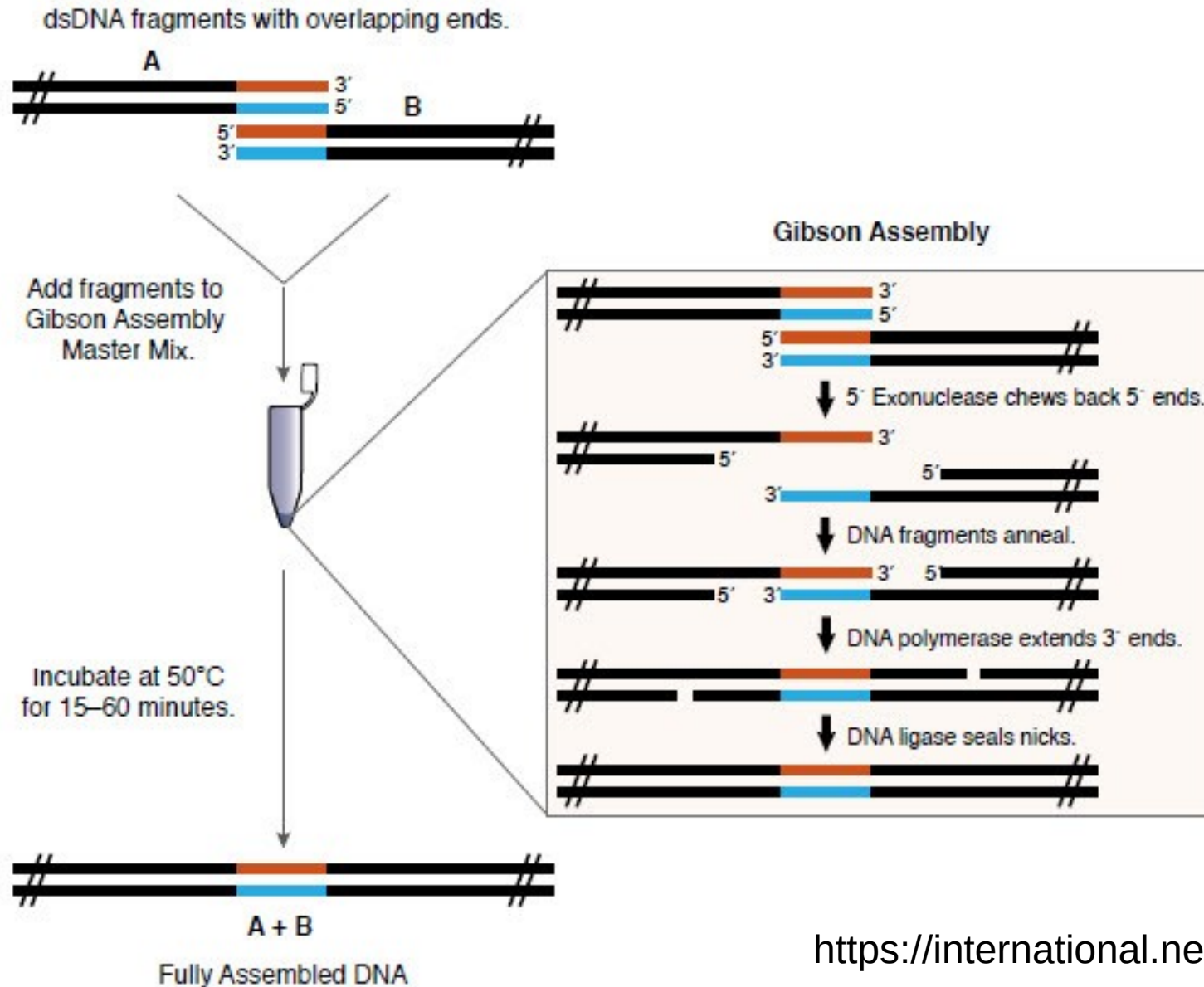
For transcriptome: 2x 75 bp  
For whole genome: 2x 150 bp

<https://www.illumina.com/science/technology/next-generation-sequencing/sequencing-technology/2-channel-sbs.html>



# Recombine DNA: Gibson cloning

Prepare fragments using PCR and special primers



# Synthesize DNA



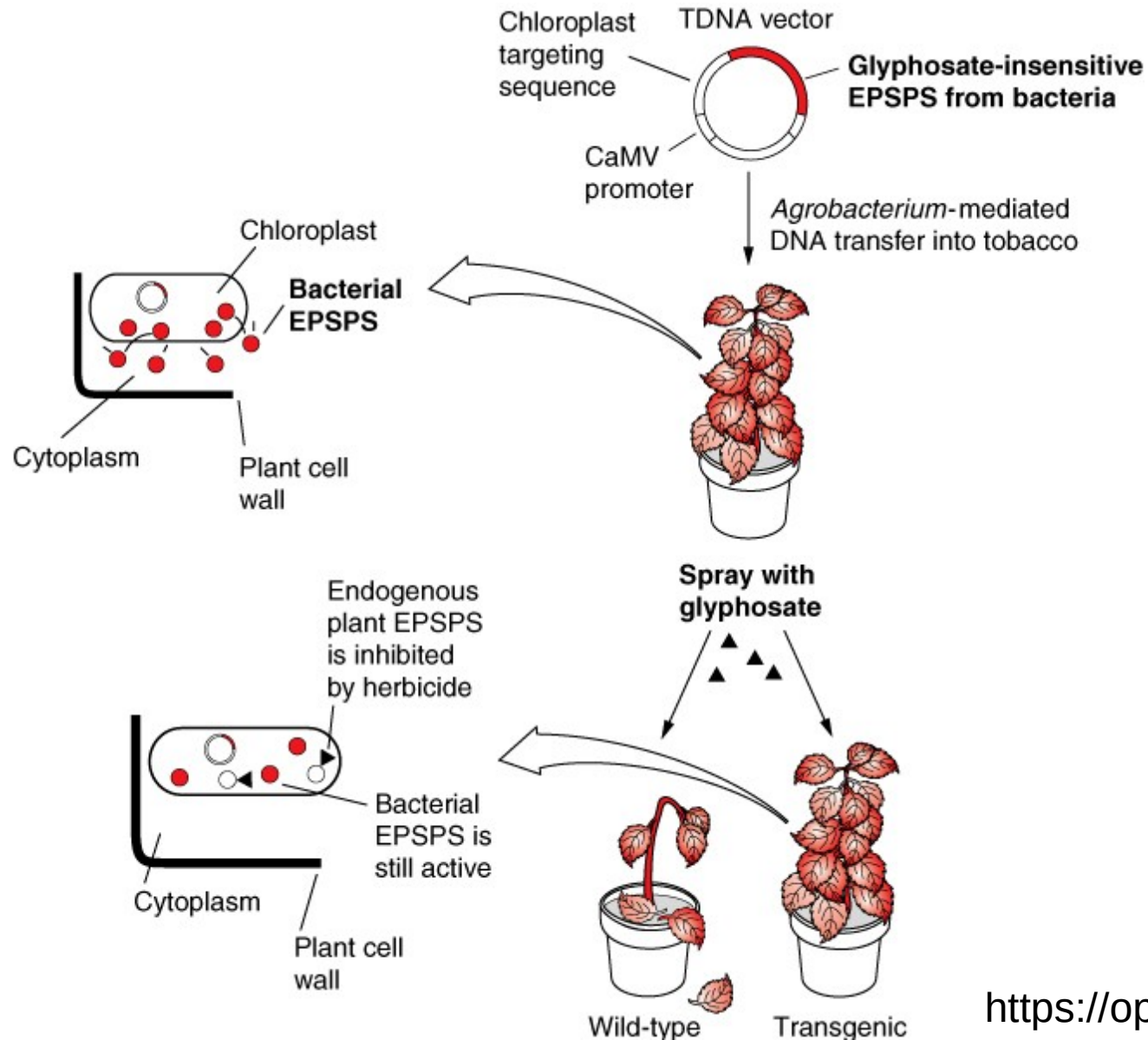
## Gene Synthesis Service Options

Types	Gene Length	Price (No hidden charge promise) †	Starting Turnaround Time (Business Days) *	Starting Turnaround Time with Plasmid Prep Service (Business Days)
Standard Gene Synthesis <i>Guaranteed</i>	≤ 8 kb	View your discounted price online in as short as 1 minute	8	10
Fast Gene Synthesis <i>Guaranteed</i>	≤ 5 kb		7	9
Rush Gene Synthesis <i>Guaranteed</i>	≤ 4 kb		4 <i>US Manufacture</i>	6 <i>US Manufacture</i>
GenPlus HT Gene Synthesis	≤ 3 kb		18	20
GenPlus Economy Gene Synthesis	≤ 8 kb		15	17
GenBrick® Gene Synthesis	> 8 kb		23	25

# Introduce foreign DNA

most widespread transgenic crop in 2005-2015 = soybean resistant to glyphosate

In tobacco:



# What can we do with DNA ?

## **Extract, purify**

## **Make more**

Amplify

Clone

Synthesize

## **Examine**

Quantify

Examine length

Stain, probe

Sequence

## **Modify**

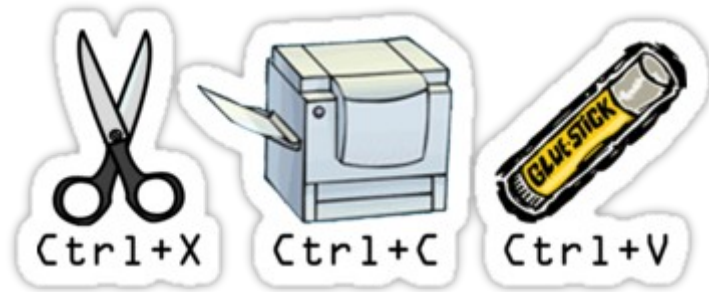
Cut

Ligate

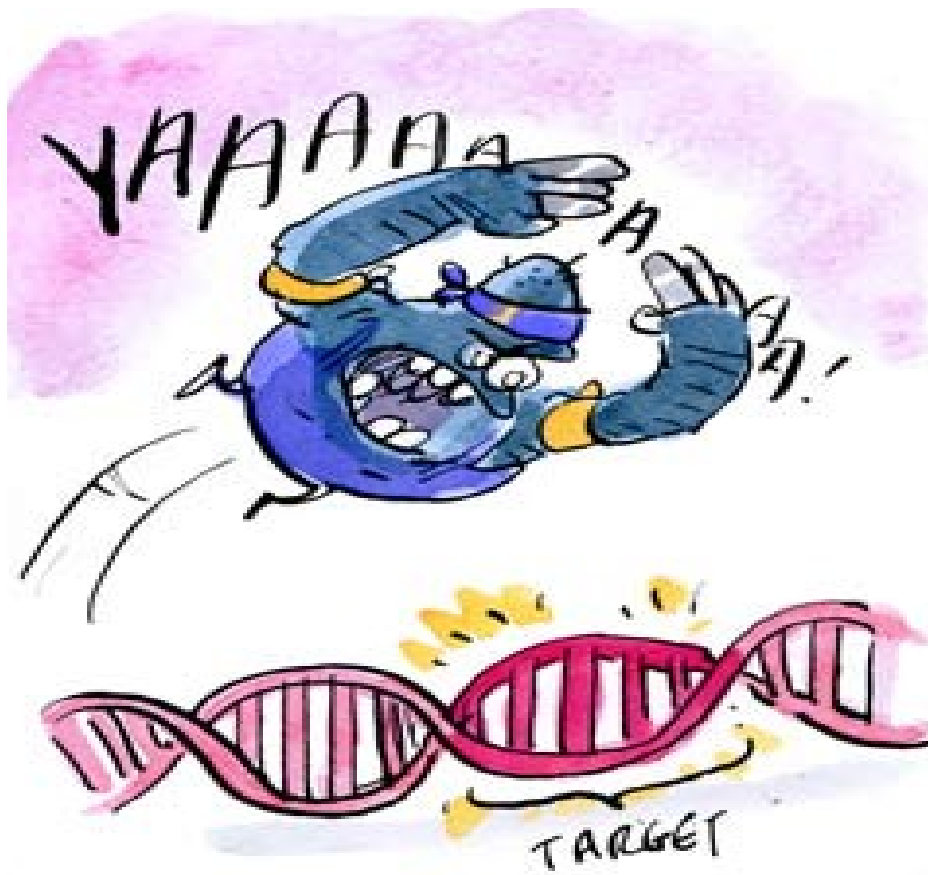
Recombine fragments

Introduce foreign DNA

Mutate



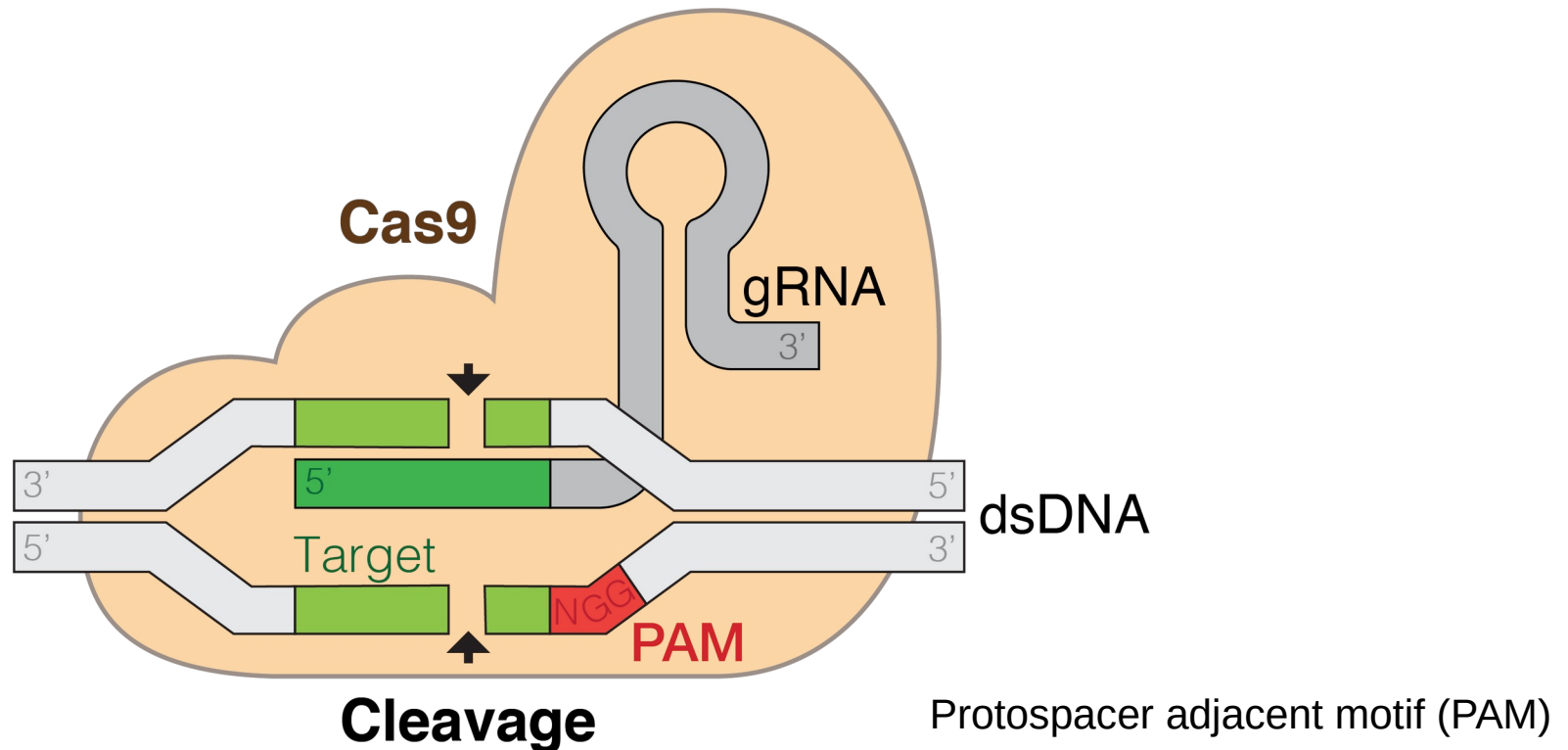
# CRISPR



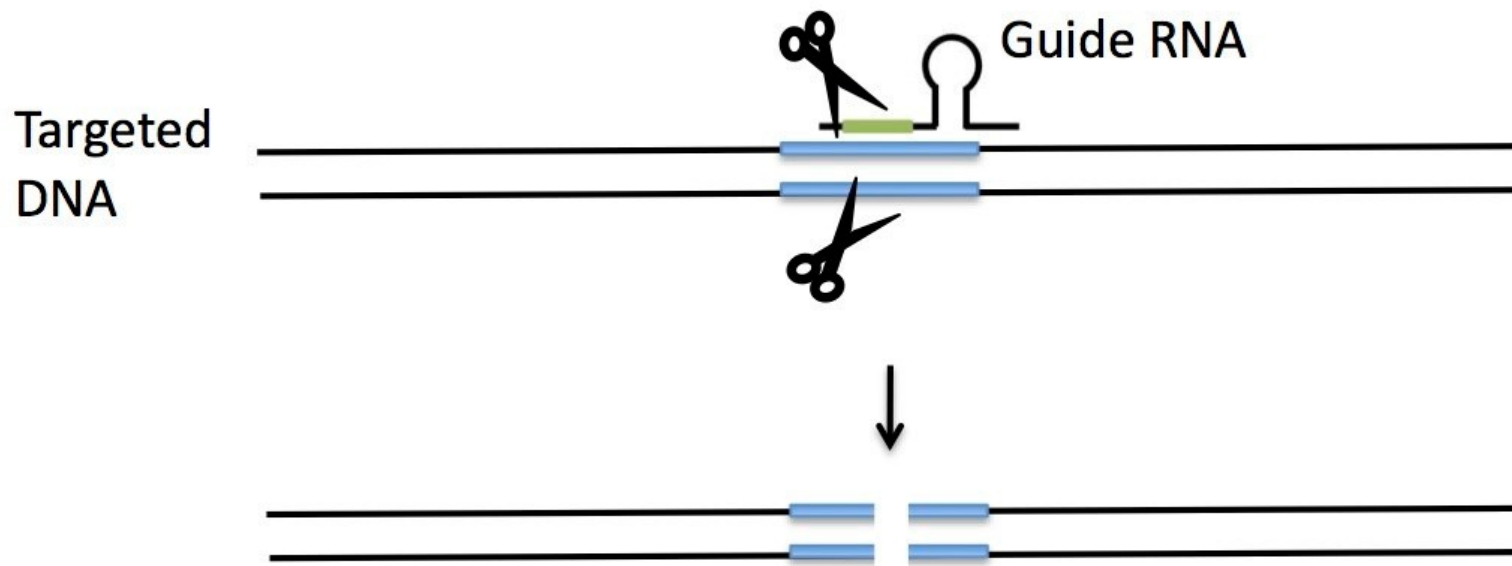
Marion Montagne



**CRISPR** = clustered regularly interspaced short palindromic repeats  
= family of DNA sequences present in bacteria and used to detect and destroy virus DNA



Can recognize and cut a specific DNA sequence (recognized by guide RNA)  
More versatile than restriction enzymes, Zn finger nucleases and transcription activator-like effector nucleases (TALENs).

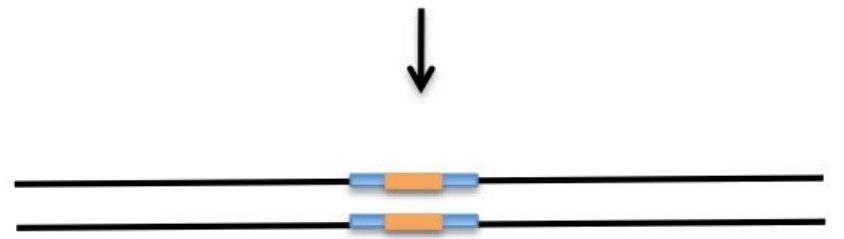
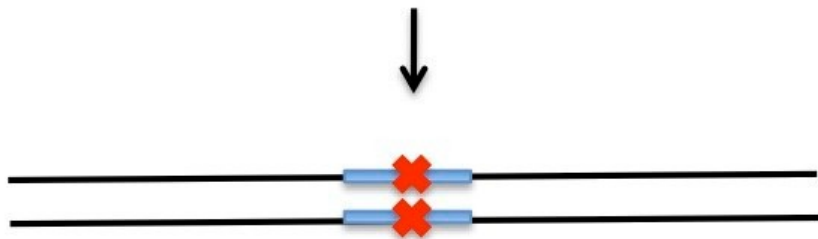
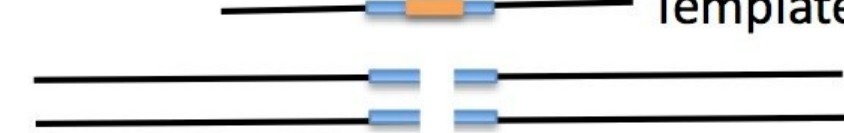
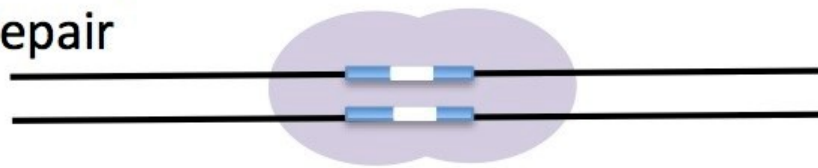


Gene Silencing

Gene Editing

Attempted Repair

Repair Template



Gene is disrupted

Gene has a new sequence

# Creating mutants with CRISPR/Cas9

GAGTTCTACAGCGTGAACCACATCAACCAGACGTACGAGTTTGTGCAGCGGATGCG	Wild type
GAGTTCTACAGCGTGAACCACATCAACCAGACGTACGAGTTG--CAGCGGATGCG	Deletion
GAGTTCTACAGCGTGAACCACATCAACCAGACGTACGAGT-----AGCGGATGCG	
GAGTTCTACAGCGTGAACCACATCAACCAGACGTACG-----CAGCGGATGCG	
GAGTTCTACAGCGTGAACCACATCAACCAGACGTA-----CAGCGGATGCG	
GAGTTCTACAGCGTGAACCACAT-----GCGGATGCG	Insertion
AGTTCTACAGCGTGAACCACATCAACCAGACGTACGAGTTTGTGACAGCGGATGCG	
TACAGCGTGAACCACATCAACCAGACGTACGAGTTTGTGGCTTTAAAGCGGATGCG	
CAGCGTGAACCACATCAACCAGACGTACGAGTTTGTGCAAGGAAACTGCGGATGCG	

*Agraulis vanillae*

*dorsal*

*ventral*



**Wild-type**

**mutant  
*optics* CRISPR**

**Wild-type**

**mutant  
*optics* CRISPR**



normal

mutant  
*WntA* CRISPR

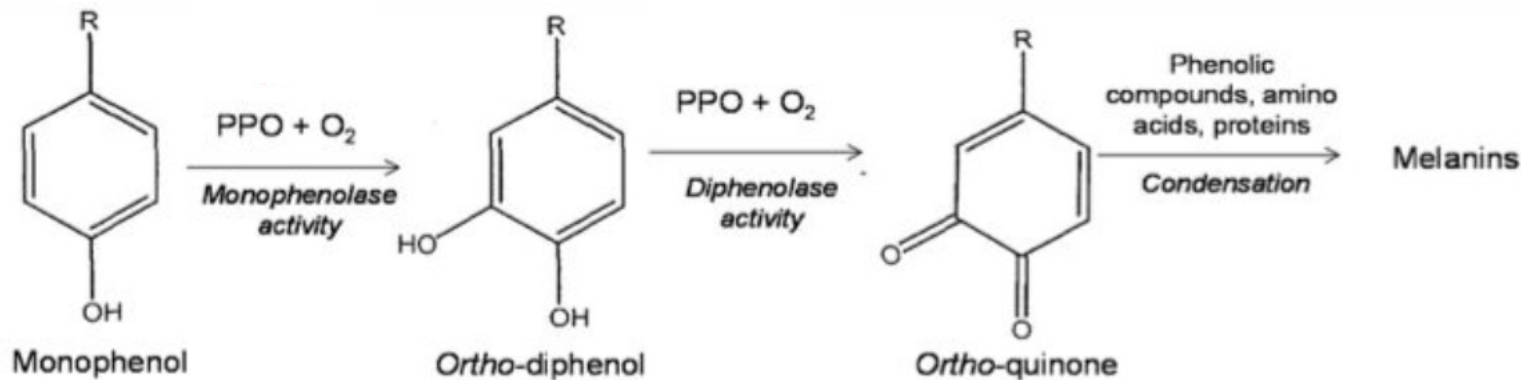
# The first CRISPR food



“Animal and Plant Health Inspection Service (APHIS) has concluded that your CRISPR/Cas9-edited white button mushrooms as described in your letter do not contain any introduced genetic material. APHIS has no reason to believe that CRISPR/Cas9-edited white button mushrooms are plant pests”

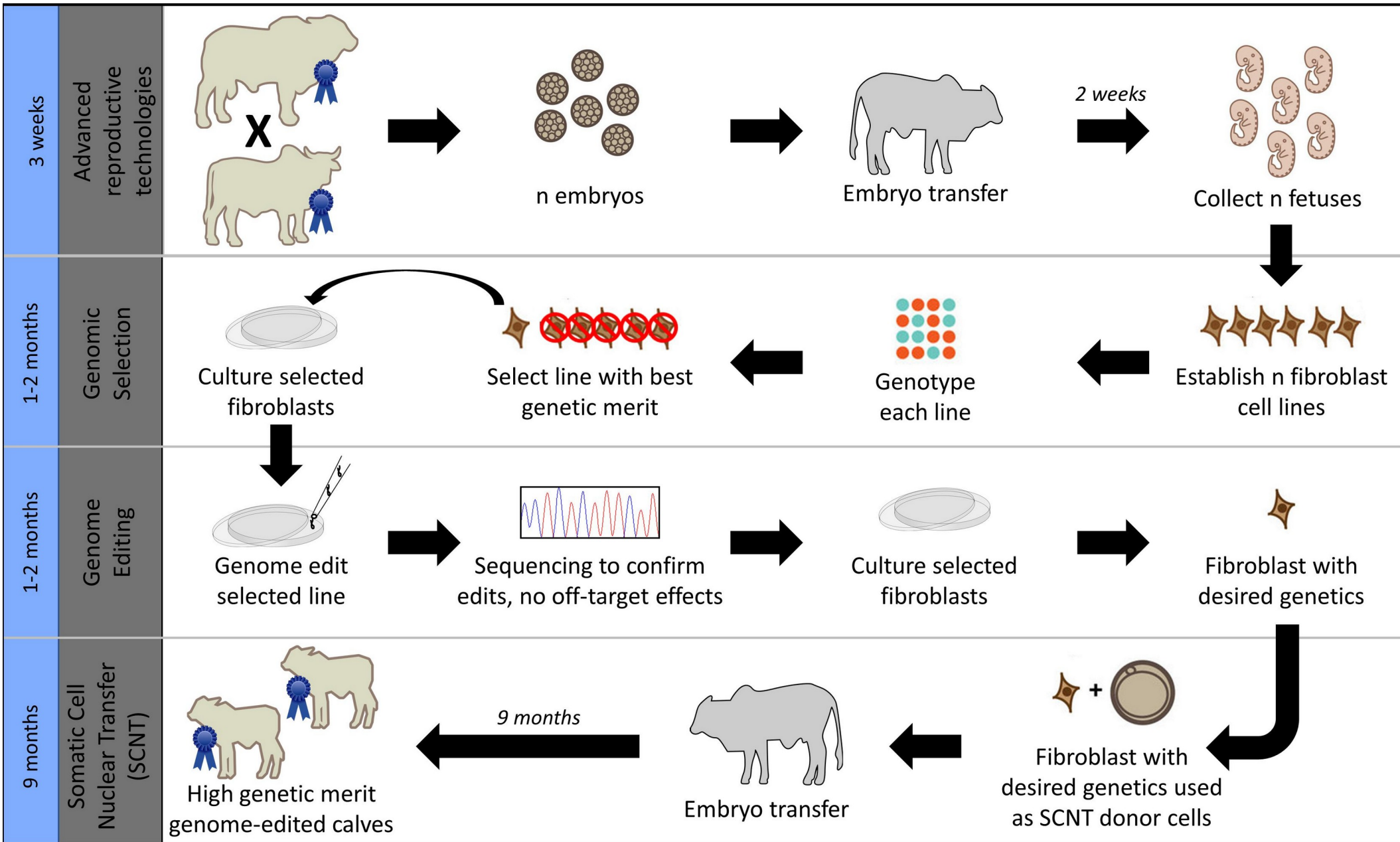
Deletion in 1 of the 6 polyphenol oxidase genes  
Reduction of 30% polyphenol oxidase activity

April 2016



**FDA does not consider CRISPR-edited food as GMO**

# Cattle advanced reproductive technologies: somatic cell nuclear transfer cloning (SCNT), embryo transfer (ET)

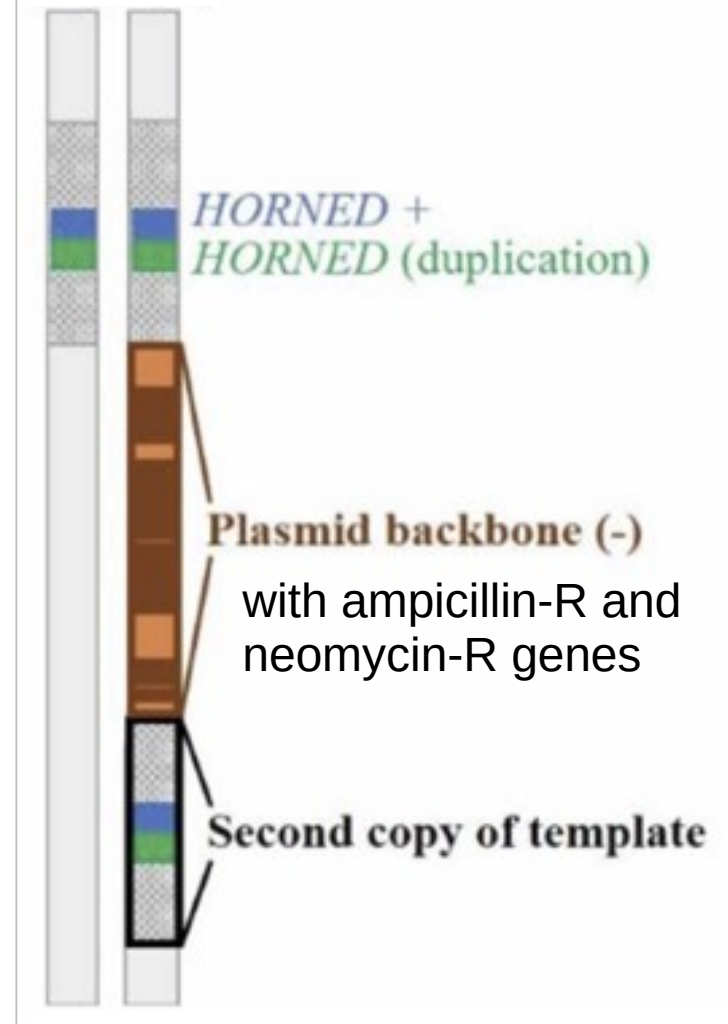


# CRISPR-edited hornless cows were supposed to be exempt of transgenes



Expected Sequence

Obtained Sequence





# Regulation about CRISPR-edited organisms in Europe and the US

Augustin  
Martin



## In the US

- For Crops : No legal framework : **co-regulation** by the USDA, FDA and EPA
- What matters is the **final characteristics** of the genetically edited organism
- 2020 USDA-APHIS' "SECURE" initiative : **disregulation** of gene-edited organisms with mutations that could have naturally occurred (CRISPR-induced SNP or addition of an endemic gene)
- Considers this type of gene editing to be an acceleration of what is **naturally occurring**



« The newest of these methods, such as genome editing, **expand traditional plant breeding tools** (...) potentially saving years or even decades in bringing needed new varieties to farmers. »

*Secretary of State Perdue for the USDA*

- CRISPR-edited animals are evaluated as **animal drugs** by the FDA : strict safety evaluation



## In Europe

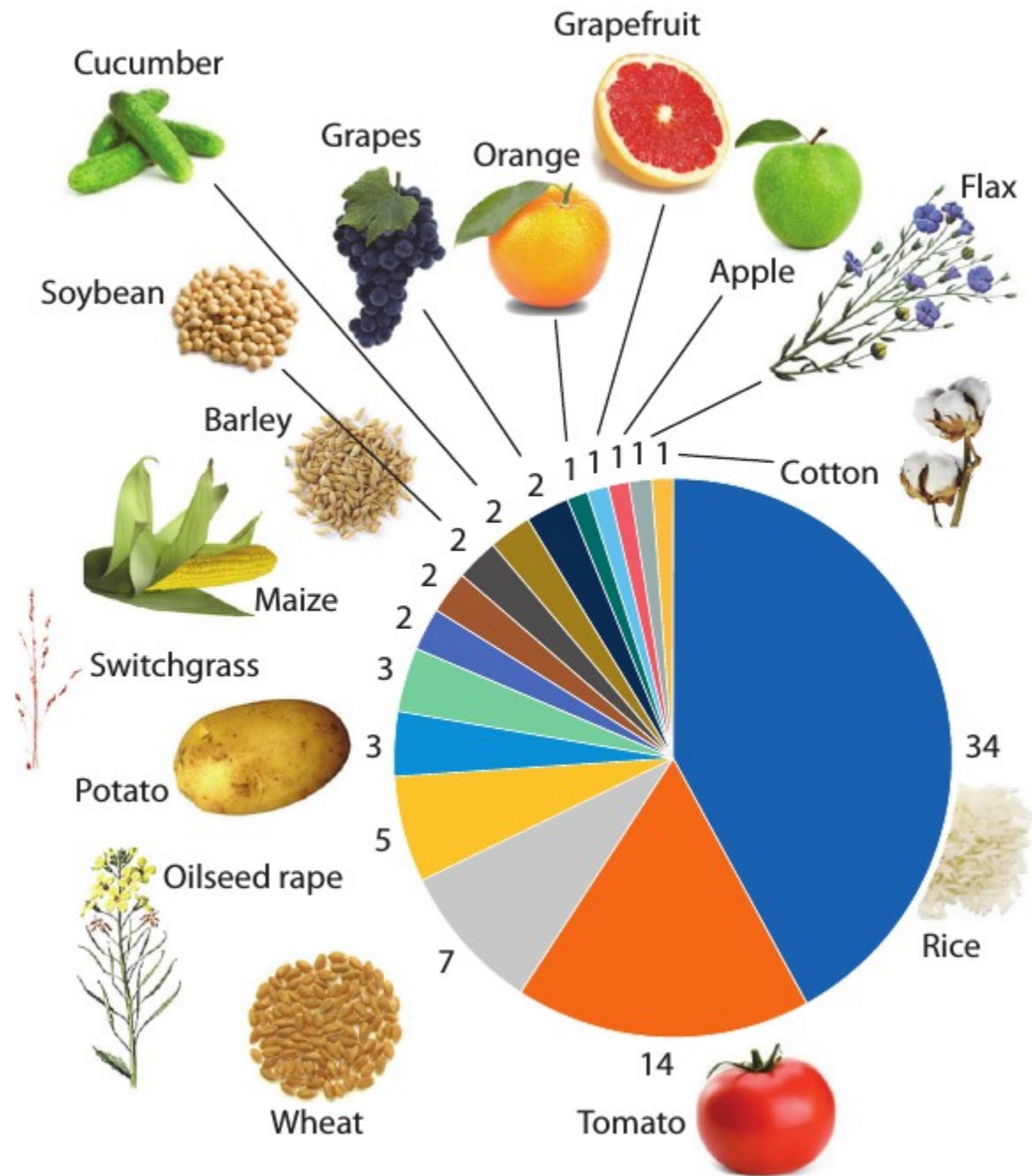
- Major judgment : ECJ "Confédération paysanne" 2018 ruling



« The Court considers that the **risks linked to the use of these new mutagenesis techniques might prove to be similar** to those that result from the production and release of a **GMO through transgenesis** »

- What matters is the **process** through which crops are obtained
- Concluded that **mutagenesis and transgenesis** are similar according to
  - Their potential danger
  - The rate of production
  - Their action of "denaturing" the genome
- CRISPR-edited crops and animals: regulated as **GMOs**
- Considers gene editing to be an **unnatural** modification of the genome

# Numerous plants modified using CRISPR



# The first CRISPR-edited foods are not “to save the world”



## Herbicide-tolerant oilseed rape

(in 2015, the German authorities authorized the release of oilseed rape developed by the US company CIBUS without being regulated as genetically engineered.)

**Soybean with modified fatty acid composition**

**Potato with improved storage capacity at cool temperatures**

**“Waxy” maize with a modified starch composition**

**Herbicide-tolerant flax**

**Sweeter-tasting strawberries**

**Seedless tomatoes**

Ueta et al. 2017

Making plants that are resistant to drought or salt requires exogenous genes

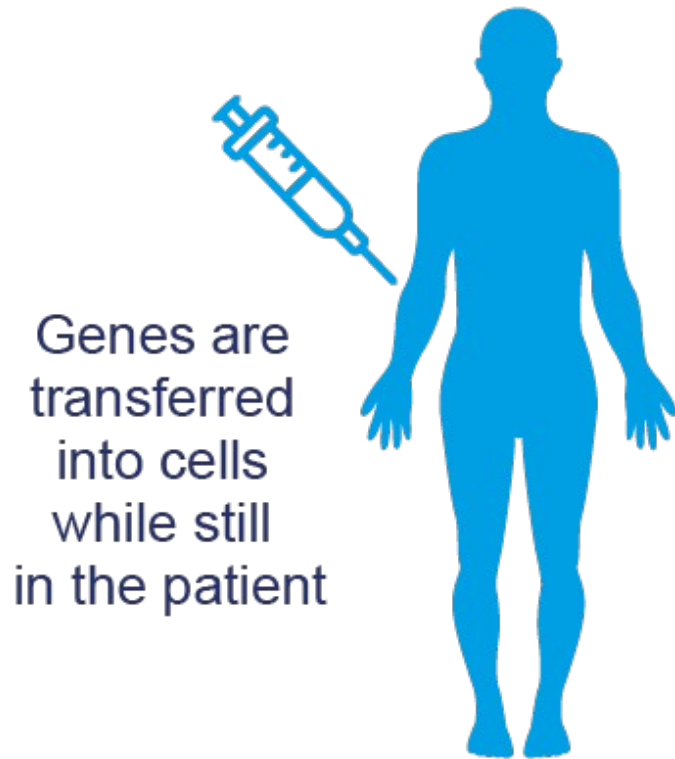
The coding loci of evolution and domestication: current knowledge and implications for bio-inspired genome editing

Virginie Courtier-Orgogozo<sup>1,\*</sup> and Arnaud Martin<sup>2</sup>

J. Exp. Biol.

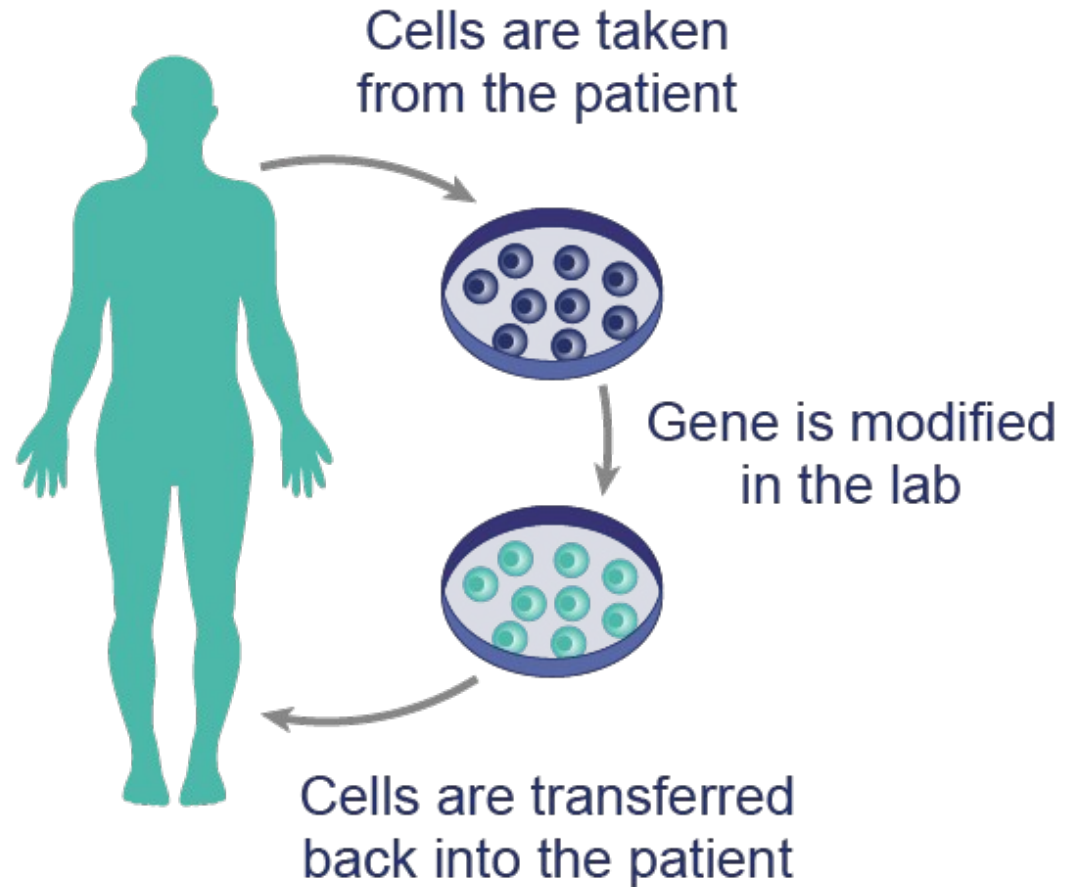
# Ongoing CRISPR clinical trials

## In Vivo



Retina disease  
First CRISPR trial in March 2020  
BRILLANCE trial, Editas

## Ex Vivo



Sickle Cell Disease  
Beta-Thalassemia

# Fundamental research is important

bacteria *Thermus aquaticus*



1969 → Taq-polymerase  
to amplify DNA

bacteria *Haemophilus influenzae*



1970 → Restriction enzymes  
To cut DNA

jellyfish *Aequorea*



1992 → Fluorescent  
proteins

bacteria *Streptococcus pyogenes*



2012 → CRISPR

# **CRISPR-based gene drive**

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# What is gene drive?

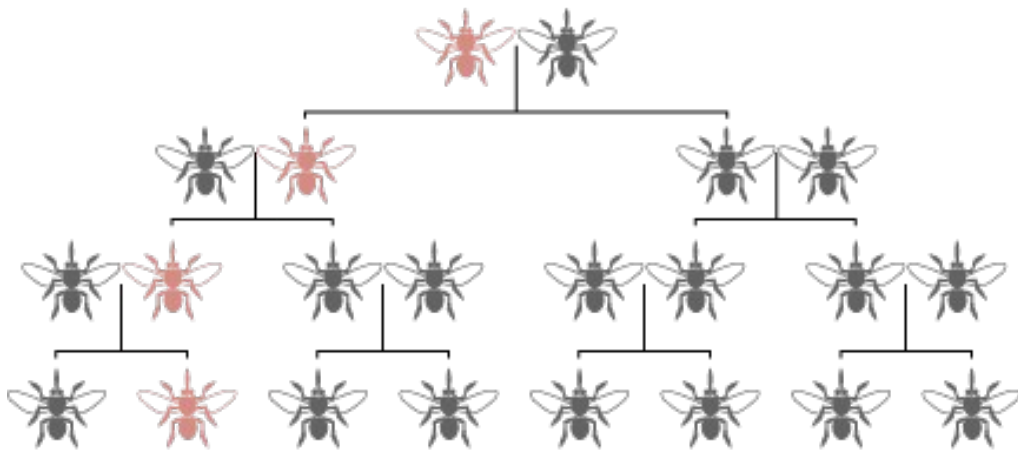
A novel biotechnology under development which aims to bias inheritance and control disease vectors, invasive species and other pests.

Public health, agriculture, conservation biology

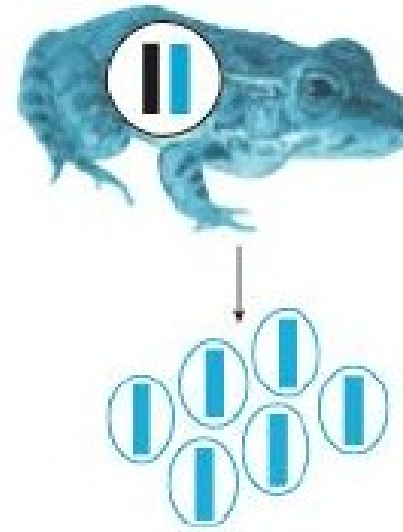
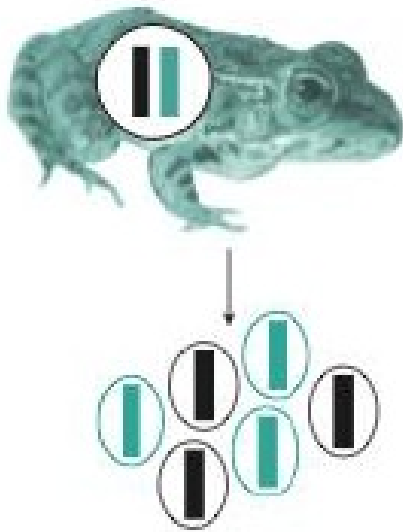
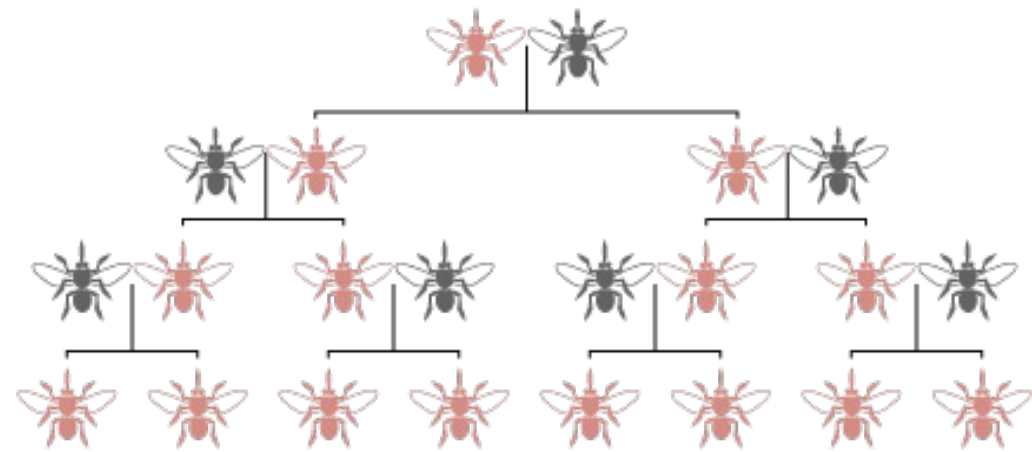
*“A natural phenomenon in several species”*

(Austin Burt, 2020)

## Normal reproduction

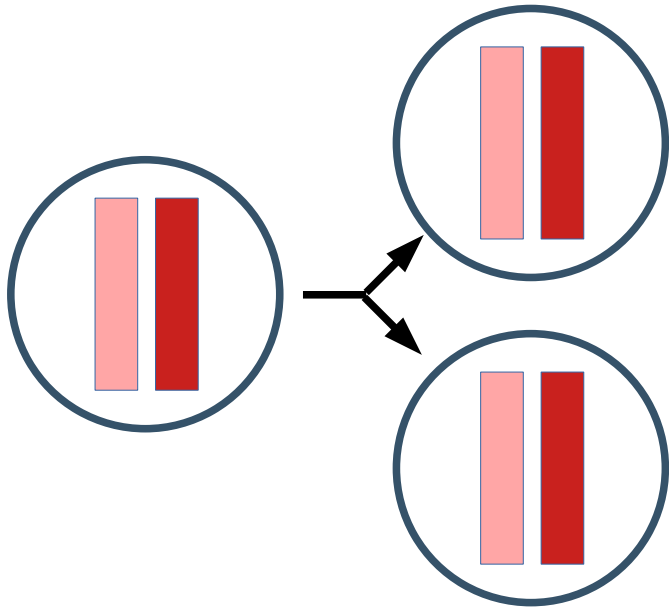


## Reproduction with gene drive

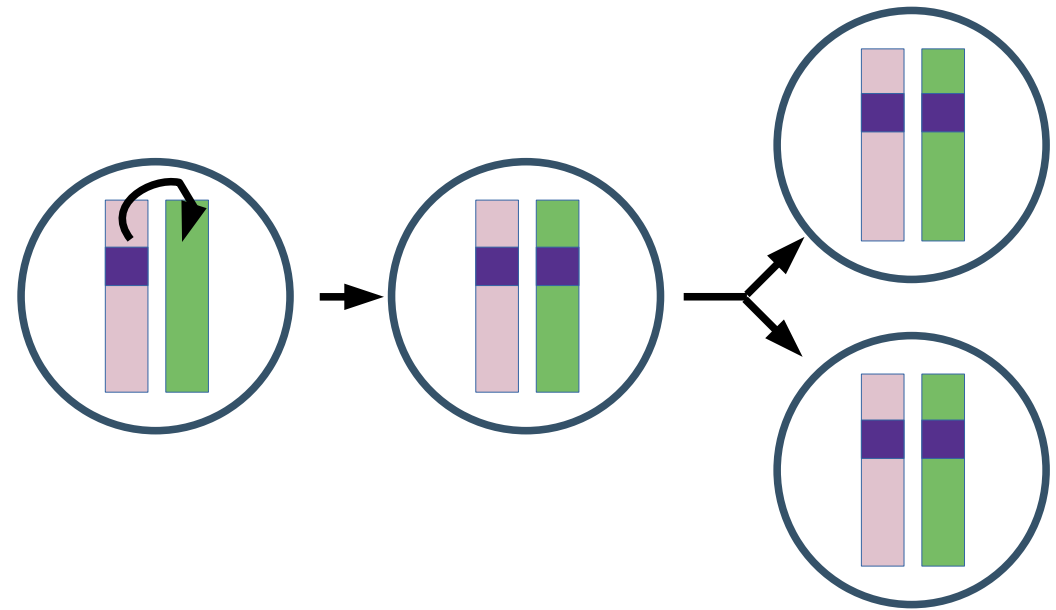




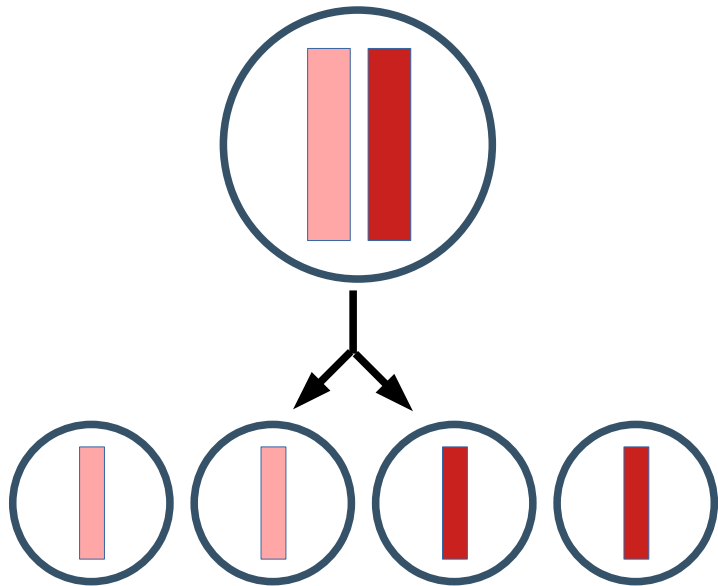
Cell division (mitosis)



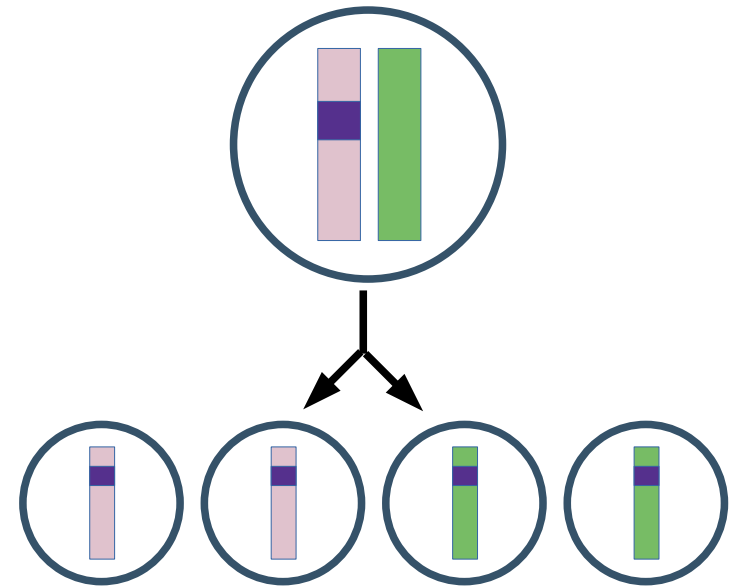
Gene drive

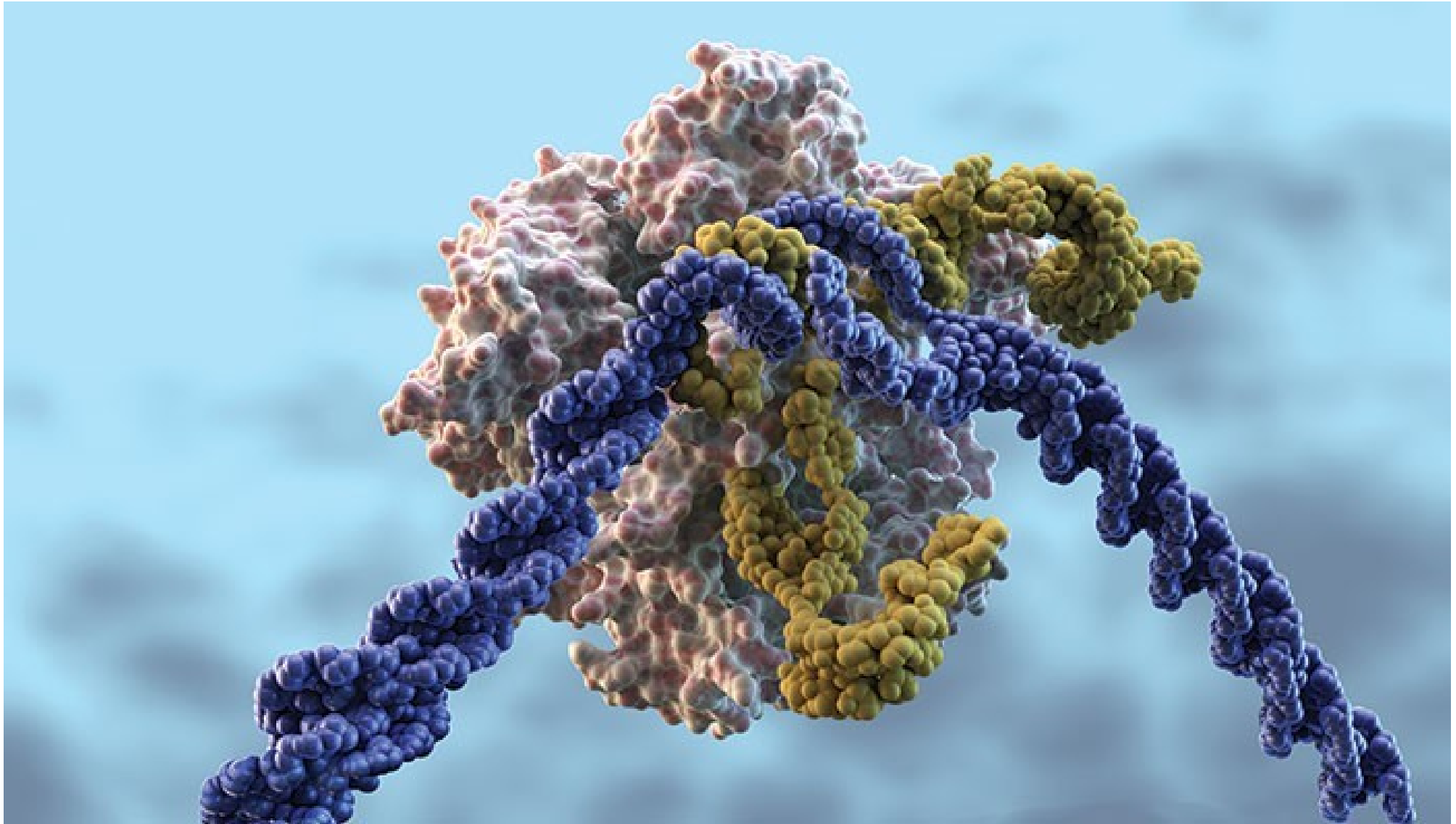


Normal reproduction

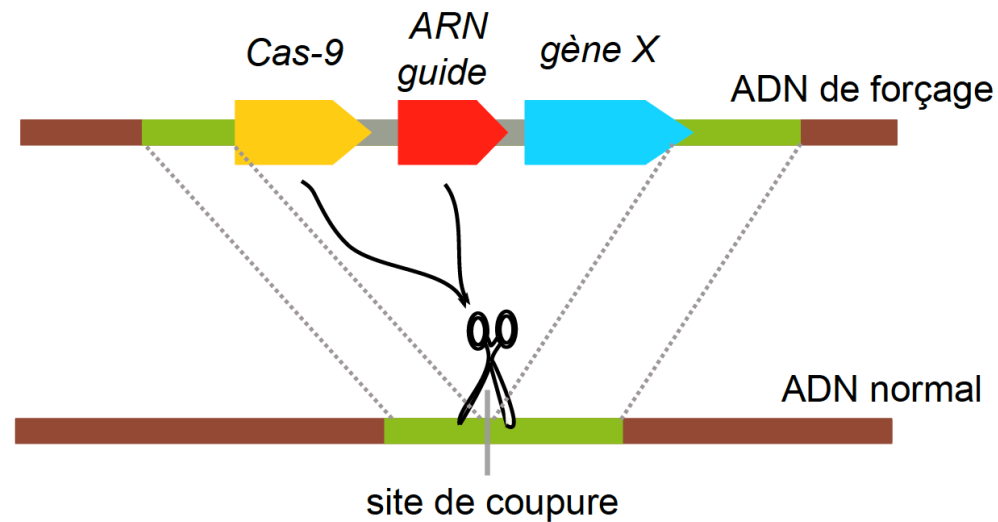
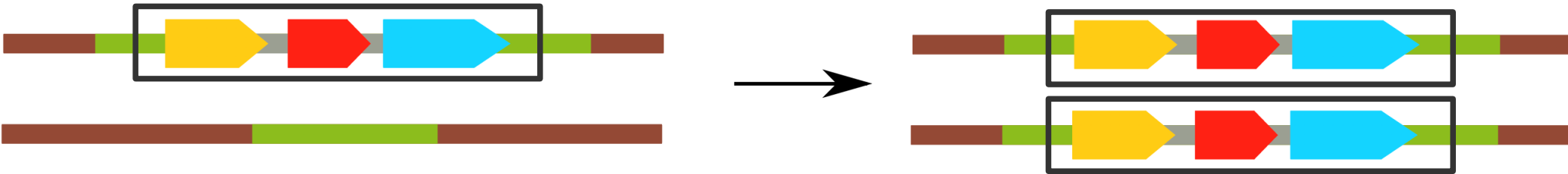


Gene drive





# How a gene drive construct copies itself



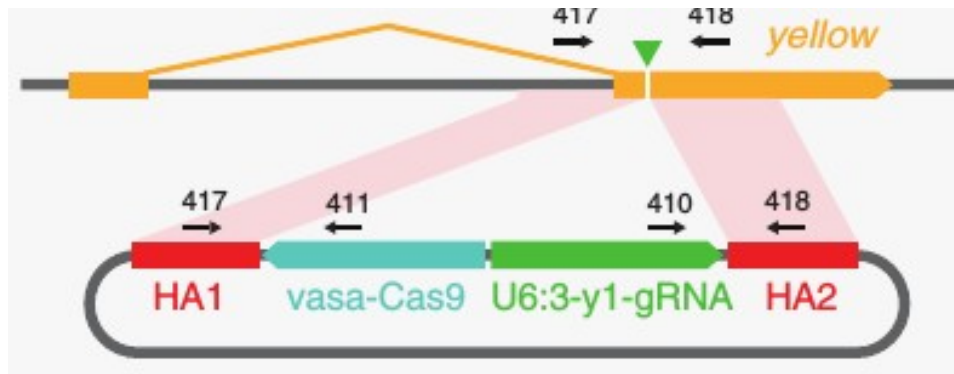
# 1st gene drive organisms

GENOME EDITING

Science, April 2015

## The mutagenic chain reaction: A method for converting heterozygous to homozygous mutations

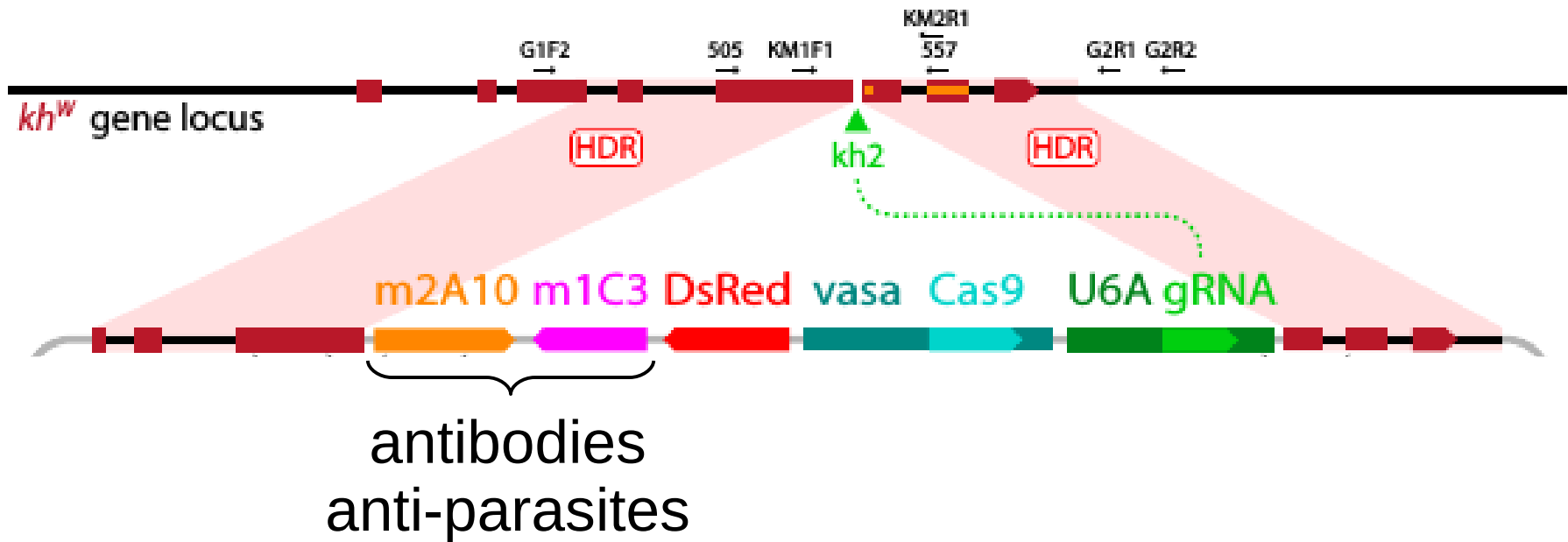
Valentino M. Gantz\* and Ethan Bier\*



**E**

	$y^- \text{♂}$	$y^- \text{♀}$	mosaic ♀	$y^+ \text{♂}$	$y^+ \text{♀}$	total
$y^{\text{MCR}} \text{♂} \times y^+ \text{♀}$	0	40	0	50	1	91
$y^{\text{MCR}} \text{♀} \times y^+ \text{♂}$	214	203	11	2	6	436

# Mosquitoes without parasites

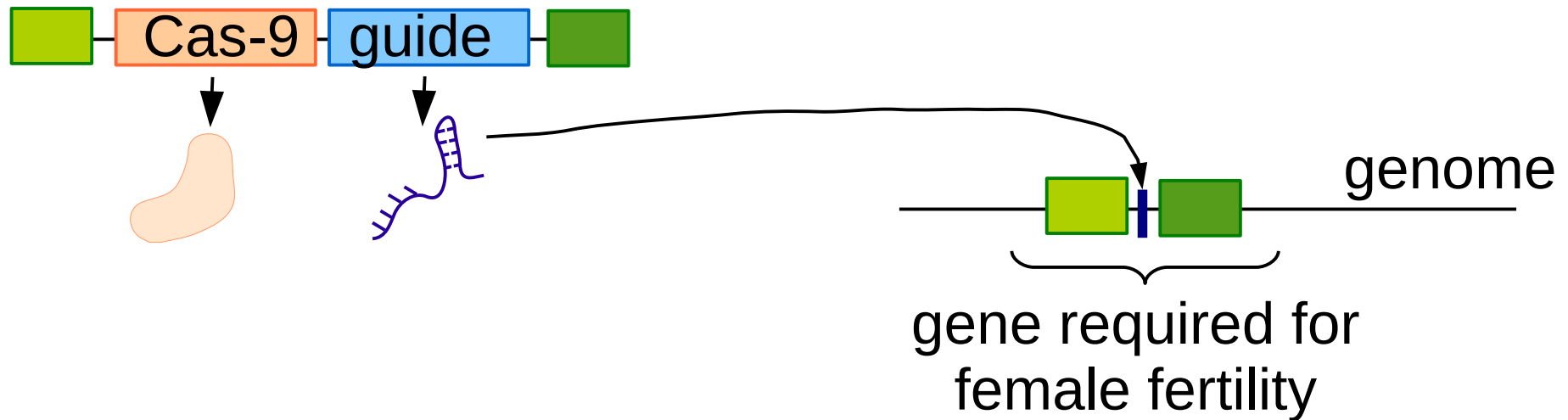


PNAS, November 2015

Highly efficient Cas9-mediated gene drive for population modification of the malaria vector mosquito *Anopheles stephensi*

Valentino M. Gantz<sup>a,1</sup>, Nijole Jasinskiene<sup>b,1</sup>, Olga Tatarenkova<sup>b</sup>, Aniko Fazekas<sup>b</sup>, Vanessa M. Macias<sup>b</sup>, Ethan Bier<sup>a,2</sup>, and Anthony A. James<sup>b,c,2</sup>

# Sterile mosquitoes



Nature Biotechnology, décembre 2015

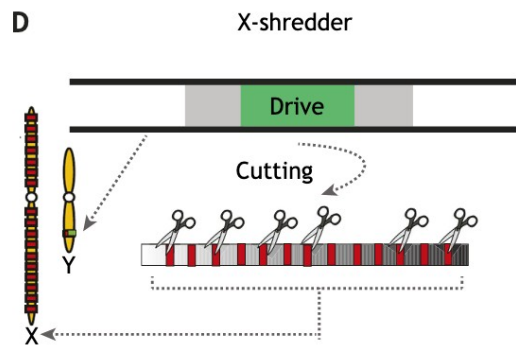
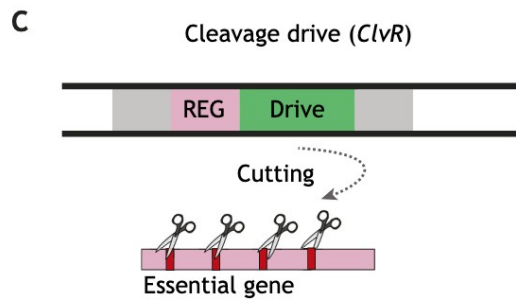
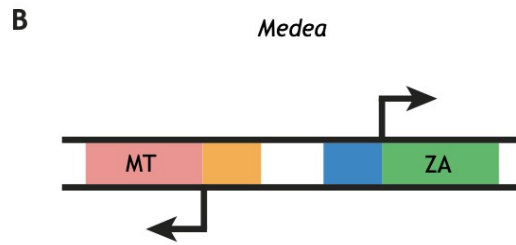
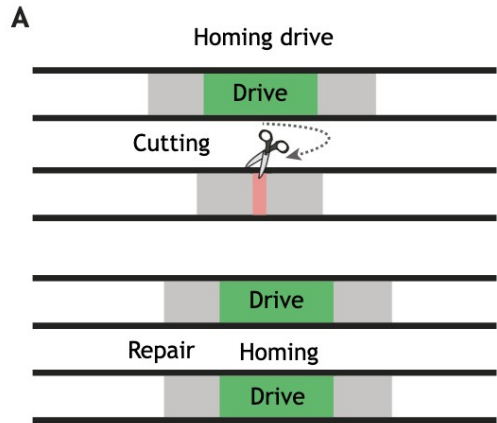
A CRISPR-Cas9 gene drive system targeting female reproduction in the malaria mosquito vector *Anopheles gambiae*

Andrew Hammond<sup>1</sup>, Roberto Galizi<sup>1</sup>, Kyros Kyrou<sup>1</sup>, Alekos Simoni<sup>1</sup>, Carla Siniscalchi<sup>2</sup>, Dimitris Katsanos<sup>1</sup>, Matthew Gribble<sup>1</sup>, Dean Baker<sup>3</sup>, Eric Marois<sup>4</sup>, Steven Russell<sup>3</sup>, Austin Burt<sup>1</sup>, Nikolai Windbichler<sup>1</sup>, Andrea Crisanti<sup>1</sup> & Tony Nolan<sup>1</sup>

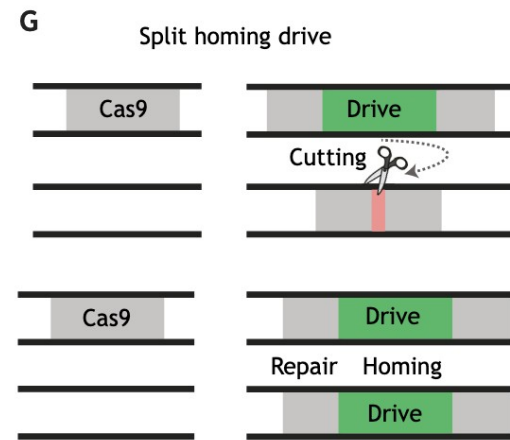
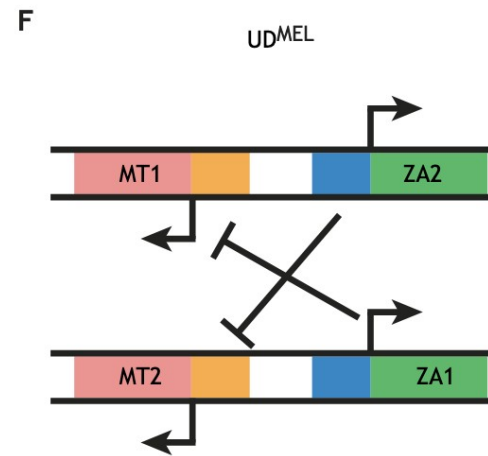
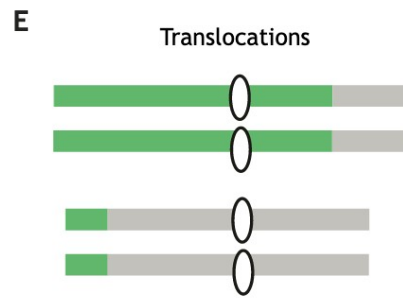
# **Various gene drives**

---

**Non-localized gene**



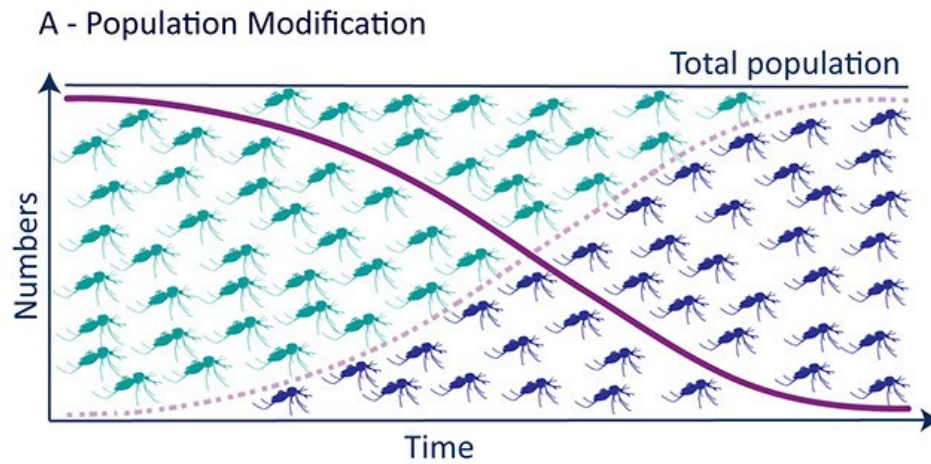
**Localized gene**



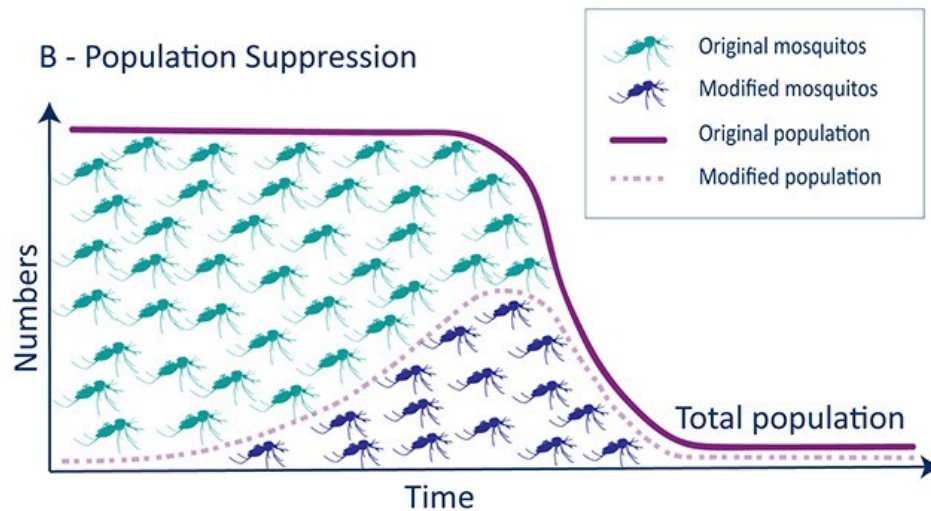
Low threshold drives  
 High threshold drives  
 Integral drives  
 Tethered drives  
 Split drives  
 Daisy drives  
 Under-dominant drives  
 Sex-limited drives

...





*“Outcome conceptually similar to “vaccinating” pest populations”*  
 (Luke Alphey, 2020)



*“Agronomic science has been modifying **crops** to increase productivity or resistance to pests or pathogens.*

*Gene drive now allows manipulating **pests**.”*

# Agricultural pest control with CRISPR-based gene drive: time for public debate

*Should we use gene drive for pest control?*

Virginie Courtier-Orgogozo<sup>1</sup> , Baptiste Morizot<sup>2</sup> & Christophe Boëte<sup>3</sup> 

# Various applications of Gene Drive

## (A) ERADICATION DRIVES

spreading strongly deleterious mutations in invasive populations

↓  
Eradicating invasive pest species



Eradicating invasive black rats that threaten the kereru (New Zealand pigeon) and other endemic species in New Zealand (NASEM 2016)

*Image: David Mudge; Ngā Manu Nature Images*

↓  
Eradicating invasive disease vectors



Eradicating invasive mosquitos, vector of avian malaria in Hawaiian honeycreeper birds (NASEM 2016)

*Image: Sean McCann; Flickr*

## (B) SUPPRESSION DRIVES

spreading mildly deleterious mutations in invasive populations



Reducing the height of invasive common ragweed to decrease its competitive pressure on native plants (Neve 2018)

*Image: Ashley Bradford; inaturalist.org*

## (C) RESCUE DRIVES

spreading beneficial mutations in endangered populations



Protecting lowland leopard frogs from highly pathogenic fungus (Esvelt et al 2014)

*Image: Brian Gratwicke; Flickr*

# Two advanced gene drives

## *Drosophila suzukii*

Invasive pest species



Scott et al. 2018

## *Anopheles* mosquitoes

Vector of malaria



<https://targetmalaria.org>

# **Risks and ethical issues associated with gene drives**

---

# What is novel about gene drive?

Several DNA pieces assembled together  
Eukaryote cis-regulatory regions with bacteria coding regions

## **Manipulates the 2 pillars of evolution**

- mutation
- transmission
- > can bypass selection and spread deleterious alleles

## **Potentially more effective than other biotechnologies**

- ease of use
- speed of change
- unprepared regulatory environment

# Classical Darwinian Evolution

1

Variation



Mutations in DNA

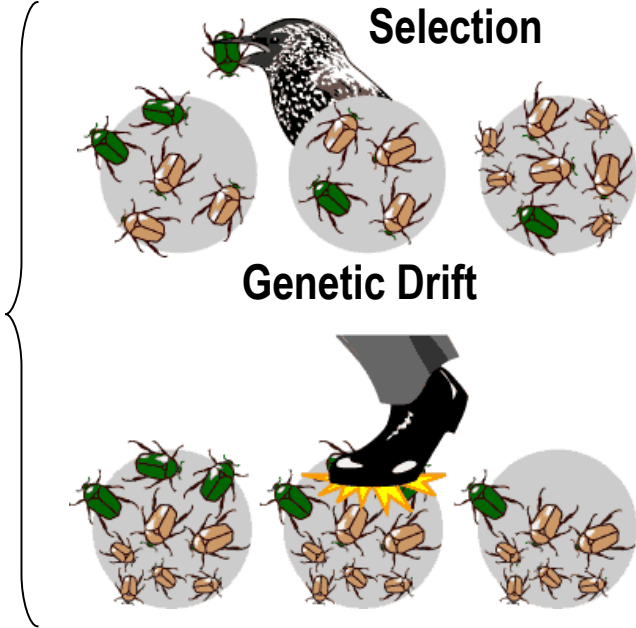
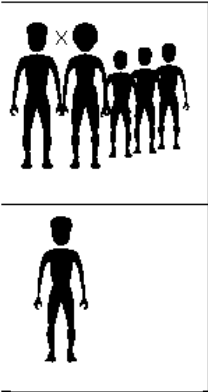
2

Transmission to the next generation



3

Reproduction  
Variability  
between individuals



# **What are the risks?**

**Molecular off-targets**

**Propagation to non-target populations**

**Propagation to non-target species**

**Consequences for ecosystems**

**Failure of counter-measures**



# What are the risks?

**Molecular off-targets**

**Propagation to non-target populations**

**Propagation to non-target species**

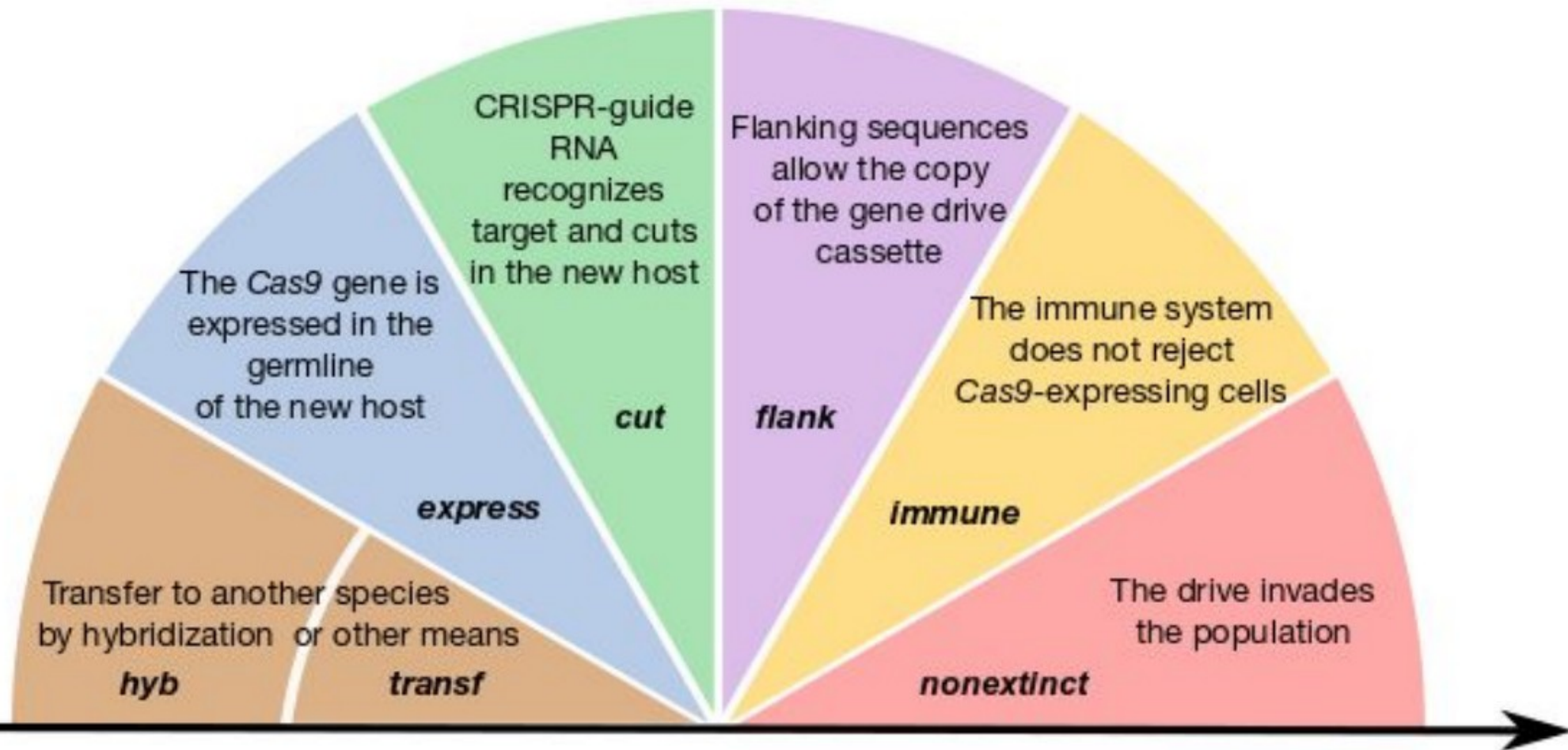
**Consequences for ecosystems**

**Failure of counter-measures**

# Evaluating the probability of CRISPR-based gene drive contaminating another species

2019

Virginie Courtier-Orgogozo<sup>1</sup>  | Antoine Danchin<sup>2</sup>  | Pierre-Henri Gouyon<sup>3</sup>  |  
Christophe Boëte<sup>4</sup> 



# Risk of hybridization

## *Drosophila suzukii*

Invasive pest species



*D. subpulchrella* India, South East Asia, China, Japan

*D. pulchrella* India, South East Asia, southern China

temperate  
tropical

## *Anopheles* mosquitoes

Vector of malaria



*An. gambiae* s.s.

*An. arabiensis*

*An. coluzzii*

*An. amharicus*

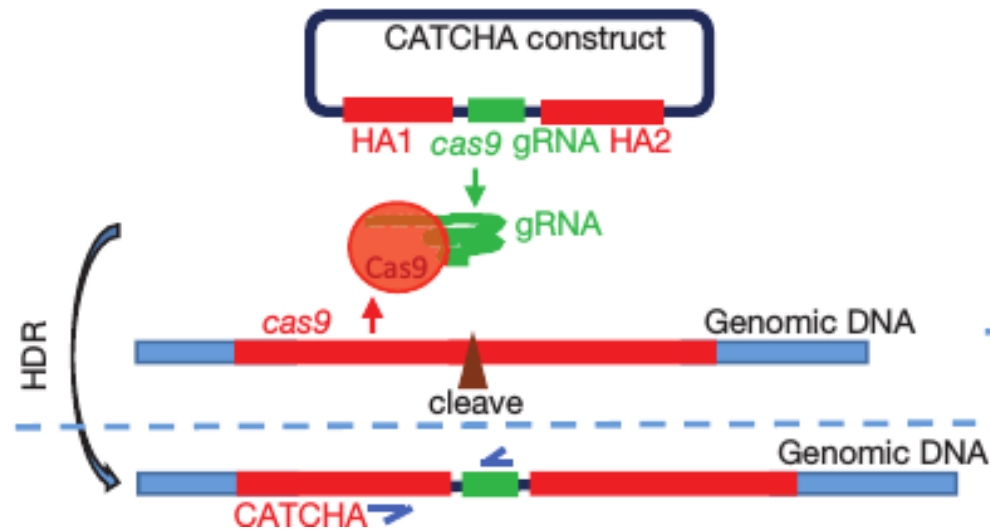
*An. melas*

*An. merus*

*An. bwambae*

*An. quadriannulatus*

# Need to stop a drive? Use another one!



CORRESPONDENCE

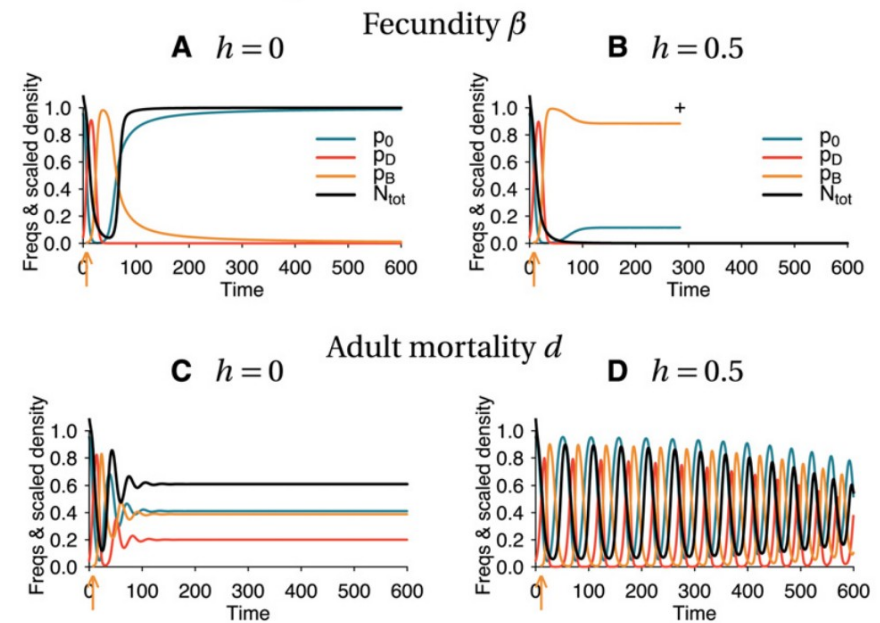
Nature Biotechnologies, Feb 2016

Bing Wu<sup>1,2</sup>, Liqun Luo<sup>1</sup> & Xiaojing J Gao<sup>1-3</sup>

**Cas9-triggered chain ablation of *cas9* as a gene drive brake**



# A brake is not guaranteed to stop an eradication drive



# Gene drives: good or bad?

May eradicate  
diseases and pest species

Less expensive than other  
methods

Potentially faster than other  
methods

Potentially more powerful  
than other methods

Potentially less efficient than  
expected (resistance via  
mutations in the target site,  
cryptic species)

An uncontrolled system  
released in the wild

Impact on other species and  
ecosystems not quantified

# Gene drives



Biases:

Living  
in malaria area

Developing  
gene drives

etc.



# Regulation

Falls under the GMO regulation

Cartagena Protocol: international agreement, established as a supplement to the Convention on Biological Diversity (CBD), which aims to protect biological diversity from the potential risks imposed by LMOs (*Living Modified Organisms=any living organism that possesses a novel combination of genetic material obtained through the use of modern biotechnology*)

Researchers added extra safety rules in their laboratories

What to do if it goes wrong?

International regulation?



# LE FORÇAGE GÉNÉTIQUE (GENE DRIVE) ET SES APPLICATIONS



@Biol4Ever

*GENE DRIVE AND ITS APPLICATIONS*

*Par Virginie COURTIER-ORGOGOZO<sup>(1)</sup>*

<http://documents.irevues.inist.fr/handle/2042/70673>

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