

Computational modeling of protein-ligand interactions

O. Taboureau, AC. Camproux, P. Tuffery

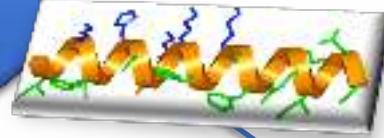
UMR-S 973

2014-2018

UMR-S 973
(Dir : B. Villoutreix)

Team 1
P. Tuffery

**Peptide
design**



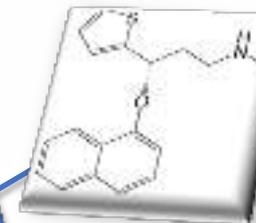
Team 2
AC. Camproux
O. Taboureaux

Profiling



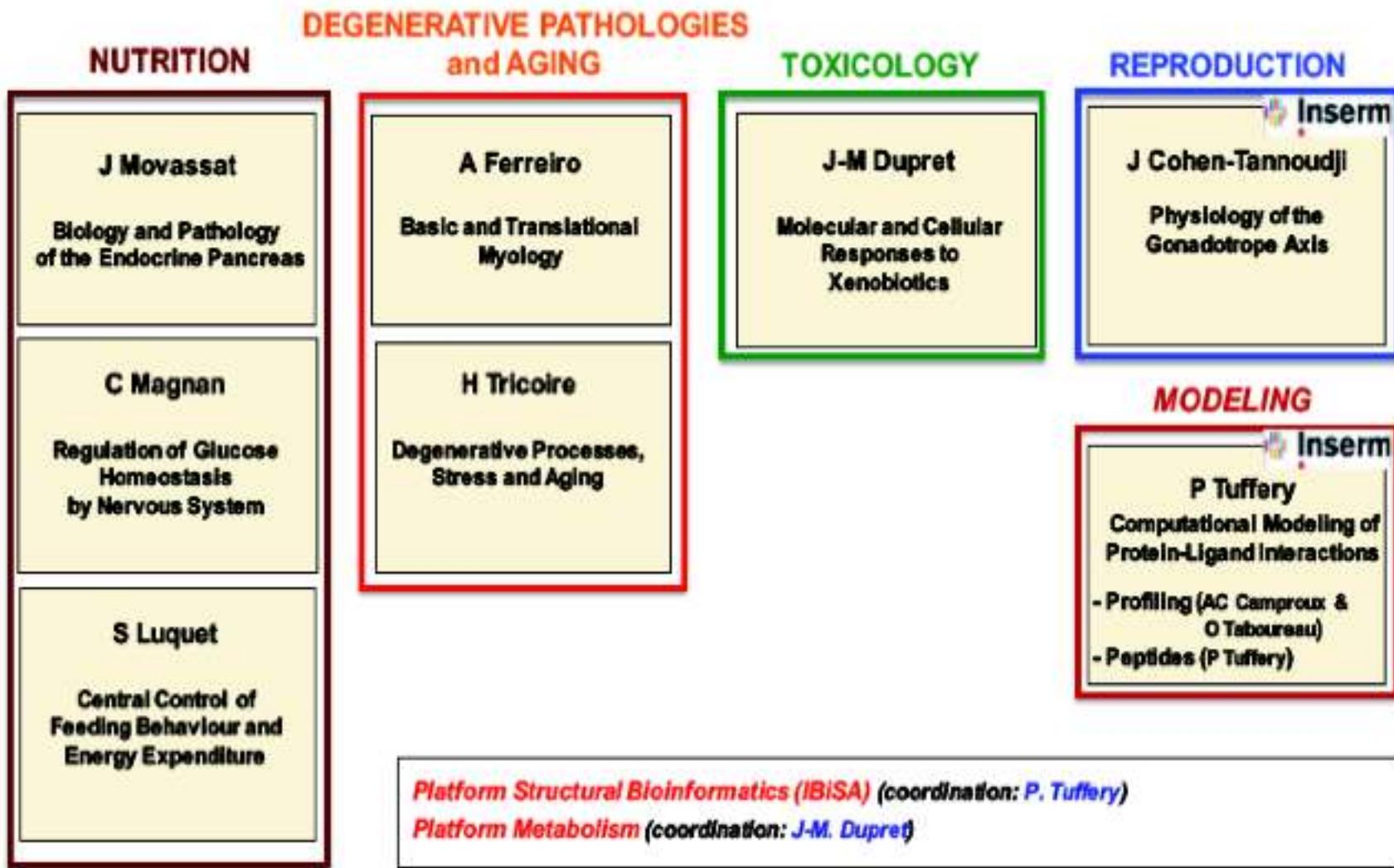
Team 3
M. Miteva
B. Villoutreix

**Virtual
screening
& ADMET**



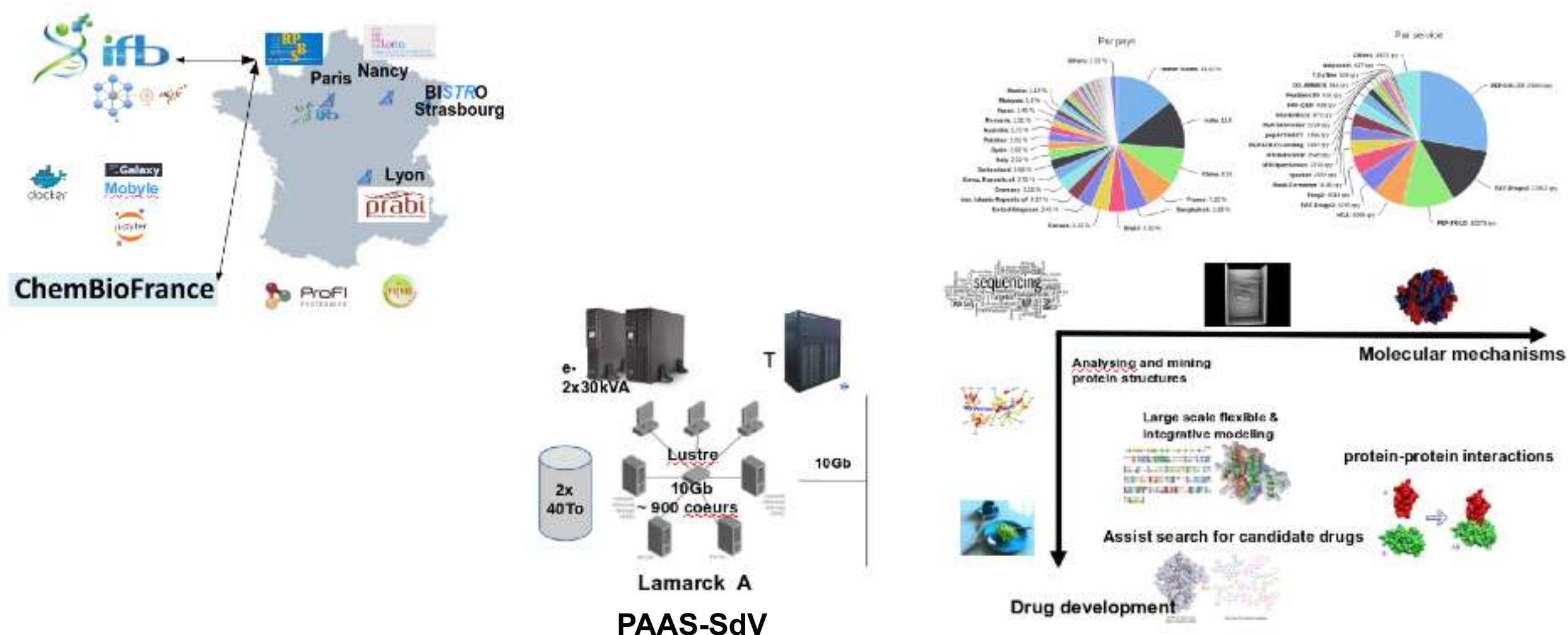
Computational modeling of protein-ligand interactions

2019-2023



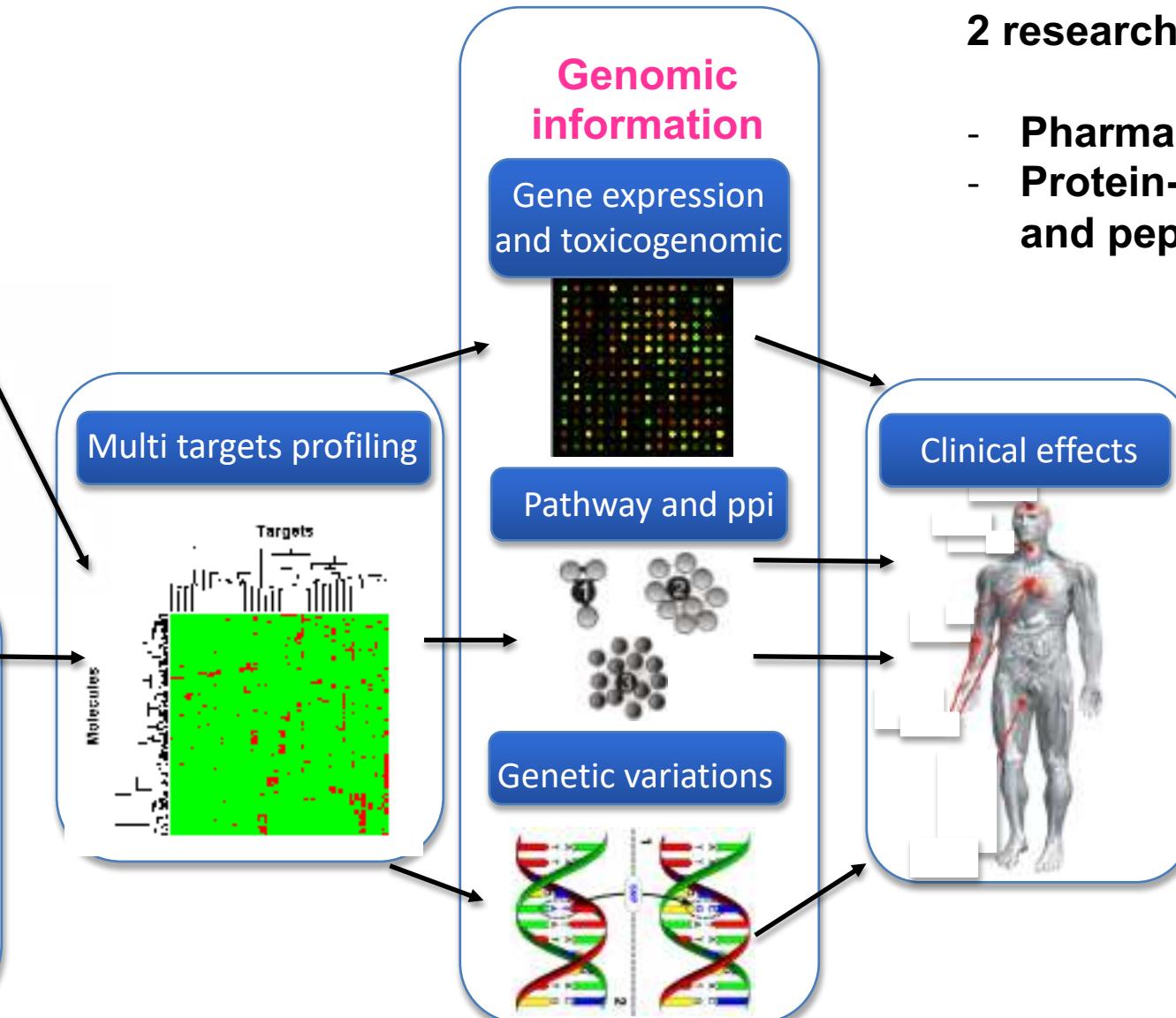
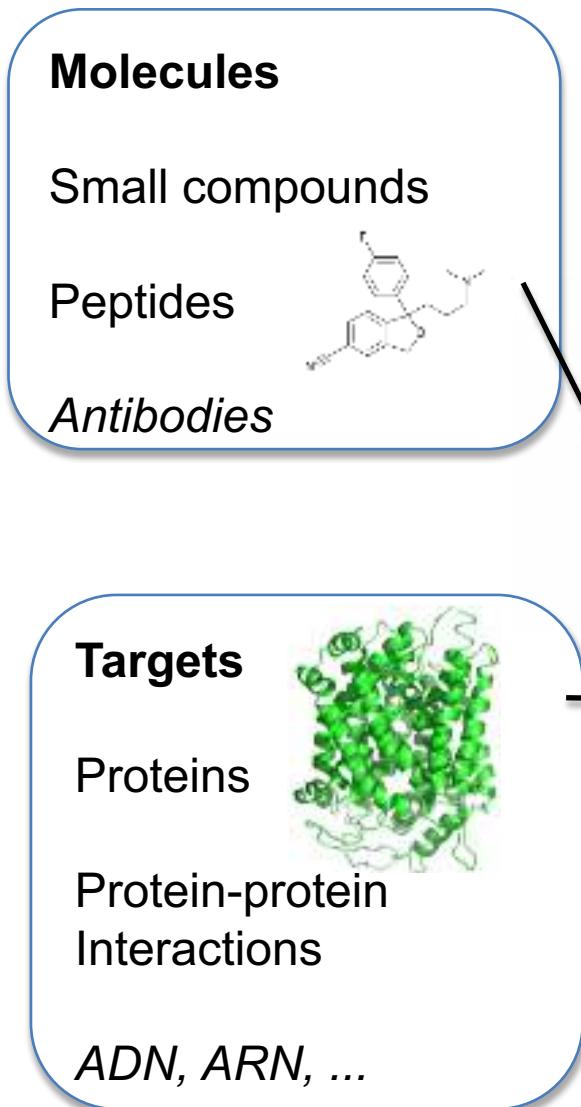
Computational modeling of protein-ligand interactions

Plate-forme RPBS (*IBiSA*) (P. Tufféry, S. De Vries, J. Rey)
(Ressource Parisienne en Bioinformatique Structurale)



Computational modeling of protein-ligand interactions

Objectives

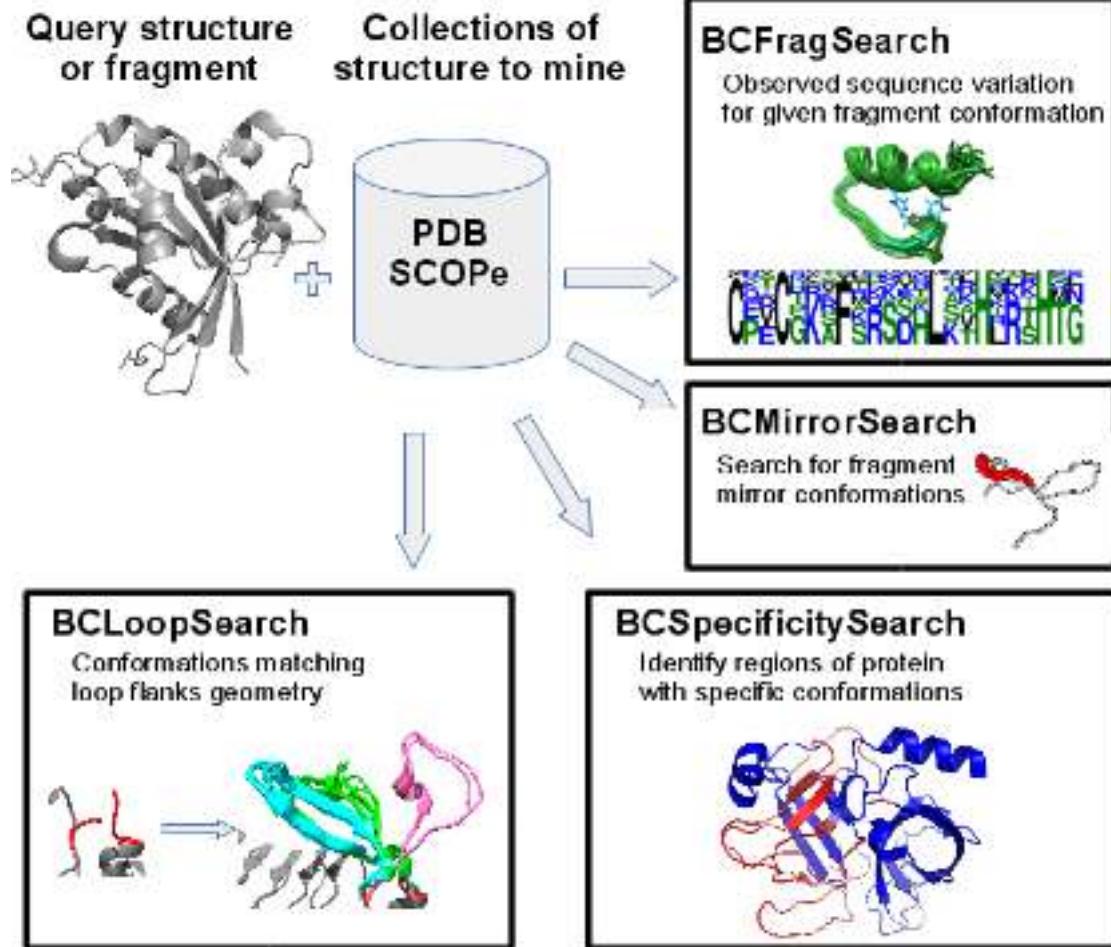


2 research areas :

- Pharmacological profiling
- Protein-peptide interactions and peptides design

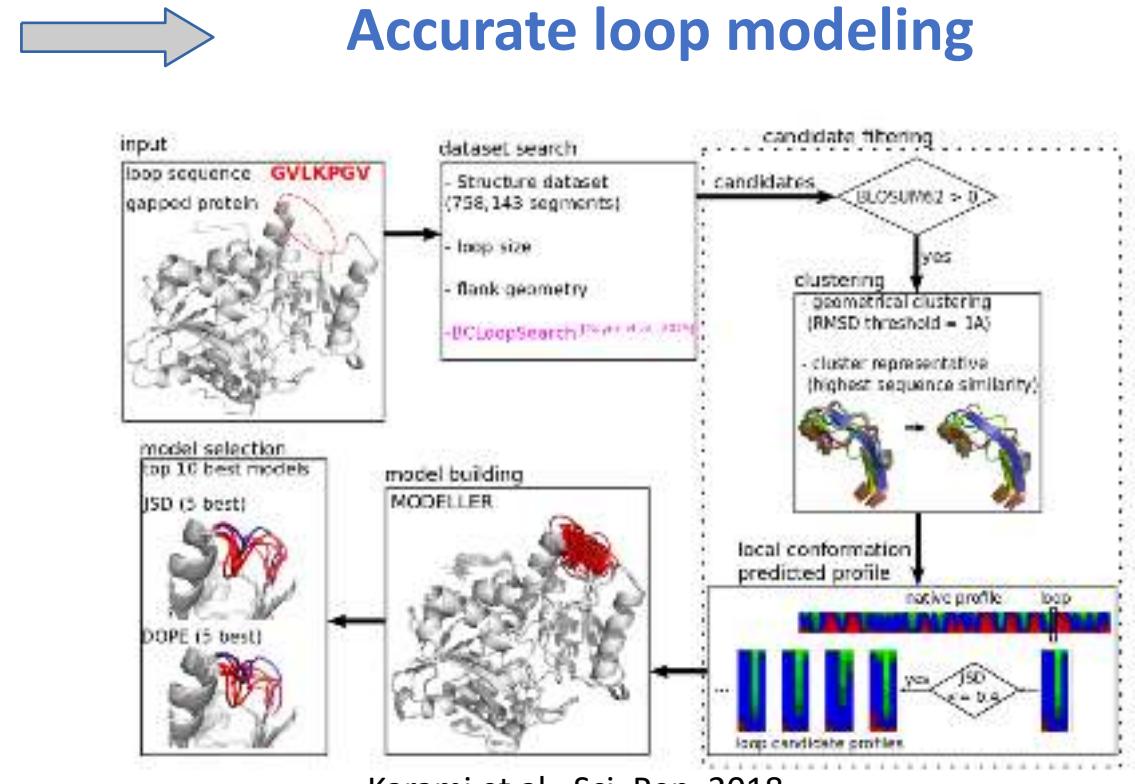
Peptide design: Recognition

Fast geometric search in protein structures



Input: gapped amino acid sequence

Output: 3D fragments matching flanks & size 3D



Karami et al., Sci. Rep. 2018

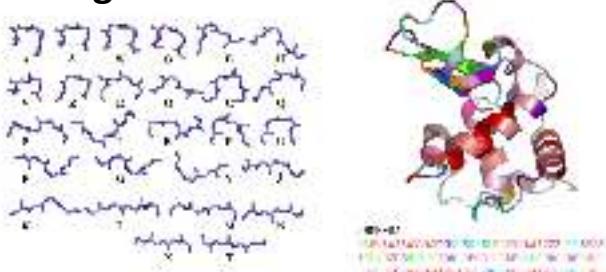
Over 50% of successful loop models are derived from unrelated proteins, indicating that fragments under similar constraints tend to adopt similar structure, beyond mere homology.

Peptide design: Modeling

Peptide structure modeling

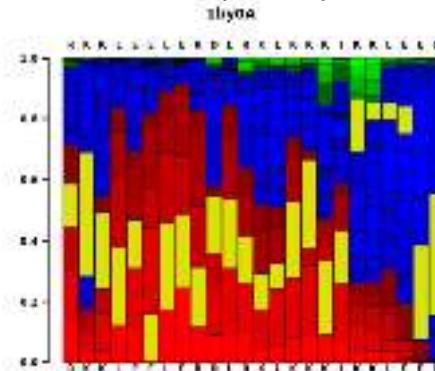
3D encoding of structures

Using Hidden Markov Models



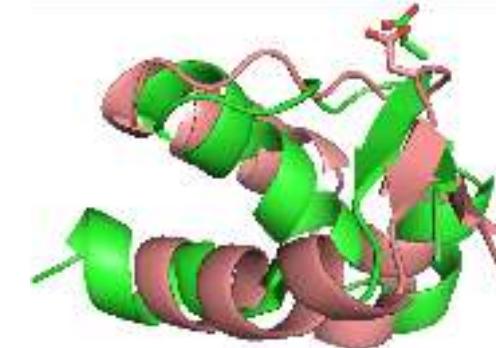
Camproux et al. J. Mol. Biol., 2004

Forward-backtrack, K-best, Taboo sampling



Maupetit et al., NAR, 2009

Thevenet et al., NAR, 2012



Shen et al., J. Chem. Theor. Comput., 2014
Lamiable et al. NAR, 2016

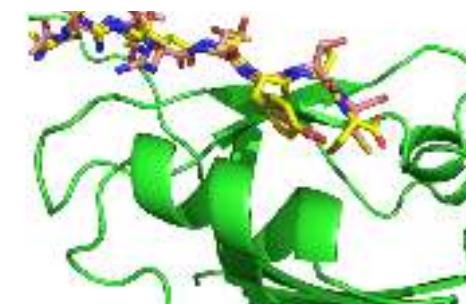
Protein-protein interactions

Protein binding site identification



Saladin et al, Nucleic Acids Res., 2014

Folding peptide at protein binding site



Lamiable et al, J. Comput Chem., 2016

<http://bioserv.rpbs.univ-paris-diderot.fr/services/PEP-SiteFinder/>

<http://bioserv.rpbs.univ-paris-diderot.fr/services/PEP-FOLD/>

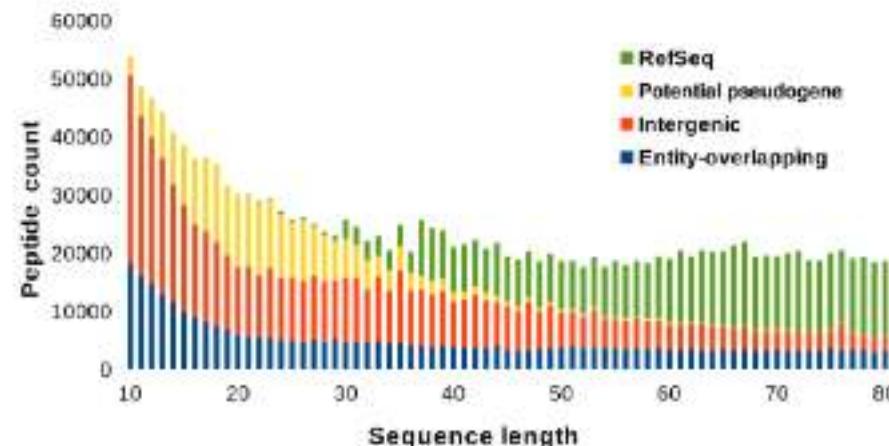
<http://bioserv.rpbs.univ-paris-diderot.fr/services/PEP-FOLD3/>

Peptide discovery: BactPepDB: a database of predicted peptides in prokaryotic genomes



Last production update (15/10/2017) :

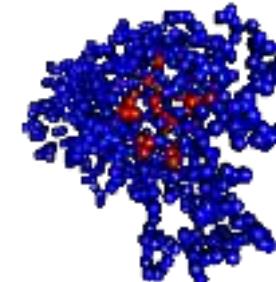
- Prokaryotes genomes :
 - ~ 100 orders
 - ~ 200 families
 - ~ 700 genera
 - ~ 1,500 species
 - ~ 2,700 strains
- Total : ~ 2,000,000 peptides
 - ~ 70 % of newcomers
 - ~ 200,000 (~ 20 %) of new intergenic SCSs are conserved to some extent : consistent with genes found in RefSeq



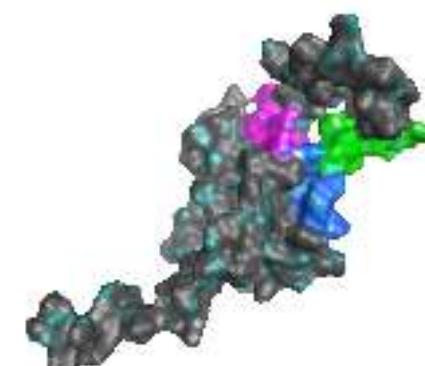
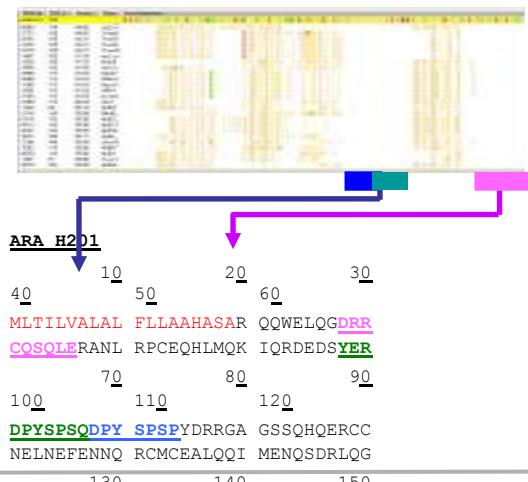
Peptide design, off-target: Non sequential alignments

Comparison of atom positions independently of the amino-acid sequence order

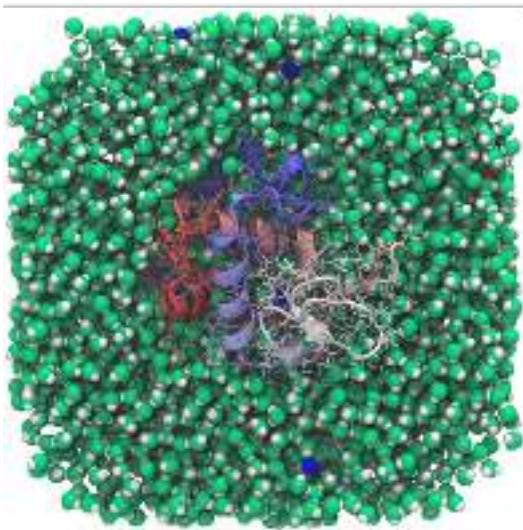
→ Far more difficult problem but useful for protein surface comparisons. atoms involved in a function, : interaction with a drug, interaction with other proteins



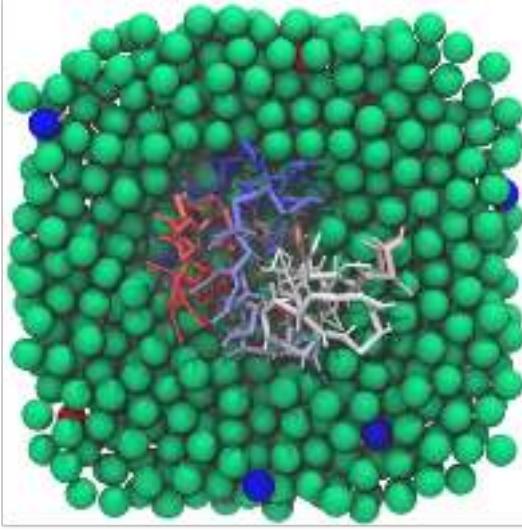
- We use graph theory : search for cliques or quasi-cliques in product graphs
- We developed a similarity measure already used in image analysis for face or object recognition
(Binet-Cauchy Kernel)



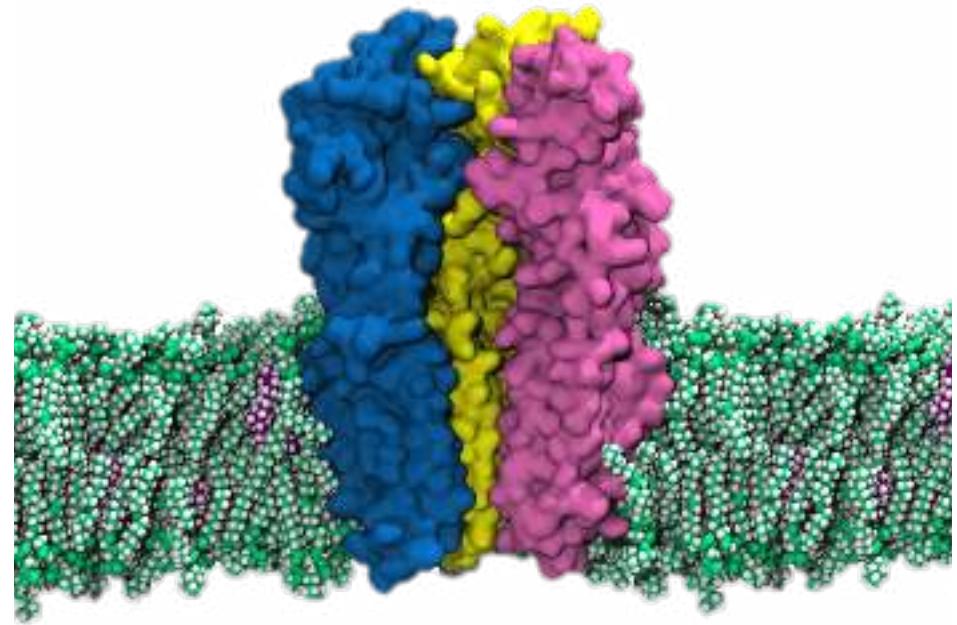
Membrane proteins motions



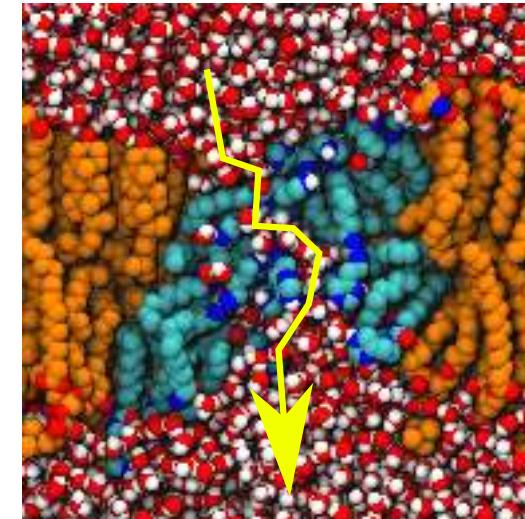
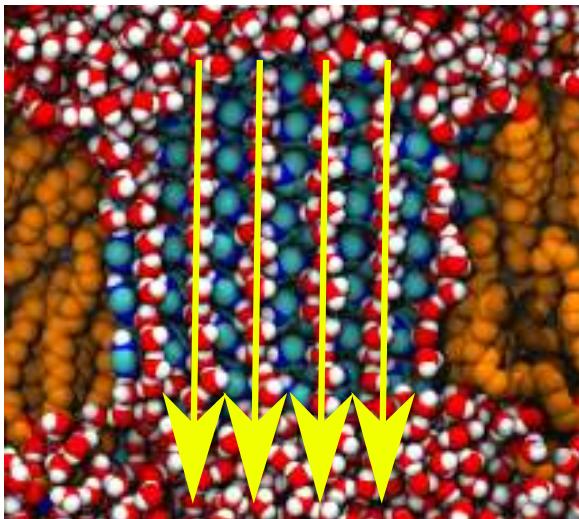
Molecular Dynamic
Simulations



Coarses grain



Synthetic channels



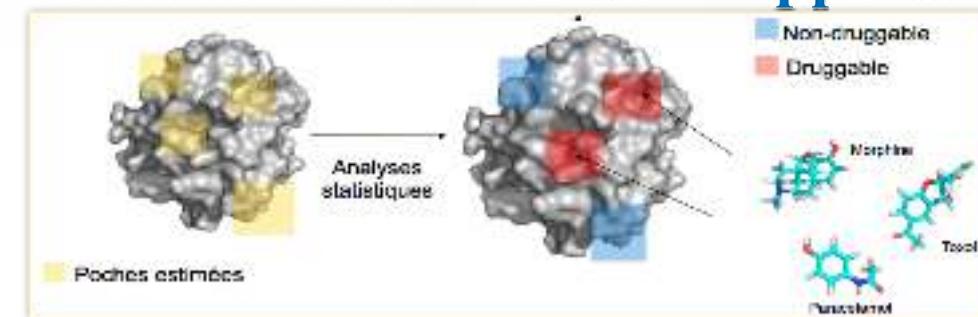
Pharmacological profiling: Target based

Characterization and validation of proteins as therapeutic targets and prediction of drug-target interactions based on statistical & bioinformatics structural approaches

Prediction of druggable targets:

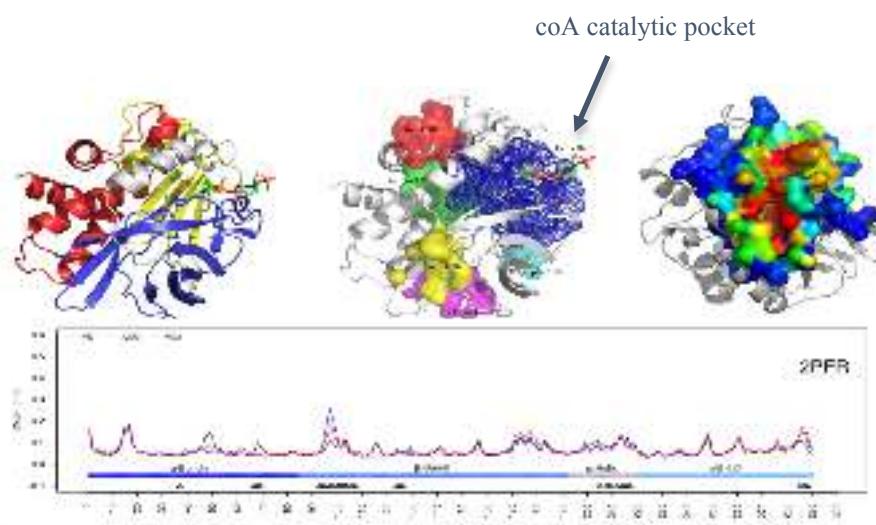
Proteins able to bind drug-like molecules, using statistical approaches: Pockdrug website

Borrel et al, 2015, Hussein et al, 2015



Proteins flexibility and transient pocket analysis

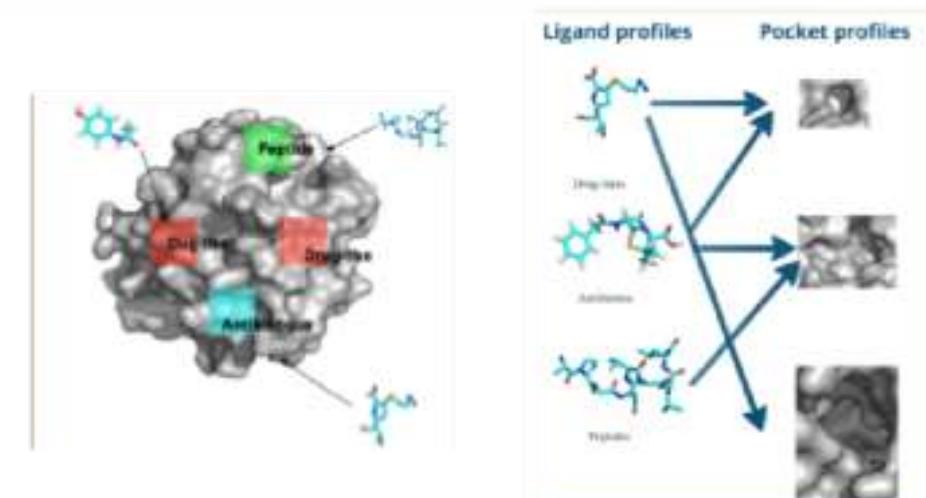
Pockdrug extension using dynamic molecular simulations:



Current applications: NS1 (non structural protein of influenza virus)

Prediction of ligand profiles using biostatistical approaches

Further deepen the profiling study: propose ligand candidates that can bind with high affinity druggable pockets



Hussein et al, 2017, Regad et al, 2017, Alam et al, 2018

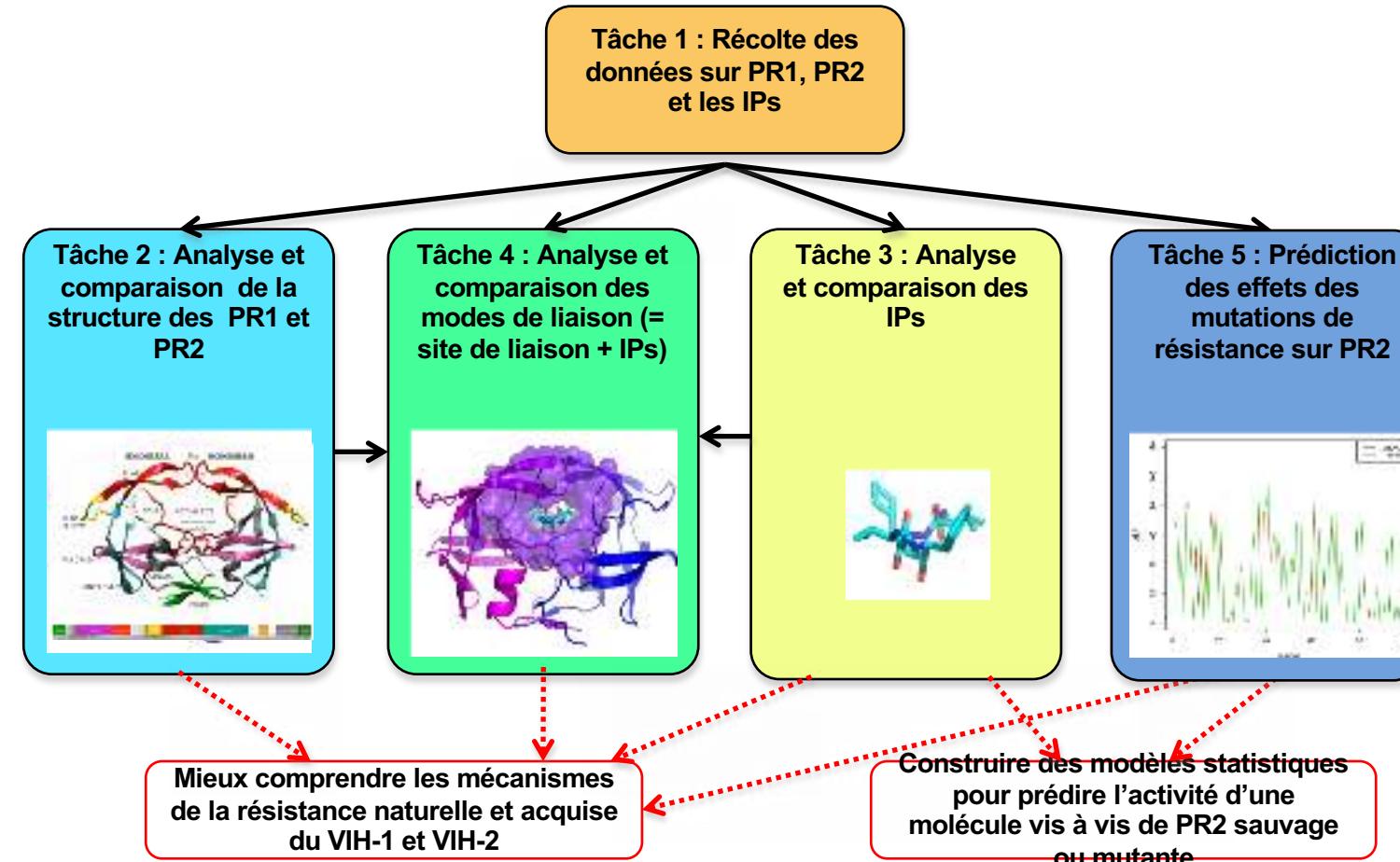
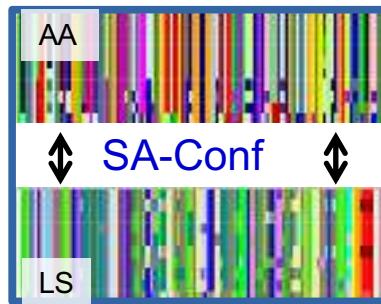
Cerisier et al, 2017a, Cerisier et al, 2017b

Pharmacological profiling: Target based – Application on HIV-2 proteases

Inhibition of HIV-2 protease: study of the resistance mechanism using in silico approaches

3D structures : 15 PR1
13 PR2

unbound, bound forms
(indinavir, darunavir, amprenavir)



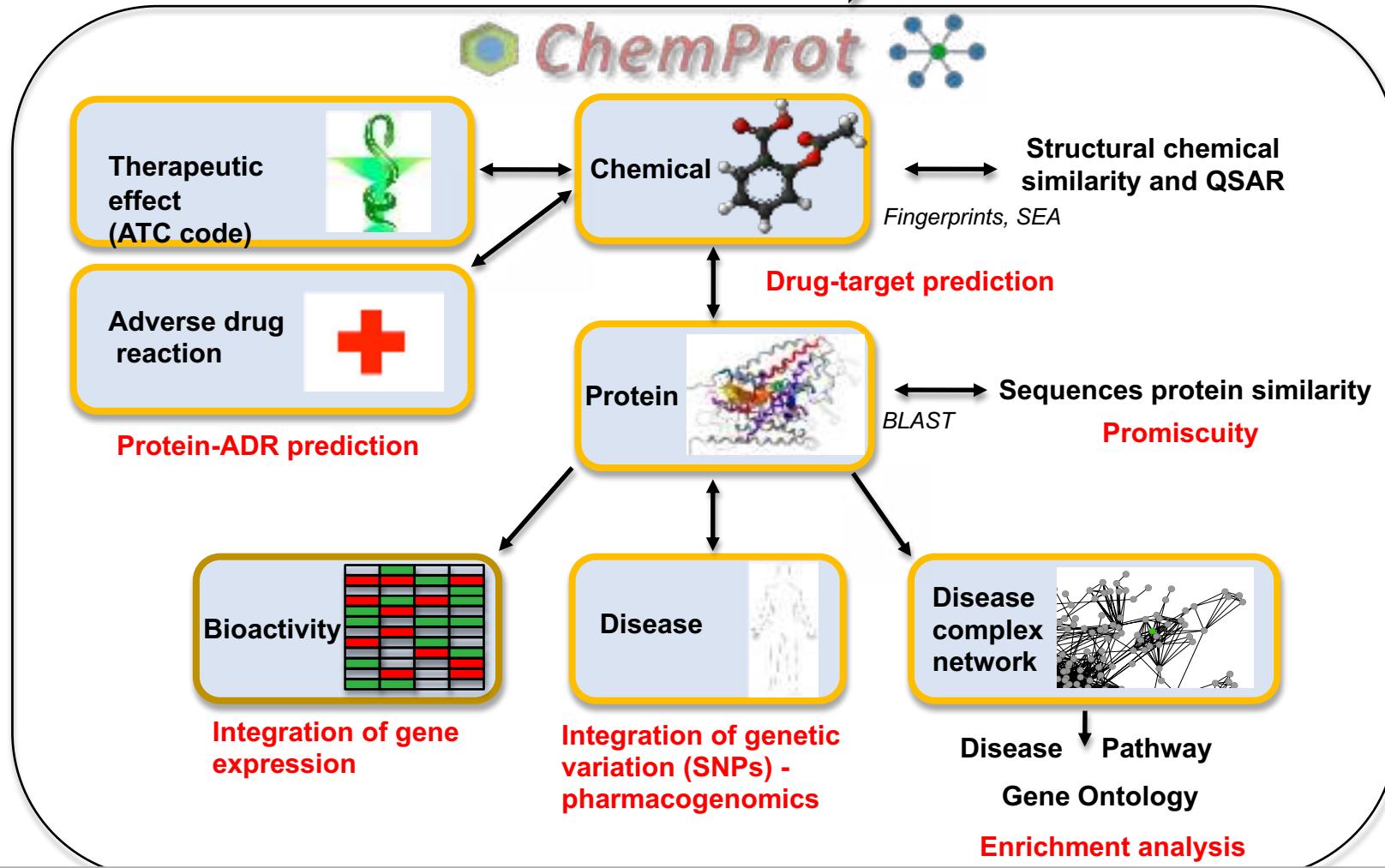
Projet VIH2-Bioinfo



Pharmacological profiling: Data integration

ChemProt: focused on Drug-Target-Biological outcomes profiling

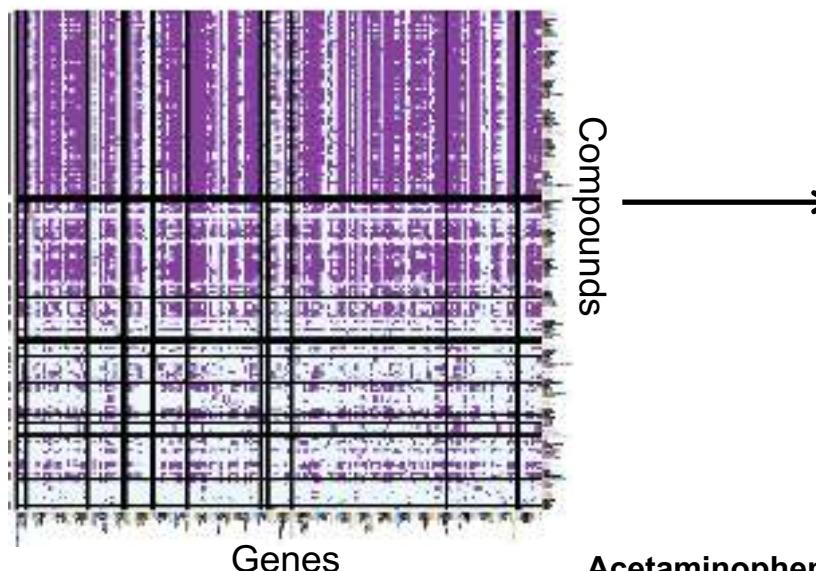
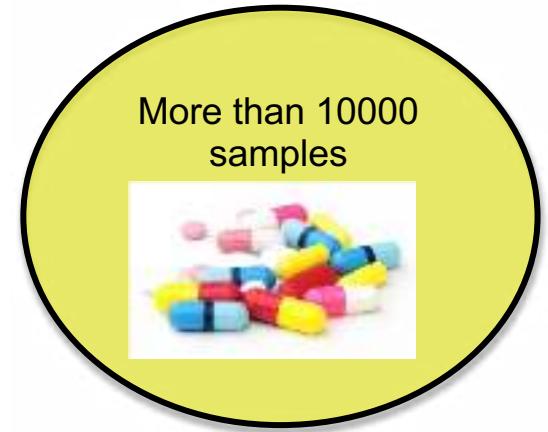
Many data not always linked together → Need of data integration



Pharmacological profiling: Systems chemical toxicology

Analysis of large scale microarray data

Toxicogenomics data analysis



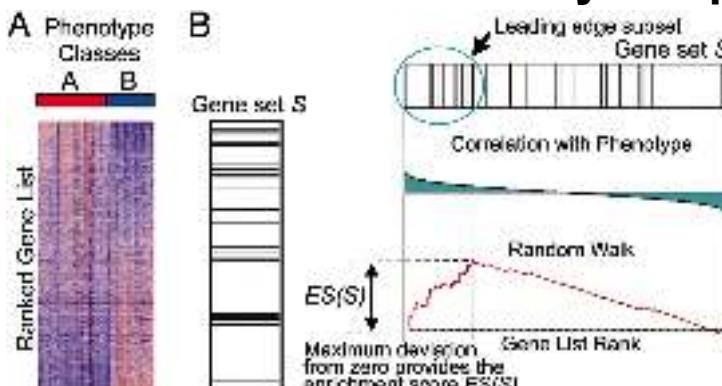
Gene	pvalNom(invivo)	pvalNom(invitro)
Acot1	4.45e-04	6.74e-08
Aig1	2.82e-04	2.14e-06
Ehhadh	4.43e-05	3.18e-06
Acox1	5.31e-04	4.97e-06
Cpt1b	1.53e-03	5.95e-06
Cpt2	2.09e-03	6.82e-06
Slc27a2	6.04e-04	1.04e-05

WY-14643

Lipid metabolism

Fatty acid transporter

Gene Set Enrichment Analysis approach (GSEA)



Rank	Term	ES	Neg. Z-Score
1	Basal Notch signaling	2.23e-05	-5.20e-05
2	Basal Notch signaling	2.02e-05	-2.02e-05
3	Basal Notch signaling	1.83e-05	-2.02e-05
4	Basal Notch signaling	1.71e-05	-3.19e-05
5	Basal Notch signaling	1.57e-05	-2.00e-05
6	Basal Notch signaling	1.43e-05	-4.24e-05
7	Basal Notch signaling	1.30e-05	-3.02e-05
8	Basal Notch signaling	1.17e-05	-1.07e-05
9	Basal Notch signaling	1.04e-05	-1.07e-05
10	Basal Notch signaling	9.13e-06	-1.07e-05
11	Basal Notch signaling	8.80e-06	-1.07e-05
12	Basal Notch signaling	8.47e-06	-1.07e-05
13	Basal Notch signaling	8.14e-06	-1.07e-05
14	Basal Notch signaling	7.81e-06	-1.07e-05
15	Basal Notch signaling	7.48e-06	-1.07e-05
16	Basal Notch signaling	7.15e-06	-1.07e-05
17	Basal Notch signaling	6.82e-06	-1.07e-05
18	Basal Notch signaling	6.49e-06	-1.07e-05
19	Basal Notch signaling	6.16e-06	-1.07e-05
20	Basal Notch signaling	5.83e-06	-1.07e-05
21	Basal Notch signaling	5.50e-06	-1.07e-05
22	Basal Notch signaling	5.17e-06	-1.07e-05
23	Basal Notch signaling	4.84e-06	-1.07e-05
24	Basal Notch signaling	4.51e-06	-1.07e-05
25	Basal Notch signaling	4.18e-06	-1.07e-05
26	Basal Notch signaling	3.85e-06	-1.07e-05
27	Basal Notch signaling	3.52e-06	-1.07e-05
28	Basal Notch signaling	3.19e-06	-1.07e-05
29	Basal Notch signaling	2.86e-06	-1.07e-05
30	Basal Notch signaling	2.53e-06	-1.07e-05
31	Basal Notch signaling	2.20e-06	-1.07e-05
32	Basal Notch signaling	1.87e-06	-1.07e-05
33	Basal Notch signaling	1.54e-06	-1.07e-05
34	Basal Notch signaling	1.21e-06	-1.07e-05
35	Basal Notch signaling	8.80e-07	-1.07e-05
36	Basal Notch signaling	5.49e-07	-1.07e-05
37	Basal Notch signaling	2.16e-07	-1.07e-05
38	Basal Notch signaling	1.83e-07	-1.07e-05
39	Basal Notch signaling	1.50e-07	-1.07e-05
40	Basal Notch signaling	1.17e-07	-1.07e-05
41	Basal Notch signaling	8.47e-08	-1.07e-05
42	Basal Notch signaling	5.16e-08	-1.07e-05
43	Basal Notch signaling	1.85e-08	-1.07e-05
44	Basal Notch signaling	1.52e-08	-1.07e-05
45	Basal Notch signaling	1.19e-08	-1.07e-05
46	Basal Notch signaling	8.68e-09	-1.07e-05
47	Basal Notch signaling	5.37e-09	-1.07e-05
48	Basal Notch signaling	2.06e-09	-1.07e-05
49	Basal Notch signaling	1.73e-09	-1.07e-05
50	Basal Notch signaling	1.40e-09	-1.07e-05

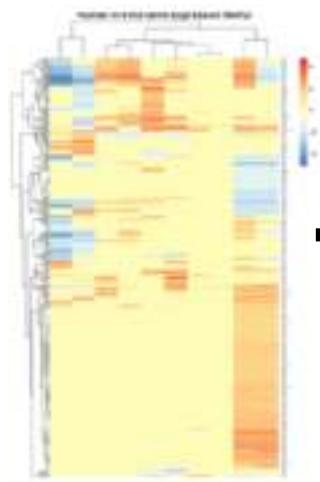
Pharmacological profiling: Example with Steatosis and DILI

Prediction of DILI compounds and genes associated to steatosis

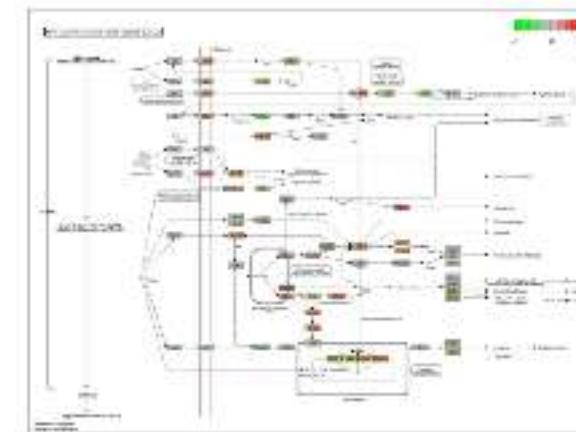
Ligand-protein interaction



Cell deregulation



Metabolic pathways affected



- Suggestion of gene's biomarkers (ALDH7A1, OSBL9 ...)
- Suggestion of molecules with a risk.

+

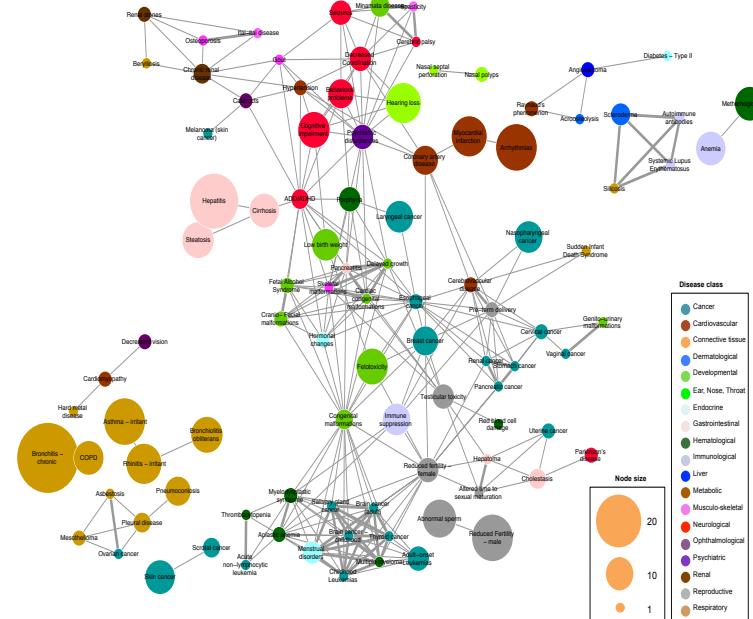
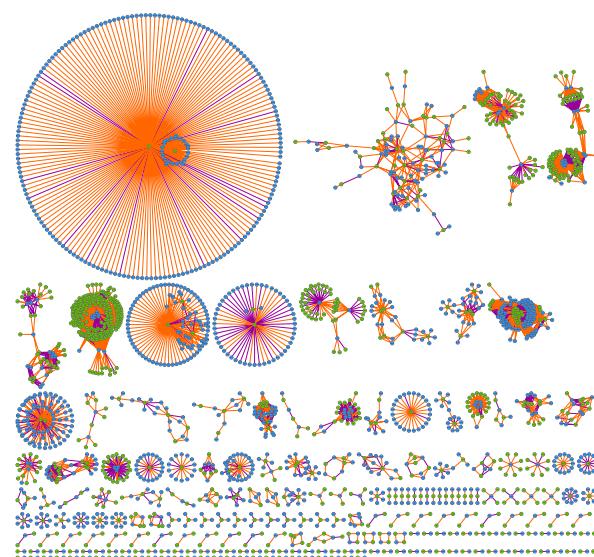
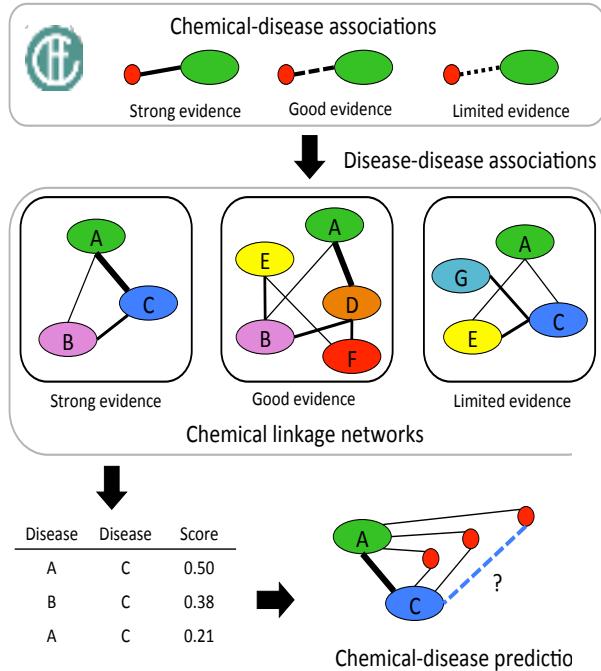
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Such analysis can be used for computational phenotypic screen.

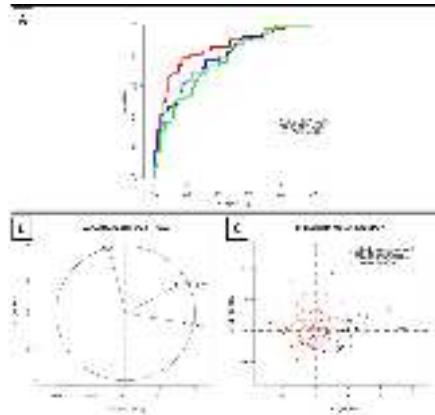
Pharmacological profiling: Network-based Analysis

Development of network-based analysis tools to predict chemical-chemical interactions to diseases and other biological outcomes

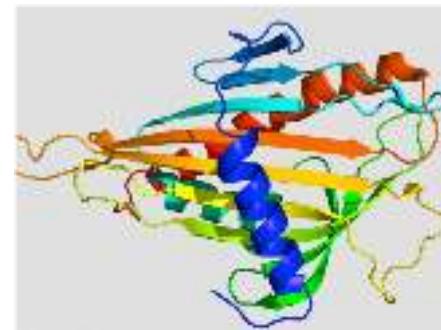


Current collaborations with teams at P7

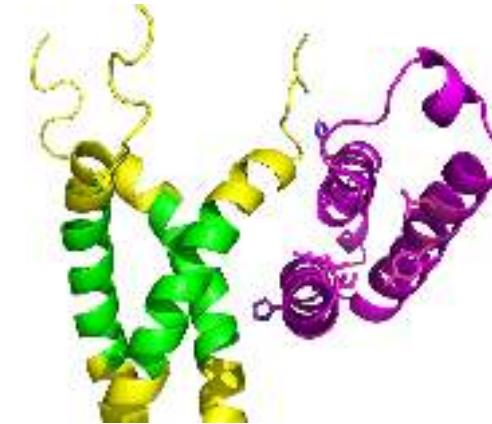
Alzheimer's disease,
DYRK1A, CBS (N. Janel)



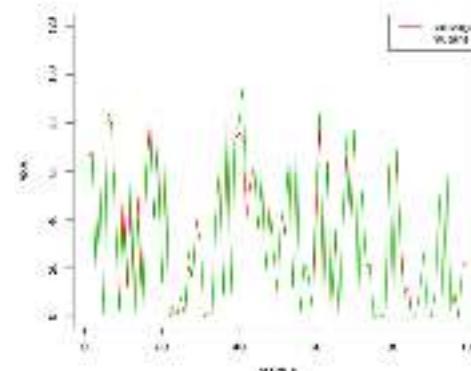
Docking study into the CERT
transporter (C. Magnan)



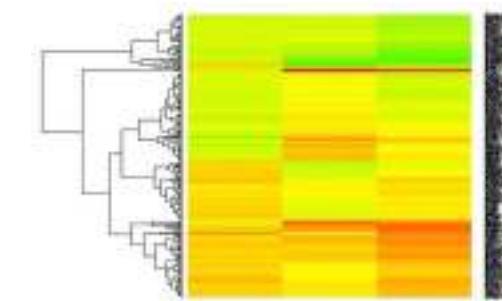
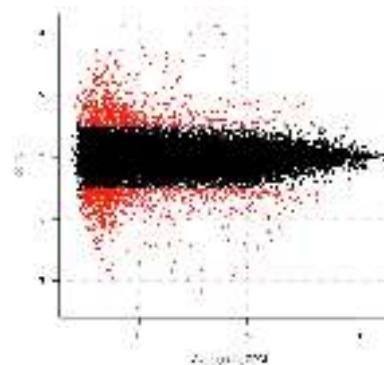
Docking study: HSF2 (A. de
Thonel and V. Lallemand-Mezger)



NAT exploration with MD
simulation (F. Rodrigues-Lima)



RNAseq study on diabetes. Gut Microbiota peptides
(C. Magnan)



Team members

Permanent staff

PR. AC. Camproux

PR. O. Taboureau

CR. M. Petitjean

MCF. A. Badel

MCF. D. Flatters

MCF. L. Regad

Tech. C. Geneix

Post doc and Ph.D in 2018

ATER. V. Leroux

Ph.D N.. Cerisier

PhD. B. Boezio

PhD. P. Laville



Permanent staff

DR. P. Tuffery

MCF. G. Moroy

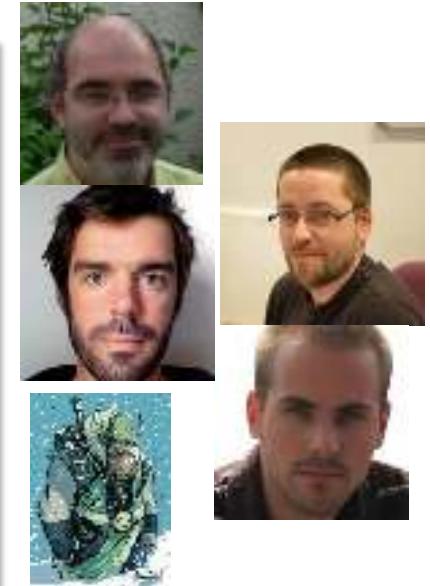
MCF. S. Murail

IG. J. Rey

IG. S. De Vries

Post doc and Ph.D in 2018

Post doc: G. Postic



Thank you